

Phytochemical Screening, Analgesic And Anti-Inflammatory Activities Of Methanol Stem Bark Extract Of *Senna Siamea* Lam. (Kassod Tree)

Sodipo O. A.,¹ Tijjani., M.A., Yakubu, J.,² Abdulrahman, F. I.² And Khan, I. Z.²

¹Department of Clinical Pharmacology and Therapeutics, College of Medical Sciences, University of Maiduguri.

²Department of Chemistry, Faculty of Science, University of Maiduguri, P.M.B 1069, Maiduguri, Borno State, Nigeria.

Corresponding Author' Sodipo O. A

Abstract

The study aims at phytochemical screening, analgesic and anti-inflammatory activities of methanol stem bark extract of *Senna siamea* Lam. (kassod tree). Fresh stem bark of *Senna siamea* were air-dried, pulverized extracted using maceration method of extraction technique with methanol and yielded 12.60 % w/w after being concentrated. The extract was screened for phytochemicals using standard methods. The phytochemical studies of the methanol extract of *Senna siamea* revealed the presence some chemical compounds such as alkaloid, flavonoids, cardiac glycosides, tannins, saponins, and terpenoids. The analgesic effect of the leaf extract was evaluated with acetic acid induced writhing and thermally induced nociception for pain while the anti-inflammatory effect was evaluated using albumin-induced rat paw oedema model. The LD₅₀ of the stem bark extract was ≥ 5000 mg/kg. The methanol stem bark extract of *Senna siamea* caused an inhibition on the writhing response induced by acetic acid in a dose dependently. Similarly, the extract doses increased the time of tail flicking in a dose dependent manner. The stem bark extract also significantly ($P < 0.05$) inhibited inflammation induced by egg albumin in the rats paw. Thus, this study has scientifically justified that the plant possess some degree of action on peripheral and central nervous system thereby acting as an antidepressant in suppressing pain and inflammation. The proves the use of the plant locally for the management and treatment of pain related health problems.

Keywords: *Senna siamea*, analgesic, bioactive, phytochemicals

Date of Submission: 20-06-2018

Date of acceptance: 05-07-2018

I. Introduction

For centuries, natural products have provided medicine for human illness and most of these remedies were obtained from higher plants (Wink, 1999). Natural products have been an integral part of the ancient traditional medicine systems, for example Chinese, Ayurvedic and Egyptian (Sarker and Nahar, 2007). The use of medicinal plants in West Africa is probably as old as the duration of human settlement in the region (Abdulrahman *et al.*, 2010; Sodipo *et al.*, 2011). The reason for the use of herbs is because of their affordability, easy accessibility and effectiveness. In the last two centuries, there has being serious investigations into the chemical and biological activities of plants and these have yielded compounds for the development of synthetic organic chemistry and the emergence of medicinal chemistry as a route for the discovery of more effective therapeutic agents (Roja and Rao, 2000).

According to the World Health Organization (WHO), a medicinal plant is any plant which, in one or more of its organs, contains substances that can be used for therapeutic purposes, or which are precursors for chemo-pharmaceutical semi synthesis. Such a plant will have its parts including leaves, roots, rhizomes, stems, barks, flowers, fruits, grains or seeds, employed in the control or treatment of a disease condition and therefore contains chemical components that are medically active (Liu, 2004). The availability, low cost and accessibility of these plants in Tropical and Sub-tropical Africa coupled with the global crisis of drug resistance incidences make it convenient for in-depth survey of medicinal plants from this part of the world (Usman *et al.*, 2009).

Senna (from Arabic *sanā*), the *sennas*, is a large genus of flowering plants in the legume family Fabaceae, and the subfamily Caesalpinioideae. This diverse genus is native throughout the tropics, with a small number of species in temperate regions. The number of species is estimated to be from about 260 (Marazzi *et al.*, 2006) to 350 (Randell and Barlow, 1998).

The leaves, stems, roots, flowers and seeds of *S. siamea* regardless of the subspecies have been used for the treatment of several illnesses including mostly malaria (Koudouvo *et al.*, 2011). According to the ethnic differences of populations from localities, the plant is used alone or in combination with other plants or with natural substances for preparation, especially in decoction (Maurya and Dongarwar, 2012).

