

## Neuro-Protection after Traumatic Brain Injury: Novel Strategies.

Alok Singh<sup>1</sup>, Parag Sharma<sup>2</sup>, Suryaprakash Mishra<sup>3</sup>

<sup>1</sup> Department Of Pharmacology, G. R. Medical College, Gwalior, 474001 Madhya Pradesh India.

<sup>2</sup> Department Of Pharmacology, Peoples College Of Medical Sciences, Bhopal Madhya Pradesh 462037 India..

<sup>3</sup> Department Of Pharmacology, Institute Of Medical Sciences, Banaras Hindu University Varanasi 221005 Uttar Pradesh India

---

**Abstract:** Traumatic brain injuries (TBI) are increasing cause of functional disability which requires aggressive researches on neuroprotective agents to prevent or rectify the consequences of organic brain damage and to enhance rehabilitation. Many of the drugs which proves beneficial in preclinical studies failed to translate into clinical practice, hence the search for the ideal neuroprotective drug continues. Since the pathophysiology of TBI is highly complex and variable with multiple factors playing their roles simultaneously, it's a very tiring job to find out a novel agent. When we try to alter one of the patho-physiological aspect of TBI, it affects many targets at a time, in the same manner as the waves affect the multiple points when we throw an object into pond. As the many aspects of the TBI are still obscured, it will be of great difficulty to search a suitable target which has got pronounced effect on TBI. The failure of newer molecules to fulfill its promise can also be attributed to the flaw in pre-clinical and clinical studies. Eventually the journey to have an ideal neuroprotective agent still continues.

**Key Words:** Traumatic Brain Injury, Neuro-Protection.

---

### I. Introduction

Traumatic brain injury (TBI), also known as acquired brain injury, head injury, or brain injury, causes substantial disability and mortality. Traumatic Brain Injury is a significant public health problem worldwide and is predicted to surpass many diseases as a major cause of death and disability by the year 2020<sup>1</sup>. The majority of TBI cases (60%) are a result of road traffic injuries (RTI), followed by falls (20-30%), and violence (10%)<sup>2</sup>. In India it is estimated nearly 1.6 million individuals will sustain TBI and seek hospital care annually<sup>3</sup>. RTI are the leading cause of TBI in India accounting for 45-60% of TBI, and falls account for 20-30% of TBI, paralleling the findings from the Global Burden of Disease Study<sup>4</sup>. Traumatic brain injury causes mechanical tissue destruction which can be supposed to be the primary mechanism of brain injury that results in neuronal cell death causing cerebral edema and rise in intracranial tension contributing to impaired cerebral vasoregulation, cerebral ischemia/hypoxia and brain damage. Primary injury itself acts as trigger for secondary mechanism responsible for brain injury i.e. the neuronal cell death associated with cerebral ischemia is due to the lack of oxygen and glucose, and may involve the loss of ATP, excitotoxicity of glutamate, oxidative stress, reduced neurotrophic support, and multiple other metabolic stresses<sup>5</sup>. One major event taking place at the moment of TBI is the massive ionic in flux referred to as traumatic depolarization. Excitatory amino acids may play a vital role in this depolarization. This represents one of the most important mechanisms of diffuse neuronal cell dysfunction and damage associated with TBI. Cerebral edema and associated increased intracranial pressure are the major immediate consequences of TBI that contribute to most early deaths. There are at least two kinds of delayed and progressive pathological changes induced by TBI. One of these is axonal damage, which is not the direct consequence of traumatic tissue tearing. Rather, results from complex axolemmal or cyto-skeletal changes, or both, which lead to cyto-skeletal collapse and impairment of axoplasmic transport. The other change in traumatized brains occurs concomitantly with compromised blood brain barrier (BBB) function. Secondary damage in TBI is influenced by changes in cerebral blood flow (CBF), cerebral metabolic dysfunction and inadequate cerebral oxygenation. Excitotoxic cell damage and inflammation may lead to apoptosis<sup>6</sup>. Furthermore, it is also becoming clear that these secondary insults are, to a significant degree, are preventable. Since multiple derangements starts simultaneously it is essential to have effective neuroprotective therapy to prevent early brain damage. Management of head injury focuses on preventing, detecting and correcting the secondary brain injury after trauma. Duration and severity of such secondary brain damage influences the possible outcome. Unfortunately, numerous neuroprotective drugs have failed to demonstrate beneficial effects in Phase II/III clinical trials, despite

previous encouraging preclinical results<sup>7</sup>. However, some drugs that have been approved for use in the clinic have neuroprotective effects, and these could be used for the treatment and improvement in functional recovery in patients of traumatic brain injury.

## **II. Neurogenesis after TBI:**

First proposed nearly a century ago, the persistence of neural stem cells and neurogenesis in the adult mammalian central nervous system (CNS) is now accepted<sup>8</sup>. Adult neurogenesis is found in these forebrain regions in all mammalian species examined, including humans and may serve to replace cells damaged by brain insults<sup>9,10</sup>. Significant self-recovery occurs following all but the most severe episodes of TBI<sup>11</sup>. The mechanisms underlying this remain unclear, though injury-induced neurogenesis is one compelling potential contributor to post-injury recovery<sup>12</sup>. The two most well studied and validated reservoirs for neural stem and progenitor cells in mammals are the SVZ of the lateral ventricles and the SGZ of the dentate gyrus<sup>13</sup>. Normally, the postnatal SVZ contributes progenitors to the rostral migratory stream to support ongoing olfactory neurogenesis, while the SGZ of the dentate gyrus provides new granular neurons throughout life<sup>13,14</sup>. Following TBI, progenitor cells in each of these areas become activated, though it is still unclear whether this activation results in stable and productive neurogenesis<sup>12</sup>. Stem cells from the adult brain proliferate and differentiate into neurons and glia in tissue culture with the same efficiency for neuronal differentiation as found in fetal stem cells. Recent studies have shown that they can differentiate to neurons in the adult human dentate gyrus in vivo. Injury to the hippocampus has been associated with learning and memory deficits, which are the hallmarks of TBI. Neurogenesis in this region has been implicated in learning and memory functions<sup>15</sup>. Despite a constant decline in NPC proliferation in the dentate gyrus from adolescence through senescence<sup>16-18</sup>, the adult brain up-regulates proliferation to almost the same extent as the juvenile brain after TBI<sup>19</sup>, indicating that NPCs conserve the ability to respond to proliferative signals. Although neurotrophins are upregulated after TBI<sup>20</sup>, the aging hippocampus has been reported to contain less brain-derived neurotrophic factor (BDNF) in comparison to the hippocampus of younger injured animals after kainic acid injury<sup>21</sup>, suggesting that the neurotrophic response to injury in the hippocampus is age dependent, which may be responsible for better outcome in children and young adults.

### ***Role of neurotrophic factors***

Neurotrophic factors are endogenous proteins which have been shown to play a critical role in the normal development of neurons and appear to mediate a protective response to trauma and disease. They may also play important role in neuro plasticity. It is seen in animal studies that a particular neurotrophic factor increases survival of specific variety of neuron and it may have negligible effect on other class of neurons. During development, neurotrophins almost undoubtedly enhance cell survival by regulating the natural cell death process termed apoptosis<sup>22</sup>. Although the exact role of neurotrophins in neurogenesis following TBI has not been well established, but studies are going on to unlock the facts behind this mystery.

