

## Evaluation of toxicity in rats of *Ruta chalepensis* L. extracts Growing in Iraq

Mustafa A. K. AL-Taie

Lecturer Department of Biology, College of sciences, university of Baghdad, Baghdad, IRAQ

\*Corresponding author: Mustafa A. K. AL-Taie

---

**Abstract:** Evaluate the acute and chronic toxicity of *Ruta Chalepensis* aerial part aqueous extract used in folk medicine in Iraq in Albino Wistar male and female rats. The study of chronic toxicity at the doses 100, 300 and 600mg/kg body weight in male and female rats for 90 days, recorded no mortality. Treated animals showed a normal weight change compared with control. The parameters of male fertility showed a significant decrease in the weight of testis, epididymis and seminal vesicle as well as a reduction in the number and the motility of spermatozooids at treated groups by the doses 300 and 600 mg/kg body weight compared to control group.

**Key words:** *Ruta Chalepensis* L., acute toxicity, chronic toxicity, testis

---

Date of Submission: 10-10-2018

Date of acceptance: 26-10-2018

---

### I. Introduction

Medicinal plants have important contributions in the healthcare system. Use of herbal medicines represent a long history of human interactions with the environment.

Plants used for traditional medicine contain a wide range of substances that can be used to treat chronic as well as infectious diseases. A number of modern drugs currently in use have been obtained through medicinal plants [1].

Despite the profound therapeutic advantages possessed by some of the medicinal plants, some constituents of medicinal plants have been found to be potentially toxic, mutagenic, carcinogenic and teratogenic [2]. This raises concern about the potential toxic effects resulting from the short-term and long-term use of such medicinal plants.

Therefore, evaluating the toxicity effects of any medicinal plants extracts intended to be used in humans and animals is of greatest significance [3].

*Ruta chalepensis* L. (family: Rutaceae), commonly known as "fringed rue", is a perennial herb (ca. 80 cm tall), its flowers are cymes with 4-5 sepals, 4-5 petals, 8-10 stamens and a superior ovary. *Ruta chalepensis* is a perennial herb that widely distributed in the Mediterranean area and also was introduced in America after the Spanish conquest [2]. It is one of the most frequently used plants for medicinal purposes [3,4]. Oil glands, that are principally present in leaves, give its strong deterrent odors [5,6]. *Ruta chalepensis* pharmacological properties, attributed to the high content of alkaloids [7-9], such as furocoumarins<sup>7</sup>, coumarins<sup>9</sup> and furoquinolone alkaloids [10], flavonoids, phenols, amino acids and saponins found in the leaves and stems of the plant [11]. *Ruta Chalepensis* is used, in the traditional medicine for the treatment of various disorders, as analgesic and antipyretic and for the treatment of rheumatism and mental disorders [1]. Also it has emmenagogue, abortifacient, antihelminthic and spasmolytic effects [12] as well as its potency as anti-inflammatory<sup>13</sup>, antihelminthic<sup>14</sup>, antifungal<sup>15</sup>, antifertility, anticonvulsant and sedative [16,17]. In children, infused *Ruta chalepensis* leave extract has been used for treatment of convulsion and other nervous disorders. In Africa, the aqueous decoction of the leaves is used for the treatment of fever [4]. More than fifty chemical compositions of *Ruta chalepensis* essential oil were studied by many research teams in Iran<sup>18</sup>, Greece<sup>19</sup>, Turkey<sup>20</sup> and India [21,22].

### II. Materials and Methods

#### • Preparation of plant extracts

The aerial parts of the plant material were cleaned with tap water, dried in shade at room temperature for 2 weeks and ground into fine powder using an electric grinder.

#### • Aqueous extract

The aqueous extract was prepared according to the method described by Mbiantcha [23], with some modifications. Briefly, 100g of *Ruta chalepensis* L. powder was mixed with 1L of boiled distilled water (100 °C) at room temperature during 72h, the mixture was filtered using Whatman filter paper n°3 and then evaporated in rotary vacuum evaporator

### 2.3 Animals

Males and females of *Albino Wistar* rats. These animals were kept in the animal house, at a temperature of 25°C and a natural photoperiod cycle. The animals were housed in plastic cages (5 rats per cage) and had free access to standard commercial diet and tap water.