In Burkina Faso, Ghana and Nigeria, the decoction of the whole stem or stem bark is taken or used for body bath against malaria and liver disorders (Adebayo and Krettli, 2011). These same uses were reported in Malaysia (Al-Adhroey *et al.*, 2010). The dried stems of *C siamea* mixed with the fruit of *Xylopi aethiopica* are pulverized and administered as a laxative (Kiepe, 1995). The decoction of the stem bark is used to treat diabetes. It is also used as a mild, pleasant, safe and purgative in Japan. Dalziel (1963); Odason and Kolawole, (2007) also indicated that this decoction is also used for scabies, urogenital diseases, herpes and rhinitis in Cambodia. In spite of the global advancement in discovery of drugs, conventional drugs still remain a major concern due to negative scientific reports regarding their adverse effects. More so, the popularity of this therapy among the healthcare workers and the general public, it is still not known whether the benefits of analgesic and anti-inflammatory therapy outweigh its risks. This has necessitated the search for a safer, affordable, available and assessable means of treatments within the plant kingdom. Thus, this study aims at screening for phytoconstituents responsible for the folkloric use of *Senna siamea* for the treatment and management of pain and inflammation.

II. Materials And Methods

Plant Extraction

One (1) kilogramme of the pulverized stem bark of *Senna siamea* was extracted exhaustively by maceration method of extraction using methanol. The crude extract was concentrated to dryness at reduced pressure in a vacuum using a rotary evaporator at 40° C. The extract was weighed, labeled and subjected to further analysis.

Preliminary Phytochemical Screening

The extract fraction of the stem bark was screened qualitatively for phytochemical constituents using standard procedures (Brain and Turner, 1975; Vishnoi, 1979; Markham, 1987; Silver *et al.*, 1998; Sofowora, 2008; Evans, 2009).

Pharmacological Investigations of the Methanol Stem Bark Extract of *Senna siamea*

All the experiments performed on laboratory animals in this study followed the standard procedure for the treatment of animals. The animals were handled according to the International Guiding Principle for Biomedical Research involving animals, (CIOMS and ICLAS, 2012).

A total of one hundred and forty eight (74) albino rats (100-180 g) and fifty (25) mice (20-28 g) of both sexes were purchased from the Animal House of the Faculty of Pharmacy, University of Maiduguri, Borno State. They were housed in clean plastic, well-ventilated cages with saw dust as beddings under 12 hours light/12 hours dark cycle conditions of normal room temperature and humidity in the Pharmacology, Physiology and Biochemistry Laboratory, Faculty of Veterinary Medicine, University of Maiduguri for the analysis. They were fed with standard feed and allowed water *ad libitum*.

Acute Toxicity Evaluation (LD₅₀)

The acute toxicity (LD₅₀) of the crude stem bark extract of methanol were determined using standard conventional procedure as described by Lorke (1983). In this study, two different routes of administration were considered; the oral and intraperitoneal. In phase I, rats were divided into 3 groups of three rats each for each route (a total of nine rats) and then treated with the crude methanol extract at doses of 10, 100 and 1000 mg/kg bd. wt. intraperitoneally and orally and observed for 24 hours for mortality. In the phase II, the animals of each group (for each route) were divided into three groups of one animal each and the methanol extract was administered at doses that were determined after the phase I. The rats were observed for signs of toxicity and mortality for the first critical four hours and thereafter daily for 7 days. The LD₅₀ was then calculated using the formula:

$$LD_{50} = \sqrt{a \times b}$$

Where a = least dose that killed a rat

b = highest dose that did not kill a rat

Analgesic Evaluation

Effect of Extract on Acetic Acid-Induced Writhing on Mice

The abdominal constriction resulting from intraperitoneal injection of acetic acid (0.6% v/v) consisting of a contraction of abdominal muscle, together with a stretching of hind limbs, was carried out according to the procedure described by Abdulrahman (2004); Correa *et al.* (1996); Nwafor (1998); Santos *et al.* (1994). Twenty (25) mice were divided into 5 groups of 5 mice each. Groups 1 and 5 served as the negative and positive controls respectively, while groups 2, 3 and 4 were pretreated (*ip*) with doses of 100, 200 and 300 mg/kg. b. wt. of the extract (*ip*). 30 minutes later, acetic acid (0.6% v/v) was administered. The number of writhing movements was counted for 30 minutes. Antinociception was expressed as the reduction of the number of abdominal constriction between negative control mice (distilled water treated mice), mice pretreated with the extract and the positive control (10 mg/kg pentazocine treated mice) and was calculated using the formula:

$$\% \text{ Protection} = \frac{(\text{Mean no. of writhes in Control group} - \text{Mean no. of writhes in Test group}) \times 100}{\text{Mean no. of writhes in Control group}}$$