### ***Neuroplasticity***

Neuroplasticity (cortical re-mapping) refers to the ability of the human brain to change as a result of one's experience, so the brain is 'plastic' and 'malleable'. A surprising consequence of neuroplasticity is that the brain activity associated with a given function can move to a different location; this can result from normal experience and also occurs in the process of recovery from brain injury. Neuroplasticity is the fundamental issue that supports the scientific basis for treatment of acquired brain injury with goal-directed experiential therapeutic programs in the context of rehabilitation approaches to the functional consequences of the injury. The evidence for neurogenesis is mainly restricted to the hippocampus and olfactory bulb, but current research has revealed that other parts of the brain, including the cerebellum, may be involved as well<sup>23</sup>. Two aspects of neuronal plasticity are important for information processing: plasticity of intrinsic excitability, that is, the change in ion channel properties; and synaptic plasticity and the change in the strength of synapses between two neurons. Several mechanisms are likely to be involved in brain plasticity. Activity-dependent modification of synaptic connections and reorganization of adult cortical areas are thought to involve long-term potentiation (LTP) and long-term depression (LTD), mechanisms by which information is stored in the mammalian central nervous system<sup>24,25</sup>. Long-term potentiation (LTP) is a rapidly developing persistent enhancement of the postsynaptic potential response to presynaptic stimulation after a brief period of rapidly repeated stimulation of the presynaptic neuron. It may be much more prolonged and can last for days. LTD is the opposite of LTP. It resembles LTP in many ways, but it is characterized by a decrease in synaptic strength. It is produced by slower stimulation of presynaptic

neurons and is associated with a smaller rise in intracellular  $Ca^{2+}$  than occurs in LTP. It is believed to be due to dephosphorylation of AMPA receptors, decreasing their conductance and facilitating their movement away from the synaptic plasma membrane. Synaptic plasticity in cortical horizontal connections has been proposed to underlie cortical map reorganization<sup>26-28</sup>. Glutamate, the main excitatory neurotransmitter, plays a crucial role. Cortical map reorganization in the primary somatosensory cortex can be prevented by blockade of N-methyl-D-aspartate (NMDA) receptors<sup>29-31</sup>.  $\gamma$ -Aminobutyric acid (GABA)-A receptor antagonists can facilitate LTP induction in neocortical synaptic systems, and the induction can be blocked by GABA-A receptor agonists<sup>28</sup>, indicating a complex interplay between excitatory and inhibitory neurotransmitters. Transmitters released by the diffuse neuromodulatory systems originating in locus coeruleus (noradrenaline), nucleus basalis (acetylcholine), lateral tegmentum (dopamine), and raphe nuclei (serotonin) may modify the process<sup>32-33</sup>.

The neurogenesis do occur after excitotoxic and mechanical lesions in dentate gyrus in rats<sup>34</sup> and must also be there in human being. Implantation of fetal neocortical cells after cortical lesions has been performed successfully in several laboratories<sup>35</sup>. Transplanted cells can interact with the host tissue by forming connections but also by being a source of trophic factors that can influence the surrounding tissue. Although both anatomic and functional integration with the host brain have been observed<sup>36</sup>, improvement in behavioral tests have been noticed only when transplantation is combined with posttransplantation housing in an enriched environment<sup>37-38</sup>.

Hippocampal neurogenesis is dependent on both genetic<sup>39</sup> and environmental factors<sup>40</sup>, but the mechanisms underlying the effect of social interaction in early and adult life on hippocampal neurogenesis are largely unknown. Neurons respond differentially to specific temporal and spatial patterns of inputs. This response specificity is not always programmed into neurons; rather, it can develop as the animal interacts with the environment. Social environments can modify neurogenesis and synaptic plasticity in adult hippocampal regions, which is associated with alterations in spatial learning and memory<sup>41</sup>, indicated by manipulating these things during rehabilitation we can improve the recovery following TBI.

Although the mechanism of neurogenesis is not clear, but it exists as indicated by recovery of learning, memory, attention etc. after few months of TBI. This may take few months to year but definitely recovery of functions do occur.

### **III. Pharmacological Targetting Of Various Mechanisms For Neuroprotection:**

#### **1. Free radical scavengers:**

The pre-clinical studies of traumatic brain injury evaluating antioxidants and free radical scavengers have shown promising results but translation into clinical practice is still required<sup>42,43</sup>. There were two multicentric clinical trials evaluating Tirilazad Mesylate, 21-aminosteroid which inhibits lipid peroxidation, which resulted in no significant effects in moderately and severely injured patients<sup>44,45</sup>. Another clinical study in Europe evaluating the NOS inhibitor, lubeluzole, to treat TBI was stopped, mainly after the drug failed to significantly decrease mortality after ischemic stroke, despite evidence for an improvement in the outcome of surviving patients<sup>46-47</sup>. Edaravone was the first free radical scavenger developed as a neuroprotective drug to be used for the treatment of stroke<sup>48</sup>. Edaravone administration following TBI inhibited free radical mediated degeneration of neurons and apoptotic cell death around the damaged area, with improvement in cerebral dysfunction following TBI in rats<sup>49</sup>. Alongwith all these pharmacological effects edaravone also increased neural stem cell numbers around the area of damage following rat TBI<sup>50</sup>. Clinical studies are going on for testing the efficacy of it<sup>51</sup>. Ebselen is a mimic of GSH peroxidase which also reacts with peroxynitrite<sup>52</sup> and can inhibit enzymes such as lipoxygenases<sup>53</sup>, NO synthases<sup>54</sup>, NADPH oxidase<sup>55</sup>, protein kinase C<sup>56</sup>. Ebselen protected the brain from ischemic damage in the acute stage<sup>57</sup>. The ceria<sup>58</sup> and yttria<sup>59</sup> nanoparticles act as direct antioxidants to limit the amount of reactive oxygen species required to kill the cells and this group of nanoparticles could be used to modulate oxidative stress in biological systems. Still research is going on with the hope to find an ideal antioxidant which will also be useful in clinical setting.

#### **2. Neuroimmunophilins:**

Cyclosporin A (CsA) an 11-member cyclic peptide is known to be potent immunosuppressant and is widely used in organ transplantation and auto-immune disorders<sup>60,61</sup>. In the last few years this useful compound has become of great interest to neuroscientists for their unique neuroprotective and neuroregenerative effects. This exerts its effects via immunophilins, the protein receptors for these agents. The immunophilin ligands show promise as a novel class of neuroprotective and neuroregenerative agents that have the potential to treat a variety of neurologic disorders. Cyclosporin A, at therapeutically relevant concentrations, acts directly on neural precursor

cells to enhance their survival both *in vitro* and *in vivo*<sup>62</sup>. This action of Cyclosporin A is promising for the development of regenerative strategies which aim to repair and regenerate damaged or diseased Central Nervous System tissue. It has been shown to block mitochondrial permeability transition pores which in open state causes mitochondrial dysfunction and preservation of mitochondrial function assessed by tissue oxygen consumption, directly translated into improvements in motor and cognitive behavior<sup>63</sup>. At higher doses the Cyclosporin A as well as Tacrolimus have been shown to markedly decrease expressions of Cytochrome C, AIF, caspase-3 and inhibited apoptosis pathways. It has also been shown that CsA treatment against spinal cord hypoxia induced damage is mediated via their antioxidant actions<sup>64</sup>, i.e. it can also act as antioxidant. Tacrolimus has also been shown to be neuroprotective in case of ischemic brain damage<sup>65</sup>. Although the CsA has shown promise in preclinical studies but in the clinical settings failed to replicate the results. In patients with acute traumatic brain injury who received cyclosporine administered intravenously, with treatment initiated within 8 hours of injury, the rate of mortality or other adverse events was not significantly different from that of the placebo group<sup>66</sup>. Tacrolimus has also shown neuroprotective activity in non human primates<sup>67</sup>.

### **3. Antidepressants and Mood stabilisers:**

Mood stabilizers, atypical anti-psychotics and anti-depressants have been reported to have delayed therapeutic effects taking few weeks to months to exhibit their efficacy<sup>68</sup>. This suggests that adaptive changes in cellular signaling cascades may underlie their therapeutic effects<sup>69-71</sup>. Mood stabilizers are known to activate the intracellular MAPK/ERK (mitogen activated protein kinase & extracellular signal-related kinase) signaling pathway<sup>72-74</sup> which is used by neurotrophins, neurotransmitters, and neuropeptides to exert their effects by increasing progenitor cell proliferation and differentiation, neuronal growth, regeneration, and survival, with long-term synaptic remodeling and plasticity<sup>75-78</sup>. The detailed discussion of MAPK/ERK pathway is beyond the scope of this review. Valproate primarily an anti-psychotic agent and also a mood stabilizer leads to activation of the ERK pathway which can be seen in primary cortical neurons<sup>79</sup>, cerebral progenitor cells<sup>80</sup>, hippocampal progenitor cells<sup>81</sup>. Atypical antipsychotics attenuate both cognitive and non-cognitive behavioral impairments in different animal models of neurotoxicity<sup>82-84</sup>. Their beneficial behavioral effects are not only related to their dopamine and serotonin receptor blockade effects, but also to their effects on neuroprotection, neurotrophins and neurogenesis which require further exploration. Among anti-depressants the SSRI fluoxetine prevents the neurotoxicity<sup>85</sup> which may involve MAPK and BDNF<sup>86</sup>. MAOIs protect against dopaminergic neural toxicity<sup>87</sup> & Ladostigil, a MAOI has promising neuroprotective effects, owing to activation of Bcl-2 & BDNF<sup>88</sup>. Early and long-term effects by these agents will guide next generation of therapeutics.