- **Animal grouping and extract administration;**

*Albino Wistar* rats of both sexes weighing between 149 and 200 g were divided into four groups of 10 rats each (5 females and 5 males). The aqueous extract dissolved in distilled water, was administered daily by gavage for 90 days to groups I to IV (doses of 0, 100, 300 and 600 mg/kg, respectively). The animals were observed for signs of toxicity and mortality throughout the experimental period. At the end of the treatment, animals were fasted overnight, but allowed access to water ad libitum. They were subsequently anesthetized with diethyl ether and blood samples were obtained by retro-orbital puncture [24] and collected in two tubes. The first one containing EDTA and it was processed immediately for hematological parameters analysis. The second containing heparin and it was centrifuged at 4000g at 4°C for 15 min to obtain serum (stored at -20°C until analysis). The organs (testes, epididymis and ovaries) were weighed and fixed in 10 % formalin for histopathological examination.

## III. Result and Discussion

### Effect of *Ruta chalepensis* L. aqueous extract on Fertility

#### 3.1 Sperm suspension

The epididymis of each rat was placed in 1 mL ofringer buffer, the spermatozoa were obtained by making small cuts in caudal epididymis and diluted by adding 9 mL ofringer buffer (1/10) and then incubated at 37°C for 10 minutes. The sperm suspension was used for analysis of motility and counts.

#### 3.2 Sperm count and motility

A sample of sperm suspension was taken and the number of sperm counted using a haemocytometer under the light microscope at a magnification of x40. Four squares were counted in triplicate. The count was expressed as 2500/mm<sup>2</sup> of suspension. Sperm motility (%) was also assessed immediately by counting both motile and immotile spermatozoa at the magnification of x40 and calculated by the formula:

**Number of motile sperm x 100/total number of motile and immotile sperm.**

- **Histopathological examination**

The treated and control rats' organs were taken out. They were weighed and examined for the evidence of gross lesions. Similar samples were fixed in 10% formalin solution, dehydrated in graded (70-90%) alcohol, cleared in xylene, and placed and embedded in paraffin wax.

To perform histology of tissues, 5-6 µm sections were prepared using Microtome (Leica, RM 2145). These sections then deparaffinated in xylene, passed through 70% to 90% alcohol, and stained with hematoxylin and eosin (H&E). The slides prepared by this process was observed under light microscopy [25].

- **Chronic toxicity**

In chronic toxicity study, rats were divided into 4 groups of 10 animals (5 female and 5 male). The rats were treated with the aqueous extract of *Ruta chalepensis* L. at three doses (100, 300 and 600 mg/kg of body weight/day) for 90 days, while the control group was given only the distilled water.

No toxicity signs or death were recorded during the 90 consecutive days of treatment via oral route with *Ruta chalepensis* L. aqueous extract at doses of 100, 300 or 600 mg/kg of body weight.

- **Effect of *Ruta chalepensis* L. aqueous extract on rats' body weight**

Changes in body weight are an indicator of adverse effects of drugs and chemicals and it will be significant if the body weight loss is more than 10% from the initial body weight occurred [26,27].

The results of body weight of control and treated rats are presented in figure 1 and figure 2. Statistically, no significant difference was noted in the body weight between the control and any of treated groups (1-4) at any time period.

**Figure (1) changes in body weight of male rats after chronic oral treatment with**

**Figure (2) changes in body weight of male rats after chronic oral treatment with Effect of *Ruta chalepensis* L. aqueous extract on sperm count and motility.**

The sperm count in control group was  $14.21 \pm 2.99 \times 2500$  sperm/mm<sup>3</sup>, in group I  $8.66 \pm 2.22 \times 2500$  sperm/mm<sup>3</sup>, in group II  $5.5 \pm 0.04 \times 2500$  sperm/mm<sup>3</sup> and in group III  $1 \pm 0.00 \times 2500$  sperm/mm<sup>3</sup>. Whereas the motility in control group was  $63.73 \pm 2.27$  (%), in group I  $60.00 \pm 2.09$  (%), in group II  $19.42 \pm 4.25$  (%) and in group III  $0.00 \pm 0.00$  (%) (Table 1). In *Ruta chalepensis* L. aqueous extract treated rats the epididymidal sperm parameters showed evidence of dose dependent toxicity. The sperm count and motility were significantly decreased in group II and group III.