Tail Immersion

Method described by Owoleye *et al.* (2004) was adopted. Rats were treated intraperitoneally with 200, 400 and 600 mg/kg of the extracts, distilled water and 10mg/kg, pentazocine (10 mg/kg) served as the negative control and positive control respectively. Measurements of extract effect were carried out within time intervals of 30, 60, 90 and 120 min after administration of the extracts. Water was heated to 50.0 ± 1.0 °C in a water bath. The time taken for the animal to remove its tails out of the water was recorded.

The increase in pain threshold was calculated using the formula:

$$\% \text{ Increase in pain threshold} =$$

$$\frac{(\text{Mean reaction time in test group} - \text{Mean reaction time in control group}) \times 100}{\text{Mean reaction time in test group}}$$

Anti-inflammatory Studies

Albumin-Induced Rat Paw Oedema Model

The anti-inflammatory study was carried out using the method described by Winter *et al.* (1963). 25 rats were divided into five groups, 1 and 2 serving as negative control (distilled water 10 ml/kg) and positive control (Pentazocine, 10 mg/kg), while groups 3, 4 and 5 received 200 mg/kg, 400 mg/kg, and 800 mg/kg of the extract respectively. Treatments were administered 1 hour before albumin injection. Albumin was separated from the yolk and was injected underneath the planter region of the paws of the rats. The paw size was measured with a digital vernier calliper at 0, 1, 2, 3, 4, 5 and 6 hours after albumin injection.

$$\% \text{ Increase in pain threshold} =$$

$$\frac{(\text{Mean reaction time in test group} - \text{Mean reaction time in control group}) \times 100}{\text{Mean reaction time in test group}}$$

Statistical Analysis

Data generated during the study were expressed in mean \pm standard Error of mean (SEM) and analysed by one way analysis of variance (ANOVA) Using InStat Graphpad version 3.10 (Graphpad InStat, 2000). Values of $P < 0.05$ were considered significant at 95 % confidence level.

III. Results And Discussion

Plant Extraction

The extraction of the stem of *Senna siamea* using methanol produced extract with greenish brown colours which was powdery. The methanol extract had a yield of 12.39% . The result of the extraction profile is shown on Table 1:

Phytochemical Screening of the Stem Bark Extract

The preliminary phytochemical screening of the stem bark using methanol as solvents revealed the presence of some phytochemicals such as flavonoids, terpenoids, cardiac glycosides, saponins, tannins and flavonoids. The result of the phytochemical screening of the gradient extraction is shown in Table 2:

Table 1: The extraction profile of air dried powdered stem bark of *Senna siamea*

Extract	Mass (g)	% Recovery (^w / _w)	Colour	Texture
Ethanol stem bark extract	61.93	12.39	greenish brown	powdery

Table 2: Results of Phytochemical Screening of Stem Bark Extract of *Senna siamea*

S/N	PHYTOCHEMICAL TEST	SSMSE
1	Test Tor Carbohydrates	
i	General test-Molish	+
ii	Test for reducing sugar-fehling test	+
iii	Test for combined reducing sugar	+
iv	Test for ketoses	+
v	Test for pentoses	+
2	Test for Tannins	
i	Ferric chloride test	+
ii	Lead acetate	+
3	Test for Phlobatannins	-
4	Test for Steroids/Triterpenes	
i	Salkowski test	+
ii	Liebermann-burcharde test	+
5	Test for Flavonoids	
i	Shinoda's test	+
ii	Ferric chloride test	+
iii	Lead acetate test	+
6	Test for Saponins	
i	Frothing test	+
7	Test for Soluble Starch	-
8	Test for Alkaloids	
i	Dragendroff's reagent	+
ii	Meyer's reagent	+
9	Test for Steroidal Nucleus	
i	Keller- killiani's test	+
10	Test for Terpenoids	+