### **4. Antiepileptics :**

There has been a growing interest in the use of antiepileptic drugs (AEDs) for neuroprotection. Voltage-gated sodium channels contribute to the development of axonal degeneration in white matter tracts, so sodium channel blocking drugs can have a protective effect on injured white matter axons. Neuronal injury results due to accumulation of calcium within injured neurons and their axons due to reverse operation of the Na-Ca exchanger, that is triggered by an increase in intracellular sodium due to sodium influx via persistently activated voltage-gated sodium channels<sup>89-90</sup>. Pharmacologic blockade of voltage-gated sodium channels can prevent axonal degeneration and preserve function after a variety of insults to axons. Levetiracetam is found to be neuroprotective in clinically relevant animal models of subarachnoid hemorrhage & closed head injury<sup>91</sup>. Levetiracetam may be a therapeutic alternative to phenytoin following acute brain injury in the clinical setting when seizure prophylaxis is indicated<sup>92-93</sup>. Topiramate is shown to prevent excitotoxic brain damage<sup>94</sup> hence can be considered as a candidate therapy for neuroprotection. Phenytoin provides neuroprotection and improves functional outcome after experimental spinal cord injury<sup>95</sup>, and warrants further exploration as a potential treatment strategy in clinical setting. So these are the potential neuroprotective antiepileptic drugs which can be exploited for the same purpose.

### **5. Anti-inflammatory agents :**

Overexpression of COX-2 appears to be both a marker and an effector of neural damage after a variety of acquired brain injuries, and in natural or pathological aging of the brain. COX-2 expression is increased for prolonged periods in brain regions specifically associated with functional deficits after neurotrauma<sup>96</sup>. COX-2 has been associated with worse outcomes after brain injury, as well as early onset dementia<sup>97</sup>. COX-2 inhibitors may be neuroprotective in the brain by reducing prostanoid and free radical synthesis, or by directing arachidonic acid down alternate metabolic pathways.

Inhibition of COX-2 after pathological insult has been shown to benefit recovery in the brain and spinal cord<sup>98</sup>. COX-2 inhibitors are potent neuroprotectants in vitro and in vivo. Inhibition of COX-2 protected neurons in mixed cultures against N-methyl D-Aspartate (NMDA) excitotoxicity<sup>99</sup>.

Minocycline, a tetracycline antibiotic, & has demonstrated neuroprotective qualities in experimental models of CNS trauma, stroke, spinal cord injury, and neurodegenerative diseases. Recent studies have focused on the anti-inflammatory<sup>100,101</sup> & antiapoptotic<sup>102,103</sup> properties of minocycline. Minocycline in human trials of stroke<sup>104</sup> and spinal cord injury<sup>105</sup> has shown positive results, but further study is warranted in order to draw a handsome conclusion.

Further interleukin-1 receptor antagonist (IL-1ra) is an important anti-inflammatory cytokine which blocks all known actions of IL-1 showing potent, sustained, neuroprotective effects<sup>106</sup>. In experimental models of TBI, IL-1 receptor following trauma contributes to the pathology and that antagonism can reduce both anatomical and functional consequences of neuroinflammation<sup>107</sup>. A randomised phase II study of interleukin-1 receptor antagonist in acute stroke patients was undertaken showing its biological activity relevant to the disease process and its clinical outcome indicated by a greater proportion of patients receiving rhIL-1ra with minimal or no disability at 3 months compared with placebo<sup>108</sup> so it's a potential new therapeutic agent for neuroprotection. Beside antithrombotic property Aspirin also has direct neuroprotective effects because of inhibition of glutamate release via recovery of ATP levels<sup>109</sup> and inhibition of serotonergic activity<sup>110</sup>. It also inhibits glutamate-mediated induction of nuclear factor kappa B which is an anti-inflammatory action which may contribute indirectly to neuroprotection<sup>111</sup>. Prostaglandin E<sub>2</sub> EP1 receptors are essential for the neurotoxicity mediated by COX-2-derived prostaglandin E<sub>2</sub>. EP1 receptors disrupt Ca<sup>2+</sup> homeostasis by impairing Na<sup>+</sup>-Ca<sup>2+</sup> exchange, a key mechanism by which neurons cope with excess Ca<sup>2+</sup> accumulation after an excitotoxic insult<sup>112</sup>. Subsequent increase in intracellular calcium concentration is an important factor for excitotoxic death ultimately contributing to neuronal death<sup>113</sup>. Microglial modulation of neuronal excitotoxicity through interaction with the EP1 receptor and may have important role in neuronal injury as microglia are associated with neuronal injury<sup>114</sup>. So EP1 receptor inhibition may be a potentially valuable strategy for neuroprotection that deserves further investigation.

## **6. Anti-apoptosis agents :**

Apoptosis classically defined as Programmed cell death which is a highly complex process and discussion of its mechanism is beyond the scope of review. Caspases, or cysteine-aspartic proteases or cysteine-dependent aspartate-directed proteases are a family of cysteine proteases that play essential roles in apoptosis, necrosis, and inflammation<sup>115</sup>. Activation of microglia and inflammation-mediated neurotoxicity are suggested to play a decisive role in the pathogenesis of several neurodegenerative disorders. Activated microglia release pro-inflammatory factors that may be neurotoxic. Caspase-8 and caspase-3/7 are involved in regulating microglia activation and inhibition of these caspases could be neuroprotective<sup>116</sup>. Caspase-3 contributes to delayed cell death and brain injury after neonatal hypoxia-ischemia and that calpain activation is associated with and likely a marker for the early component of excitotoxic/necrotic brain injury<sup>117</sup>. Olesoxime is a cholesterol-oxime compound family of mitochondrial pore modulators<sup>118</sup> having cholesterol-like structure and exhibits potent neuroprotective properties in preclinical studies by promoting the function and survival of neurons through interactions with the mitochondrial permeability transition pore (mPTP)<sup>119</sup>. Presently Olesoxime is undergoing a phase II clinical trial as treatment for spinal muscular atrophy<sup>120</sup>. The calpains are calcium-dependent intracellular proteases that are activated in a number of pathogenic conditions. The proposed physiologic roles of calpains in the brain include regulation of neurite outgrowth<sup>121</sup>, long term potentiation<sup>122</sup>, and synaptic remodeling<sup>123</sup>. A bimodal increase in activity of calpain has been observed after brain ischemia, transient at 1 hour, peak activity at 24-48 hours with continuous increase in number of neurons showing calpain activity<sup>124</sup>. Targeting those intracellular processes, represents a viable therapeutic strategy for limiting neurological damage after ischemia as indicated by cell-penetrating calpain inhibitor when administered systemically<sup>125</sup>. Leupeptin, also known as *N*-acetyl-L-leucyl-L-leucyl-L-argininal, is a naturally occurring protease inhibitor that can inhibit cysteine, serine and threonine peptidases. Both calpain inhibitor I and leupeptin protected neurons against ischemic and hypoxic damage i.e they are neuroprotective as indicated by increased cell viability and protein content in the cultures, and fewer damaged neurons in the hippocampal CA1-subfield<sup>126</sup>. The nuclear enzyme poly(ADP-ribose) polymerase (PARP) has been shown to be activated following traumatic brain injury (TBI), binds to DNA strand breaks and utilizes nicotinamide adenine dinucleotide which can lead to cell injury via severe, irreversible depletion of the NAD and ATP pool, and PARP-1 inhibitors have been expected to rescue neurons from degeneration in a number of disease models<sup>127</sup>.