**Table 1. Effect of *Ruta chalepensis* L. aqueous extract on sperm count and motility**

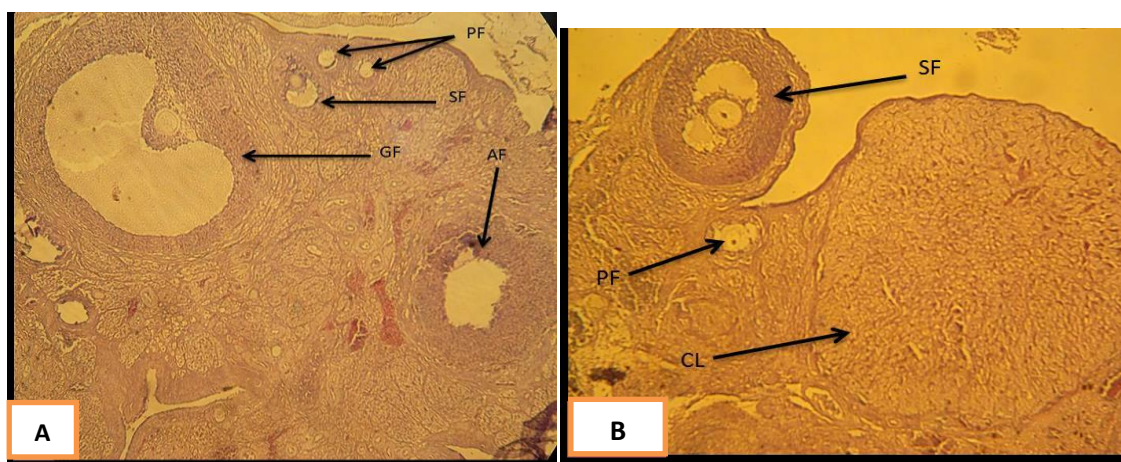
Control	Group I (0 mg/kg)	Group II (100mg/kg)	Group III (300mg/kg)	Group III (600mg/kg)
Count (2500/mm <sup>3</sup> )	$14.21 \pm 2.99$	$8.66 \pm 2.22$	$5.5 \pm 0.04^{**}$	$1 \pm 0.00^{***}$
Motility (%)	$63.77 \pm 2.27$	$60.00 \pm 2.09$	$19.40 \pm 4.25^{***}$	$0.00 \pm 0.00^{***}$

Values are presented as means  $\pm$  SD; \*\* P < 0.01, \*\*\* P < 0.001 compared with control

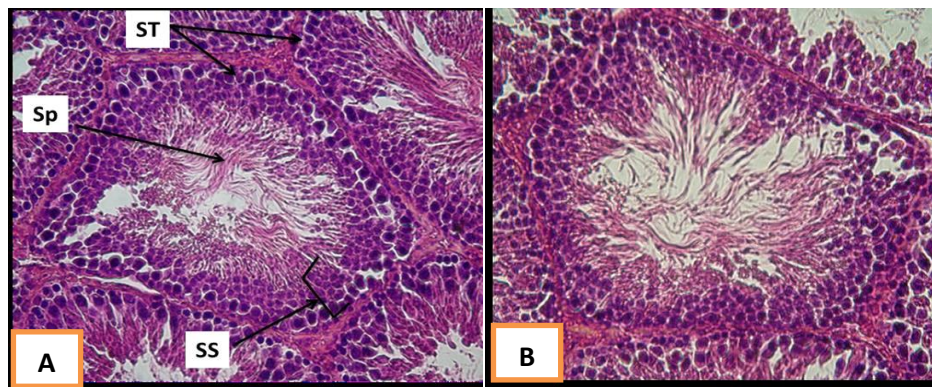
• **Effect of *Ruta chalepensis* L. aqueous extract on rats' organs histology**

there are no histological changes observed in ovary in all treated groups compared with control group (Figure 3).