SSESE- *Senna siamea* methanol stem bark extract;

Acute Toxicity (LD₅₀)

Tables 3 present the result of acute toxicity of the methanol stem bark extract of *Senna siamea* on rats. No death was recorded on administration of up to 5000 mg/kg dose of the methanol extract via both the oral and intraperitoneal routes. Thus LD₅₀ of the crude methanol extract of *Senna siamea* on rats administered via both oral and intraperitoneal routes was ≥ 5000 mg/kg bd. wt.

Analgesic Effect of Methanol stem bark Extract of *Senna siamea*

Acetic Acid-Induced Writhing

The methanol stem bark extract of *Senna siamea* also exerted an inhibition on the writhing response induced by acetic acid in a dose dependent manner at ($P < 0.05$) [Table 4]. 32.80 ± 0.37 , 27.80 ± 0.37 and 22.00 ± 2.17 mean number of writhing for doses of 100, 200 and 300 mg/Kg bd. wt.(i,p) was observed as compared to the reference drug(positive control) (18.60 ± 0.51) as shown in table (4). The effect was more pronounced at a high dose of 300mg/kg bd. wt. which gave a high percentage of inhibition (64%) of the abdominal constriction induced by acetic acid. This was found to be significantly lower than the effect of the synthetic drug (pentazocine, 20 mg/kg bd. wt) and significantly higher than animals treated with distilled water with mean number of writhes at 21.40 ± 1.28 and 62.00 ± 0.70 in the extent to which the writhing or stretching induced by acetic acid was reduced.

Thermally-Induced Nociception (Tail Immersion Test)

Figure 4 represent the mean time of tail flick at increasing doses of methanol stem bark extract of *Senna siamea* in the evaluation of thermally induced nociception of ethanol extract on rats. The extract doses of 200, 400 and 600 mg/kg bd. wt. significantly ($p < 0.05$) increased the time of tail flicking. The extract is observed to be more effective at 60 minutes after administration in a dose dependent manner (6.20 ± 0.20 , 6.20 ± 0.20 , 207.00 ± 0.54 at doses of 200, 400 and 600 mg/kg respectively). However pentazocine significantly increased the time of tail flick with a superior effect when compared to the extract.

Anti-inflammatory Effect

The methanol stem bark extract of *Senna siamea* (200, 400 and 800 mg/kg) caused statistically significant ($P < 0.05$) inhibition of inflammation induced by egg albumin in the rats paw with decrease in diameter of 4.92 ± 0.25 , 4.32 ± 0.11 and 3.80 ± 0.17 respectively. The percentage inhibition of the inflammation caused by the extract was comparable to that obtained with Pentazocine (20 mg/kg) which was used as standard (Figure 1). The effect of the stem bark extract was also dose-dependent.

Table 3: Acute toxicity effect of methanol stem bark extract of *Senna siamea* on rats

Phase	Dose (mg/kg)	No. of rat	Mortality rate	
Oral route	IP route			
I	10	3	0/3	0/3
	100	3	0/3	0/3
	1000	3	0/3	0/3
II	1600	1	0/1	0/1
	2900	1	0/1	0/1
	5000	1	0/1	0/1

LD₅₀ ≥ 5000 mg/kg

Table 4: Effect of methanol stem bark extract of *Senna siamea* on acetic acid induced writhes in mice

Group	Treatment (mg/kg)	No. of Writhes mean±S.E.M	% Protection
A	H ₂ O (-ve control)	62.00±0.70	0
B	100	32.80±0.37	47
C	200	27.80±0.37	55
D	300	22.00±2.17	64
E	10 pentazocine (+ve control)	21.40±1.28	65

Values across column with same superscript are statistically ($p > 0.05$) not significant

Values across column with no or/different superscript are statistically ($p > 0.05$) significant

Table 5: Analgesic effect of *Senna siamea* stem bark extract on rats (Tail Immersion Method)