## **7. Excitotoxicity:**

Glutamate is supposed to be involved in an acute excitotoxic process that occurs immediately after ischaemic or traumatic injury but, after this early time period, glutamate may then resume its normal physiological functions, including facilitation of neuronal survival. NMDA (N-methyl-D-aspartate) receptor targeting drugs developed as neuroprotective agents, specifically selective NMDA receptor antagonists, have not been effective in large randomised controlled trials of adequate methodological quality for most selected indications, primarily ischaemic stroke and traumatic brain injury<sup>128</sup>. The drugs that completed trials included antagonists of the glutamate site (selfotel)<sup>129</sup> and the glycine site (gavestinel)<sup>130</sup>, an antagonist of the ionchannel site (aptiganel)<sup>131</sup>, and an NR2B subunit-selective antagonist (traxoprodil)<sup>132</sup>. Recently, increasing evidence based on molecular studies suggests that memantine, an uncompetitive NMDA receptor blocker with fast channel unblocking kinetics to prevent it from occupying the channels and interfering with normal synaptic transmission, is a potent neuroprotectant<sup>133</sup>. As a neuroprotective agent, memantine can reduce functional as well as morphological sequelae induced by ischemia<sup>134</sup>.

Glutamate carboxypeptidase II (GCPII), also known as N-acetyl-L-aspartyl-L-glutamate peptidase I (NAALADase I), NAAG peptidase, is a zinc metalloenzyme that resides in membranes, which catalyzes the hydrolysis of N-acetylaspartylglutamate (NAAG) to glutamate and N-acetylaspartate (NAA)<sup>135-136</sup>. NAAG is one of most commonly found neurotransmitters the central nervous system<sup>137</sup>. Glutamate is a common and abundant excitatory neurotransmitter in the central nervous system; however excess of glutamate transmission, can damage neurons due to excitotoxicity and has been implicated in many neurological diseases and disorders<sup>137</sup>. NAALADase inhibitor 2 (phosphonomethyl) pentanedioic acid (2-PMPA) has shown efficacy in protecting neurons against injury caused by cellular anoxia<sup>138</sup>. NAALADase inhibition may represent a novel glutamate regulating strategy devoid of the side-effects that have hampered development of postsynaptic glutamate receptor antagonists. A lead NAALADase inhibitor termed GPI 5693 in phase I clinical testing was found to be safe and tolerable in healthy subjects<sup>139</sup>.

AMPA (alpha-amino-3-hydroxy-5-methyl-4-isoxazole propionic acid) receptor are both glutamate receptors and cation channels which are important to plasticity and synaptic transmission. Modifications in AMPA receptors at the postsynaptic membrane cause changes in synaptic strength, and is a key regulator of various forms of synaptic plasticity. Thus, enhancing the activity of AMPA receptors has been shown to increase production of certain growth factors<sup>140</sup> and to regulate the mechanism of neurite growth<sup>141</sup>. The agents which positively modulate the activity of AMPA receptors can be exploited to treat a variety of neuropsychiatric disorders.

Dexanabinol is a synthetic cannabinoid derivative which does not act as a cannabinoid receptor agonist, instead act as NMDA receptor antagonist<sup>142</sup>. It is anticonvulsant and neuroprotective, and is implicated in studies for treatment of head injury<sup>143</sup>. It was shown to be safe in clinical trials<sup>144</sup>.

## **8. Erythropoietin (EPO) :**

EPO is made locally in response to injury or metabolic stress as a protective factor in brain, spinal cord, or heart<sup>145</sup>. A large body of experimental evidence suggests that rhEPO is neuroprotective after different types of cerebral tissue injuries such as ischemia<sup>146</sup> or traumatic brain injury<sup>147</sup>. The protective effects of rhEPO were associated with a decrease in the number of apoptotic neurons<sup>148</sup> and an increase in the expression of the anti-apoptotic bcl-2 gene<sup>149</sup>. A recent retrospective study in patients with severe TBI, showed that administration of rhEPO was associated with a significantly lower in hospital mortality in comparison to match cases controls<sup>150</sup>. A large phase II trial, initially conducted to study the effects of rhEPO in critically ill patients as an erythropoietic agent, demonstrated in a post hoc subgroup analysis demonstrated that weekly administered subcutaneous rhEPO significantly decreased mortality in the trauma subgroup<sup>151</sup>. rhEPO administration after TBI may be neurorestorative<sup>152</sup> by enhancing neurogenesis and also probably by affording vascular protection, thereby promoting the creation of vascular niches for neurogenesis.

## **9. $\alpha 2$ Agonist :**

Dexmedetomidine has shown neuroprotective property in a many in vivo and in vitro studies<sup>153</sup>, however, the mechanism of neuroprotection remains unclear and may involve Modulation of pathways leading to excitatory cell death and apoptosis. Clinical trial for assessing safety in TBI patients abandoned<sup>154</sup>.

## 10. Davunetide :

Activity-dependent neuroprotective protein (ADNP) differentially interacts with chromatin to regulate essential genes. NAP is a peptide derived from ADNP that interacts with microtubules and provides potent neuroprotection<sup>155</sup>. Phase 2 trials are being carried out<sup>156</sup>.

## 11. Neurosteroids:

Naturally occurring neurosteroids are potent allosteric modulators of the GABA receptor and through augmentation of this receptor function can protect neuronal cells against NMDA over activation, ischemia and TBI<sup>157</sup>. Progesterone appears to exert its neuroprotective effects in TBI by protecting or rebuilding the BBB, decreasing the development of cerebral edema, down-regulating the inflammatory cascade, and limiting cellular necrosis and apoptosis<sup>158</sup>. Recognition of the neuroprotective potential of progesterone has recently led to the completion of a clinical trial named “ProTECT” (progesterone for traumatic brain injury (TBI), experimental clinical treatment), this trial tested the usefulness of progesterone as treatment for moderate to

Severe TBI<sup>159</sup> The patients in the moderate TBI group given progesterone tended to have better functional outcomes, although progesterone had no effect on the disability of severe TBI survivors at the 30-day time point.

Reduced levels of estradiol have been shown to compromise the functioning and survival of neurons and to result in alterations in memory processes<sup>160</sup>. In the brain, the synthesizing enzyme aromatase plays a very important role in neuroprotection. Males as well as females are sensitive to the protective effects of estrogen, as estradiol has been shown to prevent ischemia-induced learning disability and neuronal loss in both sexes. Some of the neuroprotective effects of androgens are mediated by their conversion to estrogens. However, androgens also exert neuroprotective and neuroregenerative effects on their own.

This outline shows that the major groups of steroid hormones exert pleiotropic effects in the nervous system and influence the viability and regenerative capacity of neurons.

There exists a variety of other classes of drugs exist which we haven't discussed in this review, alternatively some novel non-pharmacological approaches are also there which are worth mentioning e.g. Oxygen carriers, hypothermia, exercise, trans-cranial magnetic stimulation etc. The use of a particular approach after TBI depend on patient's status and varies from patient to patient. Since the newer aspects of mechanism of conventional drugs are being elaborated this will boost our approach to look for neuroprotective agent.

## References

- [1] World Health Organization. Projections of Mortality and Burden of Disease to 2030: Deaths by Income Group. Geneva; 2002 12/01/06.
- [2] Gururaj G. An Epidemiological Approach To Prevention – Prehospital Care And Rehabilitation In Neurotrauma. *Neurology India* 1995;43(3):95-105
- [3] Gururaj G. Epidemiology of traumatic brain injuries: Indian scenario. *Neurological Research* 2002;24:24-8.
- [4] Murray CJ, Lopez AD. *Global Health Statistics: A Compendium of Incidence, Prevalence and Mortality Estimates for Over 200 Conditions* Cambridge: Harvard University Press; 1996.
- [5] Dirnagl, U.; Iadecola, C.; Moskowitz, M.A. Pathobiology of ischaemic stroke: An integrated view. *Trends Neurosci.* 1999, 22, 391–397.
- [6] Werner, C. and Engelhard, K. (2007) Pathophysiology of traumatic brain injury. *Br.J. Anaesth.* 99, 4–9
- [7] Amaro, S.; Chamorro, A. Translational stroke research of the combination of thrombolysis and antioxidant therapy. *Stroke* 2011, 42, 1495–1499.
- [8] Allen, E., 1912. The cessation of mitosis in the central nervous system of the albino rat. *J. Comp. Neurol.* 22, 547–568.
- [9] Eriksson, P.S., et al., 1998a. Neurogenesis in the adult human hippocampus. *Nat. Med.* 4, 1313–1317.
- [10] Curtis, M.A., et al., 2007. Human neuroblasts migrate to the olfactory bulb via a lateral ventricular extension. *Science* 315, 1243–1249.
- [11] Anderson, V.A., et al., 2000. Recovery of memory function following traumatic brain injury in pre- school children. *Brain Inj.* 14, 679–692.
- [12] Richardson, R.M., et al., 2007. Neurogenesis after traumatic brain injury. *Neurosurg. Clin. N. Am.* 18, 169–181 xi.
- [13] Alvarez-Buylla, A., et al., 2001. A unified hypothesis on the lineage of neural stem cells. *Nat. Rev. Neurosci.* 2, 287–293.
- [14] Temple, S., Qian, X., 1995. bFGF, neurotrophins, and the control of cortical neurogenesis. *Neuron* 15, 249–252.
- [15] Leuner B, Gould E, Shors TJ. Is there a link between adult neurogenesis and learning? *Hippocampus* 2006;16(3):216–24.
- [16] Kuhn HG, Dickinson-Anson H, Gage FH. Neurogenesis in the dentate gyrus of the adult rat: age-related decrease of neuronal progenitor proliferation. *J Neurosci* 1996;16(6):2027–33.
- [17] Driscoll I, Howard SR, Stone JC, et al. The aging hippocampus: a multi-level analysis in the rat. *Neuroscience* 2006;139(4):1173–85.
- [18] Heine VM, Maslam S, Joels M, et al. Prominent decline of newborn cell proliferation, differentiation, and apoptosis in the aging dentate gyrus, in absence of an age-related hypothalamus-pituitary-adrenal axis activation. *Neurobiol Aging* 2004;25(3):361–75