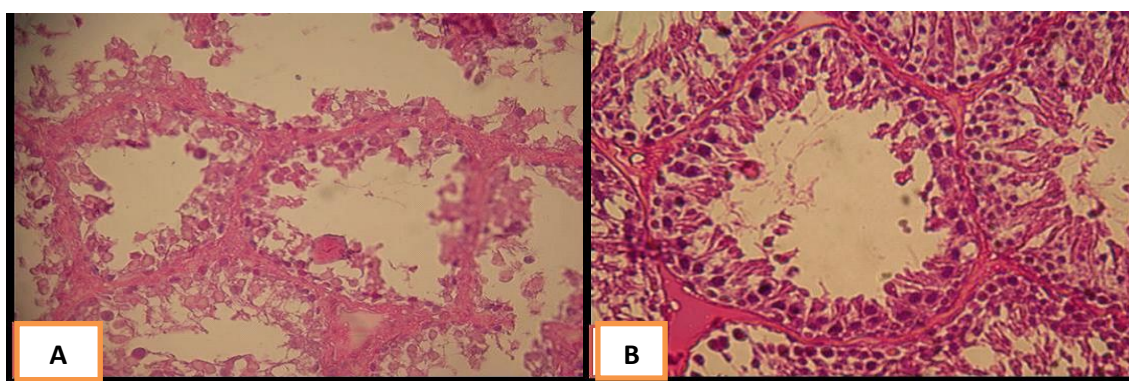
Histological examination of the testes of control group showed normal histological structure of the seminiferous tubules associated with complete spermatogenic series as demonstrated in figure (4 A). The testes of rats given 100 mg/kg of body weight of *Ruta chalepensis* L. aqueous extract had similar histological appearance as the control rats (Figure 4B), and showed active spermatogenesis, however, the testes of the rat treated with 300 and 600 mg/kg of body weight of *Ruta chalepensis* L. revealed degeneration of seminiferous tubules with absence of sperm in tubular lumen as shown in figure (5A) and figure (5B). For the epididymis of control group and the group treated by 100 mg/kg of body weight showed normal tubules that contained spermatozoa in the lumen (Figure 6A and B). Whereas, the treatment with the doses 300 and 600 mg/kg of body weight caused damage in epididymis tubules and absence of sperm (Figure 7A and B).



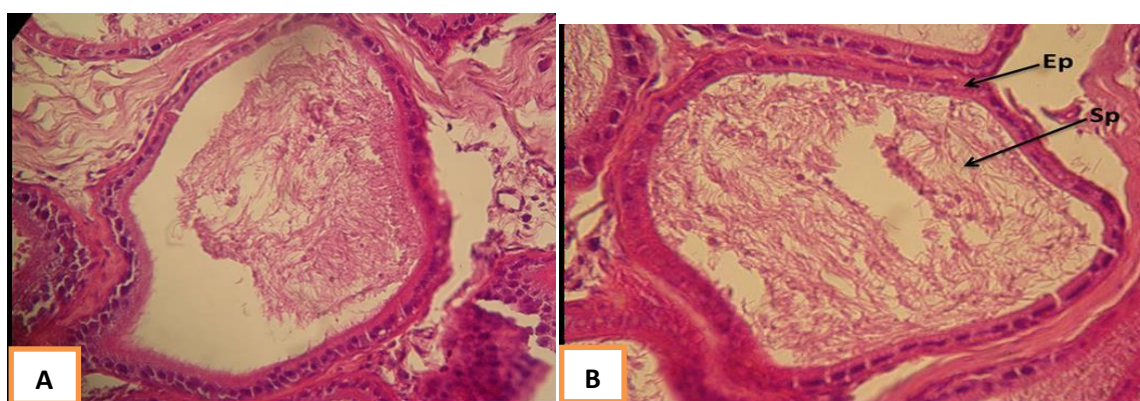
**Figure 3:** Histological sections of Ovary of control group (A) and group 1 (B) treated with 100 mg/kg bw of *Ruta chalepensis* L. aqueous extract after 90 days of treatment showing normal structure. PF: primary follicle ; SF: secondary follicle ; GF: grafian follicle ; atresia follicle ; CL : corpus luteum. (HE X 200)



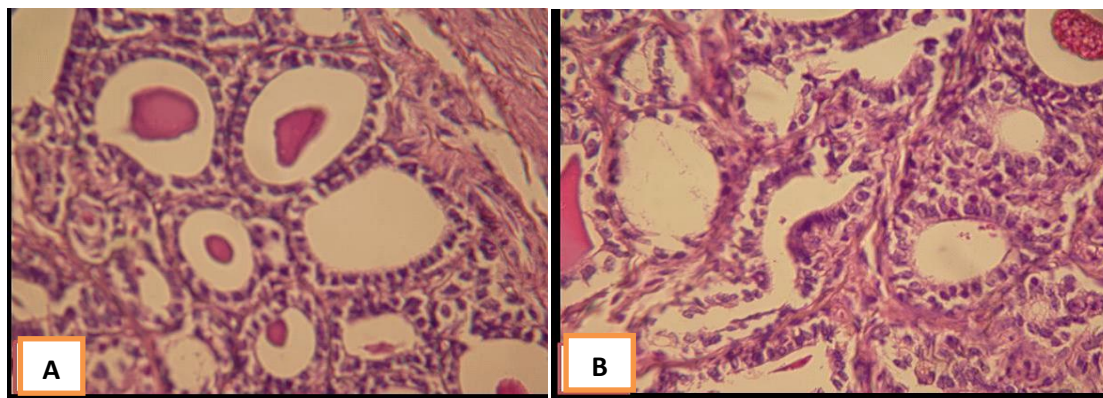
**Figure 4:** Histological sections of testes of control group (A) and group 1 (B) treated with 100 mg/kg bw of *Ruta chalepensis* L. aqueous extract after 90 days of treatment showing normal histological structure of the seminiferous tubules associated with complete spermatogenic series and spermatozoa in the lumen. ST: seminiferous tubules; Sp: sperm; SS: stages of spermatogenesis. (HE X 200)