Group	Treatment (mg/kg)	Mean±S.E.M tail flick (min)			
		30	60	90	120
A	H ₂ O (-ve control)	4.80±0.20	4.80±0.20 ^b	4.60±0.24 ^a	4.20±0.20 ^a
B	200	6.20±0.20 ^b	6.20±0.20 ^{ab}	5.20±0.20 ^a	4.80±0.20 ^a
C	400	6.40±0.24 ^{ab}	6.20±0.37 ^{ab}	5.20±0.37 ^a	4.80±0.37 ^a
D	600	7.40±0.40 ^a	7.00±0.54 ^a	7.20±0.37	5.20±0.37
E	10 pentazocine (+ve control)	9.60±0.24	9.60±0.50	8.60±0.24	6.80±0.37

Values across column with same superscript are statistically ($p > 0.05$) not significant

Values across column with no or/different superscript are statistically ($p > 0.05$) significant

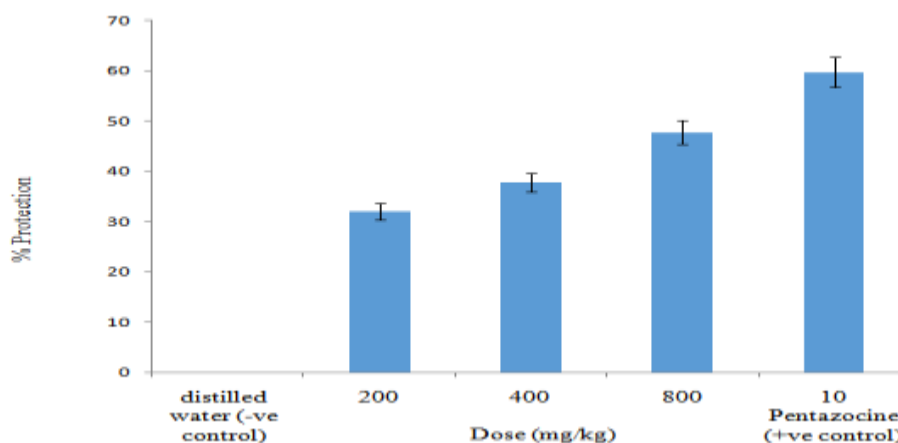


Figure 2: Anti-inflammatory Effect of Methanol Stem Bark Extract of *Senna siamea*

IV. Discussion

The phytochemical studies of the methanol stem bark extract of *Senna siamea* revealed some useful chemical compounds such as flavonoids, cardiac glycosides, tannins, saponins, terpenoids and alkaloids. These compounds have been known to exert pharmacological and antagonistic effects and still some are capable of protecting the active ingredient in herbs from decomposing either chemically or physiologically

Many researchers have given various reasons for anti-inflammatory activity. It was observed that the flavonoids detected in both extracts are known to be good anti-inflammatory agents. Studies of Raju *et al.* (2005) on anti-inflammatory potential of *Cassia fistula* revealed the responsibility of flavonoids and alkaloids in anti-inflammatory reactions. Similarly, flavonoid with anti-inflammatory potential are reported from *Morinda tinctoriaroxb.* and *Vernonia amygdalina* (Sivaraman and Muralidharan, 2010; Udemé *et al.*, 2009). In spite of flavonoids, steroids were noticed in both the extracts (ethyl acetate and methanol) and studies of Neto *et al.* (2005) reported the presence of steroids with anti-inflammatory potential in *Pfafaffia glomerata*. Terpenes have been reported to possess important biological activities, such as analgesic (Guimaraes *et al.*, 2013; Quintans *et al.*, 2013), anticonvulsant (De Sousa *et al.*, 2007), cardiovascular (Silva-Filho *et al.*, 2012) antimalarial and antibacterial (Evans, 2009). Alkaloids have pharmacological applications as anesthetics and CNS stimulants (Madziga *et al.*, 2010). More than 12,000 alkaloids are known to exist in about 20 % of plant species and only few have been exploited for medicinal purposes.