- [19] Sun D, Colello RJ, Daugherty WP, et al. Cell proliferation and neuronal differentiation in the dentate gyrus in juvenile and adult rats following traumatic brain injury. *J Neurotrauma* 2005;22(1):95–105.
- [20] Oyesiku NM, Evans CO, Houston S, et al. Regional changes in the expression of neurotrophic factors and their receptors following acute traumatic brain injury in the adult rat brain. *Brain Res* 1999;833(2):161–72.
- [21] Shetty AK, Rao MS, Hattiangady B, et al. Hippocampal neurotrophin levels after injury: relationship to the age of the hippocampus at the time of injury. *J Neurosci Res* 2004;78(4):520–32.
- [22] Franklin, J.L. and Johnson, E.M., Jr. (1994) *Phil. Trans. Roy. Soc. B. London* 345-251
- [23] Ponti, Giovanna; Peretto, Paolo; Bonfanti, Luca; Reh, Thomas A. (2008). "[Genesis of Neuronal and Glial Progenitors in the Cerebellar Cortex of Peripuberal and Adult Rabbits](#)". *PLoS ONE* 3 (6): e2366
- [24] Bear MF, Malenka RC. Synaptic plasticity: LTP and LTD. *Curr Opin. Neurobiol.* 1994;4:389–399.
- [25] Buonomano DV, Merzenich MM. Cortical plasticity: from synapses to maps. *Annu Rev Neurosci.* 1998;21:149–186.
- [26] Hess G, Donoghue JP. Long-term potentiation of horizontal connections provides a mechanism to reorganize cortical motor maps. *J Neurophysiol.* 1994;71:2543–2547.
- [27] Das A, Gilbert CD. Long-range horizontal connections and their role in cortical reorganization revealed by optical recording of cat primary visual cortex. *Nature.* 1995;375:780–784.
- [28] Hess G, Aizenman CD, Donoghue JP. Conditions for the induction of long-term potentiation in Layer II/III horizontal connections of the rat motor cortex. *J Neurophysiol.* 1996;75:1765–1777
- [29] Kano M, Lino K, Kano M. Functional reorganization of adult cat somatosensory cortex is dependent on NMDA receptors. *Neuroreport.* 1991;2:77–80.
- [30] Hess G, Jacobs KM, Donoghue JP. N-Methyl-D-aspartate receptor mediated component of field potentials evoked in horizontal pathways of rat motor cortex. *Neuroscience.* 1994;61:225–235.
- [31] Garraghty PE, Muja N. NMDA receptors and plasticity in adult primate somatosensory cortex. *J Comp Neurol.* 1996;367:319–326.
- [32] Kilgard MP, Merzenich MM. Cortical map reorganization enabled by nucleus basalis activity. *Science.* 1998;279:1714–1718
- [33] . Kirkwood A, Rozas C, Kirkwood J, Perez F, Bear MF. Modulation of long-term synaptic depression in visual cortex by acetylcholine and norepinephrine. *J Neurosci.* 1999;19:1599–1609.
- [34] Gould E, Tanapat P. Lesion-induced proliferation of neuronal progenitors in the dentate gyrus of the adult rat. *Neuroscience.* 1997;80:427–436.
- [35] . Johansson BB. Neurotrophic factors and transplants. In: Goldstein LB, ed. *Restorative Neurology: Advances in Pharmacotherapy for Recovery After Stroke*. Armonk, NY: Futura Publishing Co; 1998:141–166.
- [36] Grabowski M, Brundin P, Johansson BB. Functional integration of cortical grafts placed in brain infarcts of rats. *Ann Neurol.* 1993;34:362–368.
- [37] Grabowski M, Sørensen JC, Mattsson B, Zimmer J, Johansson BB. Influence of an enriched environment and cortical grafting in functional outcome in brain infarcts of adult rats. *Exp Neurol.* 1995;133:96–102.
- [38] Mattsson B, Sørensen JC, Zimmer J, Johansson BB. Neural grafting to experimental neocortical infarcts improves behavioral outcome and reduces thalamic atrophy in rats housed in enriched but not in standard environments. *Stroke.* 1997;28:1225–1232.
- [39] Kempermann, G., Kuhn, H.G., Gage, F.H., 1997b. Genetic influence on neurogenesis in the dentate gyrus of adult mice. *Proc. Natl. Acad. Sci. USA* 94, 10409–10414.
- [40] Jones, G.H., Hernandez, T.D., Kendall, D.A., Marsden, C.A., Robbins, T.W., 1992. Dopaminergic and serotonergic function following isolation rearing in rats: study of behavioural responses and postmortem and in vivo neurochemistry. *Pharmacol. Biochem. Behav.* 43, 17–35.
- [41] Lu, L., Bao, G. et al. Modification of hippocampal neurogenesis and neuroplasticity by social environments. *J Experimental Neurology* 183 (2003) 600–609
- [42] Rigg, J.L.; Elovic, E.P.; Greenwald, B.D. *J. Head Trauma Rehabil.*, 2005, 20, 389-391
- [43] Faden, A.I. *Neuroprotection and Traumatic Brain Injury. Arch. Neurol.*, 2001, 58, 1553-55
- [44] Schouten, J.W. *Curr. Opin. Crit. Care*, 2007, 13, 134-142
- [45] Marshall LF et al. (1998) A multicenter trial on the efficacy of using tirilazad mesylate in cases of head injury. *J. Neurosurg.* 89:519–25.
- [46] Andrews, P.J 1998. Head injury: complications and management. *Curr. Opin. Anaesthesiol.*, 11,5: 473-477
- [47] Grotta J. Lubeluzole treatment of acute ischemic stroke. [Stroke.](#) 1997 Dec;28(12):2338-46.
- [48] Yoneda, Y.; Uehara, T.; Yamasaki, H.; Kita, Y.; Tabuchi, M.; Mori, E. Hospital-based study of the care and cost of acute ischemic stroke in Japan. *Stroke* 2003, 34, 718–724.
- [49] Itoh T, Satou T, Nishida S, et al. Edaravone protects against apoptotic neuronal cell death and improves cerebral function after traumatic brain injury in rats. *Neurochem Res.* 2009;35:348–355.
- [50] Itoh T, Satou T, Nishida S, Tsubaki M, Hashimoto S, Ito H. The novel free radical scavenger, edaravone, increases neural stem cell number around the area of damage following rat traumatic brain injury. *Neurotox Res.* 2009;16:378–389.
- [51] Hao L et al. Phase I Clinical Study of Edaravone in Healthy Chinese Volunteers: Safety and Pharmacokinetics of Single or Multiple Intravenous Infusions. *Drugs in R & D.* 2012;12(2): 65-70
- [52] Masumoto H, Sies H. The reaction of ebselen with peroxynitrite. *Chem Res Toxicol.* 1996;9(1):262-7.
- [53] Walther M. et al. The Inhibition of Mammalian 15-Lipoxygenases by the Anti-Inflammatory Drug Ebselen: Dual-Type Mechanism Involving Covalent Linkage and Alteration of the Iron Ligand Sphere molecular pharmacology, 56:196–203 (1999).
- [54] Koizumi H, Fujisawa H, Suehiro E, Shirao S, Suzuki M. Neuroprotective effects of ebselen following forebrain ischemia: involvement of glutamate and nitric oxide. *Neurol Med Chir* 2011;51(5):337–43.
- [55] Wakamura K, Ohtsuka T, Okamura N, Ishibashi S, Masayasu H. Mechanism for the inhibitory effect of a seleno-organic compound, Ebselen, and its analogues on superoxide anion production in guinea pig polymorphonuclear leukocytes. *J Pharmacobiodyn.* 1990;13(7):421-5
- [56] Cotgreave IA, Duddy SK, Kass GEN, Thompson D, and Moldeus P: Studies on the anti-inflammatory activity of ebselen: ebselen interferes with granulocyte oxidative burst by dual inhibition of NADPH oxidase and protein kinase C. *Biochem Pharmacol* 1989;38,649–656.