**Figure 5:** Histological sections of testes of group 2 (A) and group 3 (B) treated with 300 mg/kg and 600 mg/kg bw respectively of *Ruta chalepensis* L. aqueous extract after 90 days of treatment showing degeneration of seminiferous tubules with absence of sperm in tubular lumen. (HE X 200)



**Figure 6:** Histological sections of the epididymis of control group (A) and group 1 (B) treated with 100 mg/kg bw of *Ruta chalepensis* L. aqueous extract after 90 days of treatment showing normal tubules that contained spermatozoa in the lumen. Ep: Epithelial cells; Sp: Sperm. (HE X 200)



**Figure 5:** Histological sections of the epididymis of group 2 (A) and group 3 (B) treated with 300 mg/kg and 600 mg/kg bw respectively of *Ruta chalepensis* L. aqueous extract after 90 days of treatment showing damage in epididymal tubules and absence of sperm. (HE X 200).

#### IV. Discussion

Medicinal plants have been in use all over the world to treat various diseases including inflammation, heart diseases, cancer, etc. Today the large numbers of drugs in use is derived from plants, which are rich in secondary metabolites and essential oils of therapeutic importance. The important advantages claimed for therapeutic uses of medicinal plants in various ailments are their safety besides being economical, effective and their easy availability [28]. In rats receiving the *Ruta chalepensis* L. aqueous extract orally at doses of 100, 300 or 600 mg/kg during 90 consecutive days, there was no significant change in the body weight in the treated groups compared to the control group. Changes in body weight have been used as an indicator of adverse effects of drugs and chemicals [29]. Since, no significant changes were observed in the body weight, in the treated groups compared to that of the control group, it suggested that at the oral doses administered, *Ruta chalepensis* L. aqueous extract had no effect on the growth of rats. Generally, the alterations of body weight gain and internal organ weights of mice would reflect the toxicity after exposure to the toxic substances [30]. Organ weight is an important index of physiological and pathological status in animals. The relative organ weight is fundamental to diagnose whether the organ was exposed to the injury or not [31]. In this study, no significant changes were observed in the internal organ weight after the 90 days' period, except a significant decrease in testis, epididymis and seminal vesicle in groups treated by the doses 300 and 600mg/kg when compared to the control group. In addition, there is no changes were observed in gross observation of organs of both control and treated groups. In the present study, a significant weight reduction was seen in the testes, epididymis and seminal vesicle in the doses 300 and 600 mg/kg when compared to the control group. The weight reduction was dose dependent. These results are similar to those obtained by Khouri and EL-Akawi [32], which suggest that the administration of aqueous extract of *Ruta graveolens* L. at a dose of 500 mg/kg body weight for 60 days induces a significant decrease in the weight of reproductive organs ( $P < 0.01$ ) when compared to control. But, do not agree with the results obtained by Al Qarawi [33] which suggest that the oral administration of an aqueous extract of the leaves of *Ruta chalepensis* in daily oral doses of 0.5 g, 1.0 g and 2.0 g for 30 days caused an increase of the testicular and epididymis weights. Generally, the reductions in internal organ weight are simple and sensitive indices of toxicity after exposure to toxic substances [27]. The significant reduction of testis weight is known to be mostly related to number of spermatids and spermatozoa present in the tissue [34]. The significant reduction in the weight of reproductive organs indirectly support the reduced availability of androgen [35]. It is known that the accessory sex organs are androgen dependent target organs and manifest differential sensibility to androgens for maintenance of their structure and function. It is also known that, any change in circulating androgens would affect the internal microenvironment of epididymis and thereby lead to alteration in sperm motility and metabolism [36]. Androgen deprivation not only suppresses spermatogenesis, leading to low sperm concentration, but also alters the epididymal milieu which renders it hostile for maturation and survival of the spermatozoa [37,38]. Testosterone, an important androgen, plays a pivotal role in maturation, spermatogenesis and the maintenance of 100 accessory sex organs [39]. The structural and functional integrity of reproductive tissues depends on the circulating androgen [40] and therefore, any small change in testosterone content may result in reductions in the weights of the reproductive organs. Sperm characteristics are important reproductive indices as they account for male fecundity. The aqueous extract at the doses of 300 and 600 mg/kg body weight produced a significant reduction in the sperm count and motility. Two possible hypotheses may be proposed to explain this reduction. One hypothesis is that the principles active of the extract may alter the pituitary gonadotropins hormones: luteinizing hormone (LH) and follicle stimulating hormone (FSH) ([41]. It is well known that the weight, size and the Secretary function of testes, epididymis and seminal vesicles are