Denaturation of proteins is a well-documented cause of inflammation. Irritant-induced inflammation occurs in two qualitatively distinguishable phases (Asif, 2011). The early phase begins within minutes of phlogistic challenge due to the release of biogenic amines such as histamine, while the latter phase involves the synthesis of prostaglandins. Drugs with known cyclooxygenase inhibitory activity such as Non-Steroidal Anti-inflammatory Drugs (NSAID) suppress this later phase of oedema formation (Asif, 2011). Egg white is an alternative phlogistic agent that triggers the release of inflammatory process via release of mediators (Vogel, 2008). Edema represents the early phase of inflammation and a number of mediators have been identified to be released in a sequential manner. There is an initial release of histamine and 5-hydroxytryptamine producing an increased vascular permeability followed by release of kinins further contributing to the increased vascular permeability and finally, the prostaglandins and slow reacting substance are released to maintain the increased vascular permeability by histamine, 5-hydroxytryptamine and kinins (Crunkhorn and Meacock, 1971). The tail immersion has been used to study centrally acting analgesics (Woolfe and MacDonald, 1994; Bachlav *et al.*, 2009). In these tests, the nociceptors are sensitized by sensory nerves and the involvement of endogenous substances such as prostaglandins are minimized. Thus from the results, we can conclude that the analgesic activity of *Senna siamea* may be fully mediated through central mechanism.

Inhibition of acetic acid-induced writhing in mice by extract (200 and 400 mg/kg) suggested that the analgesic effect of the extract may be peripherally mediated via the inhibition of the synthesis and release of prostaglandins (Koster *et al.*, 1959). The acetic acid induced mouse writhing test has been used extensively to qualify analgesic agents that have peripheral analgesic activity (Neves *et al.*, 2007). Writhing induced by chemical substances injected intraperitoneally, are due to sensitization of nociceptors by prostaglandins.

Alkaloids, flavonoids and saponins are known to possess analgesic activity (Evans, 2009). The activity of the extracts was found to be dose dependent and significant at $P < 0.05$.

V. Conclusion

The phytochemical study revealed the presence of saponins, cardiac glycosides, tannins, flavonoids, terpenoids, alkaloids and carbohydrates in the stem bark extract of the plant. The stem bark extract had an $LD_{50} \geq 5000$ mg/Kg. The stem bark extract induced some degree of effects on the peripheral and central nervous system as it exerts anti-inflammatory and induced analgesia. However, Purification, isolation and characterization using physical techniques such as HNMR, ^{13}C NMR and IR-Spectroscopy should be carried out in order to confirm the chemical structures of the bioactive constituents responsible for the plant's pharmacologic actions.

Acknowledgement

The authors appreciate the Tertiary Education Trust Fund (TETFUND) sponsored Institutional Based Research (IBR) Grant of 2017 (TETFUND/DESS/UNIMAID/MAIDUGURI/RP/Vol.V) for the fund used for this study.