- [57] Ogawa A et al. Ebselen in Acute Middle Cerebral Artery Occlusion: A Placebo-Controlled, Double-Blind Clinical Trial *Cerebrovasc Dis* 1999;9:112–118
- [58] Chung D. Nanoparticles have health benefits too, *New Scientist*, (2003)179; 2410–2416
- [59] Bloor, R.J. Brook, M.C. Flemings, S. Mahajan. Yttrium oxide, in: D, *The Encyclopedia of Advanced Materials*, vol. 4, Pergamon Press, Ltd., Oxford, 1994, pp. 2957–2959
- [60] Borel JF, Feurer C, Gubler HU, Stahelin H. Biological effects of cyclosporin A: A new antilymphocytic agent. *Agents Actions* 1976;6:465-8.
- [61] Borel JF, Feurer C, Magnee C, Stahelin H. Effects of the new anti-lymphocytic peptide cyclosporine A in animals. *Immunology* 1977;32:1017-25.
- [62] Hunt J. et al. Cyclosporin A Has Direct Effects on Adult Neural Precursor Cells. *The Journal of Neuroscience*, 2010 • 30(8):2888–2896
- [63] Xie Z. et al. Neuroprotective effect of Cyclosporin A on the development of early brain injury in a subarachnoid hemorrhage model: a pilot study. *Brain Res.* 2012 Sep 7;1472:113-23.
- [64] Yousuf, S., Atif, F., Keshewani, V. and Agrawal, S. K., Neuroprotective effects of Tacrolimus (FK-506) and Cyclosporin (CsA) in oxidative injury. *Brain and Behavior*, 2011,1: 87–94.
- [65] Giordani F. et al. Tacrolimus (FK506) reduces ischemia-induced hippocampal damage in rats: a 7- and 30-day study. *Brazilian Journal of Medical and Biological Research* (2003) 36: 495-502
- [66] Hatton J. et al. Dosing and safety of cyclosporine in patients with severe brain injury. *J Neurosurg.* 2008 ; 109(4): 699–707.
- [67] Furuichi Y. et al. Tacrolimus, a potential neuroprotective agent, ameliorates ischemic brain damage and neurologic deficits after focal cerebral ischemia in nonhuman primates. [J Cereb Blood Flow Metab.](#) 2003 ;23(10):1183-94.
- [68] Goodman and Gillman *Pharmacological Basis of Therapeutics* 12th edition.
- [69] D'Sa C, Duman RS. Antidepressants and neuroplasticity. *Bipolar Disord.* 2002;4:183-194.
- [70] Parikh V, Khan MM, Mahadik SP. [Olanzapine counteracts reduction of brain-derived neurotrophic factor and TrkB receptors in rat hippocampus produced by haloperidol.](#) *Neurosci Lett.* 2004 Feb 12;356(2):135-9
- [71] Bai O, Chlan-Fourney J, Bowen R, et al: Expression of brain-derived neurotrophic factor mRNA in rat hippocampus after treatment with antipsychotic drugs. *J Neurosci Res* 2003;71:127–31
- [72] Creson TK, Yuan P, Manji HK, Chen G. Evidence for involvement of ERK,PI3K, and RSK in induction of Bcl-2 by valproate. *J Mol Neurosci.* 2009;37:123-134.
- [73] Einat H, Yuan P, Gould TD, et al. The role of the extracellular signal-regulated kinase signaling pathway in mood modulation. *J Neurosci.* 2003;23:7311-7316.
- [74] Hao Y, Creson T, Zhang L, et al. Mood stabilizer valproate promotes ERK pathway-dependent cortical neuronal growth and neurogenesis. *J Neurosci.* 2004;24:6590-6599.
- [75] Huang X, Wu DY, Chen G, Manji H, Chen DF. Support of retinal ganglion cell survival and axon regeneration by lithium through a Bcl-2 dependent mechanism. *Invest Ophthalmol Vis Sci.* 2003;44:347-354.
- [76] Sweatt JD. Mitogen-activated protein kinases in synaptic plasticity and memory. *Curr Opin Neurobiol.* 2004;14:311-317.
- [77] Chen G, Creson T, Sharon E, Hao Y, Wang G. Neurotrophic actions of mood-stabilizers: a recent research discovery and its potential clinical applications. *Curr Psych Rev.* 2005;1:173-185.
- [78] Chen G, Manji HK. The extracellular signal-regulated kinase pathway: an emerging promising target for mood stabilizers. *Curr Opin Psychiatry.* 2006;19:313-323.
- [79] Di Daniel E, Mudge AW, Maycox PR. Comparative analysis of the effects of four mood stabilizers in SH-SY5Y cells and in primary neurons. *Bipolar Disord.* 2005;7:33-41.
- [80] Jung GA, Yoon JY, Moon BS, et al. Valproic acid induces differentiation and inhibition of proliferation in neural progenitor cells via the betacatenin- Ras-ERK-p21Cip/WAF1 pathway. *BMC Cell Biol.* 2008;9:66.
- [81] Hsieh J, Nakashima K, Kuwabara T, Mejia E, Gage FH. Histone deacetylase inhibition-mediated neuronal differentiation of multipotent adult neural progenitor cells. *Proc Natl Acad Sci U S A.* 2004;101:16659-16664.
- [82] He J, Yang Y, Yu Y, Li X, Li XM. The effects of chronic administration of quetiapine on the methamphetamine-induced recognition memory impairment and dopaminergic terminal deficit in rats. *Behav Brain Res.* 2006;172:39–45.
- [83] He J, Xu H, Yang Y, Rajakumar D, Li X, Li XM. The effects of chronic administration of quetiapine on the phencyclidine-induced reference memory impairment and decrease of bcl-XL/Bax ratio in the posterior cingulate cortex in rats. *Behav Brain Res.* 2006;168:236–242.
- [84] Luo C, Xu H, Li XM. Quetiapine reverses the suppression of hippocampal neurogenesis caused by repeated restraint stress. *Brain Res.* 2005;1063:32–39.
- [85] Azmitia EC, Murphy RB, Whitaker-Azmitia PM. MDMA (ecstasy) effects on cultured serotonergic neurons: evidence for Ca2(+)-dependent toxicity linked to release. *Brain Res.* 1990;510:97-103.
- [86] Mercier G, Lennon AM, Renouf B, et al. MAP kinase activation by fluoxetine and its relation to gene expression in cultured rat astrocytes. *J Mol Neurosci.* 2004;24:207-216.
- [87] Heikkila RE, Manzino L, Cabbat FS, Duvoisin RC. Protection against the dopaminergic neurotoxicity of 1-methyl-4-phenyl-1,2,5,6-tetrahydropyridine by monoamine oxidase inhibitors. *Nature.* 1984;311:467-469.
- [88] Weinreb O, Amit T, Bar-Am O, Yogev-Falach M, Youdim MB. The neuroprotective mechanism of action of the multimodal drug ladostigil. *Front Biosci.* 2008;13:5131-5137.
- [89] Stys PK et al. Ionic Mechanisms of Anoxic Injury in Mammalian CNS White Matter: Role of Na+ Channels and Na-Ca Exchanger. *The Journal of Neuroscience*, 1992, 12(2): 430-439
- [90] Stys PK et al. Noninactivating, tetrodotoxin-sensitive Na+ conductance in rat optic nerve axons. *Neurobiology* 1993,90:6976-6980.
- [91] Wang H. et al. Levetiracetam is neuroprotective in murine models of closed head injury and subarachnoid hemorrhage. [Neurocrit Care.](#) 2006;5(1):71-8.
- [92] Jones KE, Puccio AM, Harshman KJ, et al. Levetiracetam versus phenytoin for seizure prophylaxis in severe traumatic brain injury. *Neurosurg Focus.* 2008;25(4).
- [93] Szaflarski JP, Sangha KS, Lindsell CJ, Shutter LA. Prospective, randomized, single-blinded comparative trial of intravenous levetiracetam versus phenytoin for seizure prophylaxis. *Neurocrit Care* 2010;12(2):165-72.
- [94] [Sfaello I, Baud O, Arzimanoglou A, Gressens P.](#) Topiramate prevents excitotoxic damage in the newborn rodent brain. [Neurobiol Dis.](#) 2005 Dec;20(3):837-48.