closely regulated by androgens hormones [42]. The production of the sperm cells (spermatozoa) and testosterone in the testis are mainly regulated by the follicle stimulating hormone and Luteinizing hormone, which are released from the anterior pituitary [43]. FSH stimulates spermatogenesis in the Sertoli cells, while LH stimulates the production of testosterone in Leydig cells of the testis [44]. Low levels of these hormones decrease endogenous testosterone secretion from the testis depriving developing sperm of the signal required for normal maturation and also it suppresses testicular steroidogenesis and spermatogenesis [45] since the pituitary-testicular axis is a central regulatory conduit for testicular function that culminates in the production of spermatozoa [46]. Besides hormonal alteration, the alternative hypothesis is that the principles active may induce oxidative stress in testicular tissue and stored germ cells leading to generation of free radical products, as they exert a detrimental effect on spermatogenesis [47]. The histological examination is the golden standard for evaluating treatment related pathological changes in tissues and organs [48]. In the present study, histopathological evaluation of chronic oral administration of *Ruta chalepensis* L. indicated that the aqueous extract did not cause toxicity towards the organs as there was no structural damage to the organs of ovary of the rats.

## References

- [1]. Iauk L., Mangano K., Rapisarda A., Ragusa S., Maiolino L., Musumeci R., Costanzo R., Serra A., 2004: Berberis-aetnensis C. persl. Extracts: antimicrobial properties and interaction with ciprofloxacin. *J. Ethno pharmacology*, ,90, 267-272.
- [2]. Zeichen R., Rey A., Arganara E., Bindstein E. 2000.: Perinatal toxicology of *Ruta chalepensis* (Rutaceae) in mice. *J. Ethno pharmacology*, 69(2), 93-8.
- [3]. Arenas P., G.P. Savitry. 1994 *La ruda: Ruta chalepensis* (L.) Rutaceae. *Dominguezia*. 11:7-25.
- [4]. AlSaid M.S., Tariq M., AlYahya M.A., Rafatullah S., Ginnawi O.T., Ageel A.M. 1990.: The role of ethno pharmacology in drug development. *J. Ethno pharmacology*, 28,305-312.
- [5]. Cabrera A., and Zardini E. 1978. *Manual de la flora de losAlrededores de Beunos Aires* (Ed) Acme Buenos Aires, Argentina, , p. 234.
- [6]. Trease G.E. and Evans W.Ch. 1980. *Pharmacognosy*, Baillièere Tindal Press, London, , p. 488.
- [7]. Günaydin K. and Savci S. 2005. Phytochemical studies on *Ruta chalepensis*(LAM.) lamarck*Nat. Prod. Res.*, , 19, 203-210.
- [8]. Ulubelen, A. and H. Guner. 1988. Isolation of dehydromoskachen C from *Ruta chalepensis* var. *Latifolia*. *J. Nat. Prod.* 51: 1012-1013.
- [9]. Mohr N., Budzikiewicz H., El-Tawil B. A. H. and El-Beih F. K. A., 1982. Further furoquinolone alkaloids from *Ruta chalepensis*. *Phytochemistry*, ,21 (7), 1838-1839.
- [10]. HnatyszynO., Arenas P., Moreno A.R., Rondina R., Coussio J.D. 1974. *PlantasRevista de la SociedadCientifica*, , p. 23.
- [11]. Di-Stasi L.C., Hiruma C.A., Guimaraes C.M. 1994. Medical plants popularly used in Brazilian Amazon. *Fitoterapia*, ,65, 529-540.
- [12]. Atta A. H., Alkofahi A. Anti-nociceptive and anti-inflammatory effects of some Jordanian medicinal plant extracts. *Journal of Ethnopharmacology*. 1998;60(2):117-124.
- [13]. Calzada F., YépezMulia L., Aguilar A. 2006.: In vitro susceptibility of *Entamoebahistoltyca* and *Giardia lamblia* to plants used in Mexican traditional medicine for the treatment of gastrointestinal disorders..*J. Ethno pharmacology*, ,108, 367-370.