References

- [1]. Abdulrahman FI. Studies on the chemical contents and pharmacological activities of the root-bark extract of *Vitex doniana* (Black Plum). Ph.D. Thesis, University of Maiduguri, Maiduguri, Nigeria.2004, 166pp.
- [2]. Abdulrahman, F. I., Akan, J. C., Sodipo, O. A. and Onyeyili, P. A. Effect of aqueous root-bark extract of *Vitex doniana* sweet on hematological parameters in rats. *Journal of American Science*, 2010, 6, 8-12.
- [3]. Adebayo JO, Krettli, AU. Potential antimalarials from Nigerian plants: A Review. *Journal of Ethnopharmacology*, 2011; 133: 289-302.
- [4]. Al-Adhroey AH, Nor ZM, Al-Mekhlafi HM, Mahmud R. Ethnobotanical study on some Malaysian anti-malarial plants: A community based survey. *Journal of Ethnopharmacology*, 2010;132: 362–364.
- [5]. Asif M. In vivo analgesic and anti-inflammatory effect of *Tectona grandis* Linn stem bark extracts. *Malaysian Journal of Pharmaceutical Sciences*, 2008; 9(1): 29 – 43.
- [6]. Brian KR, Turner TD. *Practical Evaluation of Phytochemicals*, Wright Scientechnical, Bristol, UK. 1975, 57-59.
- [7]. CIOMS. and ICLAS: Principles find medical sciences and the International Council for Laboratory Animal Science. Guiding Principles for Biomedical Research Involving Animals . find pdf. <http://idas.Org/wp-content/uploads/2013/03/CIOMS-ICLAS.2012>, Access Date: 22/4/2015.
- [8]. Correa CR, Kyle DJ, Chakrasvarty S, Calixto JB. Anticoceptive receptor antagonist NPC 18688 in mice. *British Journal of Pharmacology*, 1996; 117: 552 – 558.
- [9]. Crunkhorn P, Meacock SC. Mediators of the inflammation induced in the rat paw by carrageenan. *Britain Journal of Pharmacology*, 1971; 42(3): 392-402.
- [10]. Dalziel JM. *The useful plants of West Tropical Africa – An appendix to the second edition of the flora of West Tropical Africa* (Hutchinson, J and Dalziel, J.M) Great Britain Watcmangs Ltd., 1963; 596-597.
- [11]. De Sousa DP, Quintans-Jr, LJ, Almeida RN. Evaluation of the anticonvulsant activity of alfa-Terpineol. *Pharmaceutical Biology*, 2007; 45: 69–70.
- [12]. Evans WC. *Trease and Evans Pharmacognosy*. 16th Edition. Saunders Publishers, London. 2009; 42–229.
- [13]. GraphPad Software: GraphPad Software InStat guide to choosing and interpreting statistical tests, GraphPad Software, Inc., San Diego California USA Version 3.10 32 bit for windows: www.graphpad.com. 2000.
- [14]. Guimaraes AG, Quintans JSS, Quintans-Jr LJ. Monoterpenes with analgesic activity-A systematic review. *Phytotherapy Research*, 2013; 27: 1–15.
- [15]. Kiepe P. Effect of *Cassia siamea* hedgerow barriers on soil physical properties. *Geoderma*; 1995; 66: 113-120.
- [16]. Koster R, Anderson M, De-Beer EJ. Acetic acid for analgesic screening. *Federation Proceedings*, 1959; 18: 412-418.
- [17]. Koudouvo K, Karou DS, Kokou K, Essien K, Aklikokou K, Glitho IA, Simpore J, Sanogo R, Souza C, Gbeassor M. An ethnobotanical study of antimalarial plants in Togo Maritime Region. *Journal of Ethnopharmacology*; 2011; 134: 183–190.
- [18]. Liu RH. (2004). Potential synergy of phytochemicals in cancer prevention: Mechanism of action. *Journal of Nutrition*, 2004; 134(12): 3479S-3485S.
- [19]. Lorke D. Approach to acute toxicity test, *Archive Toxicology*, 1983; 54: 275 – 287.
- [20]. Madziga HA, Sanni S, Sandabe UK. Phytochemical and elemental analysis of *Acalypha wilkesiana* leaf. *Journal of American Science*, 2010; 6(11): 510-514.
- [21]. Marazzi B, Endress PK, Queiroz LP, Conti E. Phylogenetic relationships within *Senna* (Leguminosae, Cassiinae) based on three chloroplast DNA regions: patterns in the evolution of floral symmetry and extrafloral nectaries". *American Journal of Botany*. 2006; 93(2): 288–303.
- [22]. Markham KR, Mues R, Stoll M, Zinsmeister, HD. NMR spectra of flavones di-*C*-glycosides from *Apometzgeria pubescence* and the detection of rational isomerism in 8-*C*-hexosylflavones, *Zeitschrift fur Naturforschung*, 1987; 42: 1039-1042.
- [23]. Maurya R, Dongarwar N. Studies on the medicinal uses of wild trees of Nagpur District. *International Journal of Life Science and Pharmaceutical Research*, 2012; 2: 21-24.
- [24]. Nwafor PA. Anticoceptive and other pharmacological effects of *Asparagus pubescence* bark root and *Cassia nigricans* leaves. PhD Thesis, (unpublished) University of Jos. Nigeria. 1998
- [25]. Odason EE, Kolawole J. Anti-diabetic properties and brine shrimp toxicity of the aqueous extract of the root of *Cassia siamea* Lam. (Ceasalpiniaceae). *Nigerian Journal of Pharmaceutical Research*, 2007; 6: 66-69.
- [26]. Owoloye BV, Olaleye SB, Oke JM, Elegbe RA. Anti - Inflammatory and Analgesic Activities of *Nothospondias staudtii*, *Nigerian Journal of Physiology and Science*, 2004; 19(1-2): 102-105.
- [27]. Raju I, Moni M, Subramanian V. Anti-inflammatory and antioxidant activities of *Cassia fistulalinn* bark extract. *African Journal of Traditional Complementary and Alternative Medicine*, 2005; 2(1): 70-85.
- [28]. Randell BR, Barlow B A. *Senna* pp 89-138. In: A. S. George (executive editor). *Flora of Australia* volume 12. *Australian Government Publishing Service: Canberra, Australia*. pp. 1998; 89-138.
- [29]. Roja G, Rao PS. Anticancer compound from tissue cultures of medicinal plant. *Journal of Herbs, spices and medicinal plants*. 2000;7: 71-102.
- [30]. Santos ARS, Cechinel FV, Nieri R, Viano AM, Moreno PN, Campos MM, Yunes RA, Calixto JB. Analgesics of culture from selected species of Phyllantu. *Journal of Pharmacy and Pharmacology*, 1994; 46.
- [31]. Sarker SD, Nahar L. *Chemistry for Pharmacy Students: General Organic and Natural Product Chemistry*. John Wiley and Sons, England. 2007; 283-359.
- [32]. Silva GL, Lee I, Douglas KA. Special problems with extraction of plants.In: Cannel, J.P.R. (ed.). *Natural Products Isolation*. Humana press publishers, New Jersey (USA). 1998; 356-358.
- [33]. Silva-Filho JC, Oliveira NNPM, Arcanjo DDR, Quintans-Jr, LJ, Cavalcanti SCH, Santos MR. Investigation of mechanisms involved in (-)-borneol-induced vasorelaxant response on rat thoracic aorta. *Basic Clinical Pharmacology and Toxicology*, 2012; 110: 171–177.
- [34]. Sivaraman, D. and Muralidharan, P. Anti-Ulcerogenic evaluation of root extract of *Ficushispida Linn*, in aspirin ulcerated rats. *Afr. J. Phar. Pharmacol*. 2010; 4(2), 079-082.
- [35]. Sodipo OA, Abdulrahman FI, Sandabe UK, Akinniyi JA. Effects of the aqueous fruit extract of *Solanum macrocarpum* Linn. on Haematological parameters of triton-induced hyperlipidemic rats. *African Journal of Pharmacy Pharmacology*, 2011; 5(5), 632-639.
- [36]. Sofowora AE. *Medicinal Plants and Traditional Medicine in Africa*. 3rd Edition. Spectrum Books Limited, Ibadan Nigeria, 2008; 97-112.
- [37]. Udemé OG Owonari AG. Evaluation of anti-inflammatory activity of extract of *Vernonia amygdalina*. *East. Journal of Medicine*. 2009; 14: 20-22.