- [195] [Hains BC](#), [Saab CY](#), [Lo AC](#), [Waxman SG](#). Sodium channel blockade with phenytoin protects spinal cord axons, enhances axonal conduction, and improves functional motor recovery after contusion SCI. [Exp Neurol](#). 2004 Aug;188(2):365-77.
- [196] . [Gopez, J.J.](#), [Yue, H.](#), [Vasudevan, R.](#), [Malik, A.S.](#), [Fogelsanger, L.N.](#), [Lewis, S.](#), [Panikashvili, D.](#), [Shohami, E.](#), [Jansen, S.A.](#), [Narayan, R.K.](#), [Strauss, K.I.](#), 2005. Cyclooxygenase-2-specific inhibitor improves functional outcomes, provides neuroprotection, and reduces inflammation in a rat model of traumatic brain injury. [Neurosurgery](#) 56, 590–604.
- [197] [Jellinger, K.A.](#), 2004. Head injury and dementia. [Curr. Opin. Neurol.](#) 17, 719–723.
- [198] [Resnick, D.](#), [Graham, S.](#), [Dixon, C.](#), [Marion, D.](#), 1998. Role of cyclooxygenase-2 in acute spinal cord injury. [J. Neurotrauma](#) 15, 1005–1013.
- [199] [Hewett, S.](#), [Uliasz, T.](#), [Vidwans, A.](#), [Hewett, J.](#), 2000. Cyclooxygenase-2 contributes to N-methyl-D-aspartate-mediated neuronal cell death in primary cortical cell culture. [J. Pharm. Exp. Ther.](#) 293 (2), 417–425.
- [100] [Stirling DP](#), [Koochesfahani KM](#), [Steeves JD](#), [Tetzlaff W](#). Minocycline as a neuroprotective agent. [Neuroscientist](#). 2005; 11(4):308–322.
- [101] [Kim HS](#), [Suh YH](#). Minocycline and neurodegenerative diseases. [Behav Brain Res](#). 2009; 196(2):168–179.
- [102] [Garcia-Martinez EM](#), [Sanz-Blasco S](#), [Karachitos A](#), et al. Mitochondria and calcium flux as targets of neuroprotection caused by minocycline in cerebellar granule cells. [Biochem Pharmacol](#). 2010;79(2):239–250.
- [103] [Antonenko YN](#), [Rokitskaya TI](#), [Cooper AJ](#), [Krasnikov BF](#). Minocycline chelates Ca<sup>2+</sup>, binds to membranes, and depolarizes mitochondria by formation of Ca<sup>2+</sup>-dependent ion channels. [JBioenerg Biomembr](#). 2010; 42(2):151–163.
- [104] [Lampf Y](#), [Boaz M](#), [Gilad R](#), et al. Minocycline treatment in acute stroke: an open-label, evaluator-blinded study. [Neurology](#). 2007; 69(14):1404–1410.
- [105] [Casha S](#), [Zygun D](#), [McGowan D](#), [Yong VW](#), [Hurlbert JR](#). Neuroprotection with minocycline after spinal cord injury: results of a double blind, randomized, controlled pilot study. [Neurosurgery](#). 2009; 65(2):410–411
- [106] [Hannum C](#), [Wilcox et al](#). Interleukin-1 receptor antagonist activity of a human interleukin-1 inhibitor. [Nature](#). 1990;343:336–340
- [107] [Jones NC et al](#). Antagonism of the interleukin-1 receptor following traumatic brain injury in the mouse reduces the number of nitric oxide synthase-2-positive cells and improves anatomical and functional outcomes. [Eur J Neurosci](#). 2005 Jul;22(1):72-8.
- [108] [Emsley HCA](#), et al. A randomised phase II study of interleukin-1 receptor antagonist in acute stroke patients. [J Neurol Neurosurg Psychiatry](#) 2005;76:1366–1372.
- [109] [Cristóbal JD et al](#). Inhibition of glutamate release via recovery of ATP levels accounts for a neuroprotective effect of Aspirin in rat cortical neurons exposed to oxygen-glucose deprivation. [Stroke](#). 2002;33:261-267
- [110] [Ghosh A](#), et al. Evaluation of nootropic and neuroprotective effects of low dose aspirin in rats. [Journal of Pharmacology and Pharmacotherapeutics](#), 2011,2;(1):3-6
- [111] [Grilli M](#), [Pizzi M](#), [Memo M](#), [Spano P](#). Neuroprotection by aspirin and sodium salicylate through blockade of NFκB activation. [Science](#). 1996 Nov 22;274(5291):1383-5.
- [112] [Kawano T](#), [Anrather J](#), [Zhou P](#), [Park L](#), [Wang G](#), [Frys KA](#), [Kunz A](#), [Cho S](#), [Orio M](#), [Iadecola C](#): Prostaglandin E(2) EP1 receptors: downstream effectors of COX-2 neurotoxicity. [Nat Med](#) 2006, 12:225-229
- [113] [Sattler R](#), [Tymianski M](#). Molecular mechanisms of calcium dependent excitotoxicity. [J Mol Med](#) 2000, 78:3-13.
- [114] [Carlson NG et al](#). Microglial inhibition of neuroprotection by antagonists of the EP1 prostaglandin E2 receptor. 2009 [Journal of Neuroinflammation](#) 2009, 6:5
- [115] [Alnemri ES](#), [Emad S](#); et al. Human ICE/CED-3 Protease Nomenclature" [Cell](#) 87 (2): 171.
- [116] [Burguillos MA](#). et al. Caspase signalling controls microglia activation and neurotoxicity. 2011, [Nature](#);472: 318-25
- [117] [Han BH](#). Et al. Selective, Reversible Caspase-3 Inhibitor Is Neuroprotective and Reveals Distinct Pathways of Cell Death after Neonatal Hypoxic-ischemic Brain Injury. [The Journal of Biological Chemistry](#) 2002: 277, 33,(16): 30128–30136.
- [118] [Martin LJ](#). et al. Olesoxime, a cholesterol-like neuroprotectant for the potential treatment of amyotrophic lateral sclerosis. [IDrugs](#). 2010 August ; 13(8): 568–580
- [119] [Bordet T et al](#). Identification and Characterization of Cholest-4-en-3-one, Oxime (TRO19622), a Novel Drug Candidate for Amyotrophic Lateral Sclerosis". [Journal of Pharmacology and Experimental Therapeutics](#) 322 (2): 709–720.
- [120] [Safety and Efficacy of Olesoxime \(TRO19622\) in 3-25 Years SMA Patients.](#). [ClinicalTrials.gov](#).
- [121] [Song DK](#), [Malmstrom T](#), [Kater SB](#), [Mykles DL](#). Calpain inhibitors block Ca(2+)-induced suppression of neurite outgrowth in isolated hippocampal pyramidal neurons. [J Neurosci Res](#) 1994;39(4):474–481.
- [122] [Tomimatsu Y](#), [Idemoto S](#), [Moriguchi S](#), [Watanabe S](#), [Nakanishi H](#). Proteases involved in long-term potentiation. [Life Sci](#) 2002;72(4–5):355–361.
- [123] [Chan SL](#), [Mattson MP](#). Caspase and calpain substrates: roles in synaptic plasticity and cell death. [J Neurosci Res](#) 1999;58(1):167–190.
- [124] [Neumar RW](#), [Meng FH](#), [Mills AM](#), [Xu YA](#), [Zhang C](#), [Welsh FA](#), [Siman R](#). Calpain activity in the rat brain after transient forebrain ischemia. [Exp Neurol](#) 2001;170(1):27–35.
- [125] [Hong SC](#), et al. Neuroprotection With a Calpain Inhibitor in a Model of Focal Cerebral Ischemia. [Stroke](#). 1994;25:663-669
- [126] [Rami A](#), [Kriegelstein J](#). Protective effects of calpain inhibitors against neuronal damage caused by cytotoxic hypoxia in vitro and ischemia in vivo. [Brain Research](#) 1993; 609: (1–2):67–70
- [127] [LaPlaca MC](#), et al. Pharmacologic inhibition of poly(ADP-ribose) polymerase is neuroprotective following traumatic brain injury in rats. [J Neurotrauma](#). 2001;18(4):369-76.
- [128] [Muir KW](#), [Lees KR](#). Excitatory amino acid antagonists for acute stroke. [Cochrane Database Syst Rev](#) 2003; 3: CD001244
- [129] [Morris GF](#), [Bullock R](#), [Marshall SB](#), [Marmarou A](#), [Maas A](#), [Marshall LF](#). Failure of the competitive N-methyl-D-aspartate antagonist Selfotel (CGS 19755) in the treatment of severe head injury: results of two phase III clinical trials. [J Neurosurg](#) 1999; 91: 737–43.
- [130] [Lees KR](#), [Asplund K](#), [Carolei A](#), et al, for the GAIN International Investigators. Glycine antagonist (gavestinel) in neuroprotection (GAIN International) in patients with acute stroke: a randomized controlled trial. [Lancet](#) 2000; 355: 1949–54.
- [131] [Albers GW](#), [Goldstein LB](#), [Hall D](#), [Lesko LM](#). Aptiganel hydrochloride in acute ischemic stroke: a randomized controlled trial. [JAMA](#) 2001; 286: 2673–82.
- [132] [Yurkewicz L](#), [Weaver J](#), [Bullock MR](#), [Marshall LF](#). The effect of the selective NMDA receptor antagonist traxoprodil in the treatment of traumatic brain injury. [J Neurotrauma](#) 2005; 22: 1428–43.
- [133] [Chen, H.S.](#), [Pellegrini, J.W.](#), [Aggarwal, S.K.](#), [Lei, S.Z.](#), [Warach, S.](#), [Jensen, F.E.](#), [Lipton, S.A.](#), 1992. Open-channel block of N-methyl-D-aspartate (NMDA) responses by memantine: therapeutic advantage against NMDA receptor-mediated neurotoxicity. [J. Neurosci.](#) 12, 4427–4436.