- [14]. Ali-Shtayeh M.S., Abu-Ghdeib S.I. 1999.: Antifungal activity of plant extracts against dermatophytes. *Mycoses*, ,42, 665-672.
- [15]. Ulubelen A., Ertugrul L., Birman H., Yigit R., Erseven G., Olgac V. 1993.: Antifungal activity of plant extracts against dermatophytes. *Phytotherapy Res.*, 8, 233-236.
- [16]. Aguilar-Santamaria, L. and Tortoriello, J. Anticonvulsant and sedative effects of crude extracts of *Ternstroemiapringlei* and *Ruta chalepensis*. *Phytotherapy Research* 1996; 10: 531-533.
- [17]. Rustaiyan A, Khossravi M, Sultani-Lotfabadi F, Yari M, Masoudi S, Monfared A (2002). Constituents of the essential oil of *Ruta chalepensis* L. from Iran. *J. Essent. Oil Res.*, 14: 378-379.
- [18]. Tzakou O.\*, Couladis M. 2001. The essential oil of *Micromeriagraeca* (L.) Bentham et Reichenb. growing in Greece. *Flav. Fragr. J.*, 16, 107-109.
- [19]. Baser, K.H.C., T. Ozek and S.H. Beis, 1996. Constituents of the essential oil of *Ruta chalepensis* L. from Turkey. *J. Essent. Oil Res.*,8: 413-414.
- [20]. Bagchi G.D. Dwivedi P.D., Mandal S., Naqvi A.A., Kumar S. 2003.: Essential oil constituents of *Ruta chalepensis* plants grown in India. *Indian Perfumer*, 47, 39-41.
- [21]. Bagchi, G.D., P.D. Dwivedi, A. Singh, F. Haider and A.A. Naqvi, 2003. Variations in essential oil constituents at different growth stages of *Ruta chalepensis* on cultivation at north Indian plains. *J. Essent. Oil Res.*, 15: 263-264.
- [22]. **Mbiantcha, M., Kamanyi, A., Teponno, R.B., Tapondjou, L.A., Watcho, P. and Nguelafack, T.B.** (2011). Analgesic and Anti-inflammatory Properties of Extracts fromthe Bulbils of *Dioscorea bulbifera* L. var *sativa* (Dioscoreaceae) in Mice and Rats.Evidence-Based Complementary and Alternative Medicine, 1-9.
- [23]. **Waynforth, H. B.** (1980). *Experimental and surgical technique in the rat*. London.Academic Press, 17-68.
- [24]. **Bensalem-bendjelloul, M.** (1998). *Techniques histologiques*théorie et pratique. Office des publications universitaires, 6-41.
- [25]. **Raza, M., Al-Shabanah, O.A., El-Hadiyah, T.M., and Al-Majed, A.A.** (2002). Effect of prolonged vigabatrin treatment on haematological and biochemical parameters in plasma, liver and kidney of Swiss albino mice. *ScientiaPharmaceutica*, 70, 135-145.
- [26]. **Teo, S., Stirling, D., Thomas, S., Hoberman, A., Kiorpes, A., and Khetani, V.** (2002).A 90- day oral gavage toxicity study of D-methylphenidate and D, Lmethylphenidatein Sprague-Dawley rats. *Toxicology*, 179, 183-196.
- [27]. **Verma, S. K., AND Kumar, A.** (2011). Therapeutic uses of withaniasomnifera(Ashwagandha) with a note on withanolides and its pharmacological actions. *AsianJournal of Pharmaceutical and Clinical Research*, 4, 1-4.
- [28]. **El Hilaly, J., Israili, Z.H., and Lyoussi, B.** (2004). Acute and chronic toxicological studies of *Ajugaivain* experimental animals. *Journal of Ethnopharmacology*, 91, 43-50.
- [29]. **Carol, S.A.** (1995). *Acute, Sub chronic and Chronic Toxicology*. In *CRC Handbook of Toxicology*; Michael, J.D., Mannfred, A.H., Eds.; CRC Press Inc.: Boca Raton, FL, USA,;pp. 51-104.
- [30]. **Dybing, E., Doe, J., Groten, J., Kleiner, J., and O'Brien, J.** (2002). Hazard characterization of chemicals in food and diet: dose response, mechanism andextrapolation issues. *Food Chem. Toxicol*,42, 237-282.
- [31]. **Khouri, N.A., and EL-Akawi, Z.** (2005). Antiandrogenic activity of *Ruta graveolens*L in male Albino rats with emphasis on sexual and aggressive behavior. *NeuroendocrinologyLetters*. 26(6), 823-829.