- [38]. Usman H, Abdulrahman FI, Usman A. Qualitative phytochemical screening and in vitro Antimicrobial effects of methanol stem bark extract of *Ficus thonningii* Moraceae. *African Journal of Traditional and Complementary Alternative Medicine*. 2009; 6(3): 289-295.
- [39]. Vishnoi NR. *Advanced Practical Chemistry*. Yikas Publication House, PVT Ltd. Ghaziabad-India, 1979; 447-449.
- [40]. Vogel HG. *Drug Discovery and Evaluation: Pharmacological Assays*. Third Edition. Springer. Aalen- Germany. 2005; 1164-1165.
- [41]. Wink M, Schmeller T, Latz-Briining B. Modes of action of allelochemical alkaloids: Interaction with neuroreceptors, DNA and other molecular targets. *Journal of Chemical Ecology*, 1998; 24: 1888-1937.
- [42]. Winter CA, Risley EA, Nuss GW. Carrageenan-induced oedema in the hind limb of rat as an assay for anti-inflammatory activity. *Professional Society of Experimental Biology and Therapy*, 1962; 111: 544-547.

Sodipo O. A." Phytochemical Screening, Analgesic And Anti-Inflammatory Activities Of Methanol Stem Bark Extract Of *Senna Siamea* Lam. (Kassod Tree)". *IOSR Journal of Pharmacy and Biological Sciences (IOSR-JPBS)* 13.3 (2018): 38-45.