- [134] Block, F., Schwarz, M., 1996. Memantine reduces functional and morphological consequences induced by global ischemia in rats. *Neurosci. Lett.* 208, 41–44.
- [135] Rojas C, Frazier ST, Flanary J, Slusher BS . "Kinetics and inhibition of glutamate carboxypeptidase II using a microplate assay". *Anal. Biochem.* 310 (1): 50–4.
- [136] Mesters JR, et al. Structure of glutamate carboxypeptidase II, a drug target in neuronal damage and prostate cancer *EMBO J.* 25 (6): 1375–84.
- [137] Zhou J, Neale JH, Pomper MG, Kozikowski AP (December 2005). "NAAG Peptidase inhibitors and their potential for diagnosis and therapy". *Nat Rev Drug Discov* 4 (12): 1015–26.
- [138] [Tortella FC](#), et al. Neuroprotection produced by the NAALADase inhibitor 2-PMPA in rat cerebellar neurons. [Eur J Pharmacol.](#) 2000;18;402(1-2):31-7.
- [139] [J P van der Post](#) JP, et al. The central nervous system effects, pharmacokinetics and safety of the NAALADase-inhibitor GPI 5693. *Br J Clin Pharmacol.* 2005 ; 60(2): 128–136.
- [140] Bai F, Bergeron M, Nelson DL. Chronic AMPA receptor potentiator (LY451646) treatment increases cell proliferation in adult rat hippocampus. *Neuropharmacology* 2003;44:1013–1021.
- [141] Voss OP, Milne S, Sharkey J, O'Neill MJ, McCulloch J. Molecular mechanisms of neurite growth with AMPA receptor potentiation. *Neuropharmacology* 2007;52:590–597.
- [142] [Nonpsychotropic cannabinoid acts as a functional N-methyl-D-aspartate receptor blocker](#)". Proceedings of the National Academy of Sciences of the United States of America 86 (23): 9584–7.
- [143] "Development of HU-211 as a neuroprotectant for ischemic brain damage". *Neurological Research* 17 (4): 275–80.
- [144] "Efficacy and safety of dexanabinol in severe traumatic brain injury: results of a phase III randomised, placebo-controlled, clinical trial". *Lancet Neurol* 5 (1): 38–45.
- [145] Brines, M., & Cerami, A. (2006). Discovering erythropoietin's extra-hematopoietic functions: biology and clinical promise. *Kidney Int* 70, 246–250.
- [146] Minnerup, J., Heidrich, J., Rogalewski, A., Schäbitz, W. R., and Wellmann, J. (2009). The efficacy of erythropoietin and its analogues in animal stroke models: a metaanalysis. *Stroke* 40, 3113–3120.
- [147] Mammis, A., McIntosh, T. K., & Maniker, A. H. (2009). Erythropoietin as a neuroprotective agent in traumatic brain injury *Review. Surg Neurol* 71, 527–531.
- [148] Yatsiv, I., Grigoriadis, N., Simeonidou, C., Stahel, P. F., Schmidt, O. I., Alexandrovitch, A. G., et al. (2005). Erythropoietin is neuroprotective, improves functional recovery, and reduces neuronal apoptosis and inflammation in a rodent model of experimental closed head injury. *FASEB J* 19, 1701–1703.
- [149] Ozisik, K., Ozisik, P., Yildirim, E., Misirlioglu, M., & Tuncer, S. (2006). Expression of antiapoptotic survivin and aven genes in rat heart tissue after traumatic brain injury. *Transplant Proc* 38, 2784–2787
- [150] Talving, P., Lustenberger, T., Kobayashi, L., Inaba, K., Barmparas, G., Schnüriger, B., et al. (2010). Erythropoiesis stimulating agent administration improves survival after severe traumatic brain injury: a matched case control study. *Ann Surg* 251, 1–4.
- [151] Corwin, H. L., Gettinger, A., Pearl, R. G., Fink, M. P., Levy, M. M., Shapiro, M. J., et al. (2002). EPO Critical Care Trials Group. Efficacy of recombinant human erythropoietin in critically ill patients: a randomized controlled trial. *JAMA* 288, 2827–2835.
- [152] Xiong, Y., Mahmood, A., Lu, D., Qu, C., Kazmi, H., Goussev, A., et al. (2008). Histological and functional outcomes after traumatic brain injury in mice null for the erythropoietin receptor in the central nervous system. *Brain Res* 1230, 247–257.
- [153] Schoeler M. et al. Dexmedetomidine is neuroprotective in an in vitro model for traumatic brain injury. *BMC Neurology* 2012, 12:20.
- [154] [Safety of Dexmedetomidine in Severe Traumatic Brain Injury](#). [Clinicaltrials.gov](#). 2010
- [155] Gozes I, et al. NAP: research and development of a peptide derived from activity-dependent neuroprotective protein (ADNP). *CNS Drug Rev* 11:353–368.
- [156] Davunetide (AL-108) in Predicted Tauopathies - Pilot Study. [ClinicalTrials.gov](#) Identifier: NCT01056965.
- [157] Xilouri, M. et al. (2007) Neuroprotective effects of steroid analogues on P19-N neurons. *Neurochem. Int.* 50, 660–670
- [158] Stein, D.G. et al. (2008) Does progesterone have neuroprotective properties? *Ann. Emerg. Med.* 51, 164–172
- [159] Wright, D. W., Kellermann, A. L., Hertzberg, V. S., Clark, P. L., Frankel, M., Goldstein, F. C., et al. (2007). ProTECT: A Randomized Clinical Trial of Progesterone for Acute Traumatic Brain Injury. *Ann Emerg Med* 49, 391–402.
- [160] Behl, C. (2002). Estrogen as a neuroprotective hormone. *Nat Rev Neurosci* 3, 433–442.