- [32]. **Al Qarawi, A.A.** (2005). Stimulatory Effect of the Aqueous Extract of *Ruta chalepensis* on the Sex Organs and Hormones of Male Rats. *The Journal of Applied Research*, 206-211.
- [33]. **Gupta, R.S., Chaudhary, R., Yadav, R.K., Verma, S.K. and Dobhal, M.P.** (2005). Effects of saponins of *Albizialebeck (L.) benth* bark on the reproductive system of male albino rats. *J. Ethnopharmacol*, 96, 31-36.
- [34]. **Zeherea, M .N., Reddy, S. P., Reddy, S. P., Ravindra., and Saraswati, B. Patil.** (1998) Antispermatic and androgenic activities of *Momordica Charantia* (Karela) in Albino rats. *J. Ethnopharmacol*. 61, 9- 16.
- [35]. **Khan, P.K., and Awasthy, K.S.** (2003). Cytogenetic toxicity of Neem. *Food chem.Toxicol*, 41, 1325-1328.
- [36]. **Setty, B.S.** (1979). Regulation of Epididymal function and sperm maturation- endocrine approach to fertility control in male. *Endocrinology*. 74, 100-117.
- [37]. **Rao, M.V.** (1988). Effect of alcoholic extract of *Solanum xanthocarpum* seeds in adult male rats. *Indian J. Exp.Biol*. 26, 95-98.
- [38]. **Keel, A. B., and Abney, T.** (1980). Influence of bilateral cryptorchidism in the mature rat; Alteration in testicular function and serum hormonal level. *Endocrinology*, 107, 1226-1233.
- [39]. **Chinoy, N.J., Sheth, K.M., and Seethalakshmi, L.** (1982). Studies on reproductive physiology of animals with special reference to fertility control. *Comp. Physiol. Ecol*, 7- 325.
- [40]. **Kusemiju, T.O., Osinubi, A.A., Noronha, C.C., Okanlawon, A.O.** (2010). Effect of *Carica papaya* bark extract on the testicular histology of male SD rats. *Nig. Quat. Jour. Of Hosp. Med*, 20 (3), 133-137.
- [41]. **Almasad, M.M., Qazan, W.S., and Daradka, H.** (2007). Reproductive toxic effects of *Artemisia herba alba* ingestion in female Spague-Dawley rats. *Pak J Biol Sci*. 10(18), 3158-3161.
- [42]. **Steinberger, E.** (1971). Hormonal control of mammalian Spermatogenesis. *Physiology Review*, 51(1), 1-22.
- [43]. **Wangenstein, H., Samuelsen, A.B., and Maltrud, K.E.** (2004). Antioxidant activity in extracts from coriander. *Food Chem*, 88, 293-297.
- [44]. **Kusemiju, T.O., Osinubi, A.A., Noronha, C.C., Okanlawon, A.O.** (2010). Effect of *Carica papaya* bark extract on the testicular histology of male SD rats. *Nig. Quat. Jour. Of Hosp. Med*, 20 (3), 133-137.
- [45]. **Cheng, C.Y., Wong, E.W., Yan, H.H., and Mruk, D.D.** (2010). Regulation of spermatogenesis in the microenvironment of the seminiferous epithelium: new insights and advances. *Mol. Cell Endocrinol*, 315(1-2), 49-56.
- [46]. **Ghosh, D., Das, U.B., Ghosh, S., Mallick, M., and Debnath, J.** (2002). Testicular gametogenic and steroidogenic activities in cyclophosphamide treated rats: A correlative study with testicular oxidative stress. *Drug ChemTox*, 25, 281-291.
- [47]. **Subramanion, L. J., Zuraini, Z., Yeng, C., Yee, L. L., Lachimanan, Y. L. and Sreenivasan, S.** (2011). Acute Oral Toxicity of Methanolic Seed Extract of *Cassia fistulain* Mice. *Molecules*, 16, 5268-5282.

Mustafa A. K. AL-Taie. "Evaluation of toxicity in rats of *Ruta chalepensis* L. extracts Growing in Iraq."  
*.IOSR Journal of Pharmacy and Biological Sciences (IOSR-JPBS)* 13.5 (2018): 30-36