

Phylogenetic analysis of some Hymenochaetaceae members

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Abstract: The Hymenochaetaceae family of Phylum Basidiomycotina is characterized by 28 genera and 400 species to date. Phylogenetic analysis of various members of this family was attempted using morphological and molecular traits. Thirty samples belonging to Hymenochaetaceae were collected from Pune region. Morphological studies were carried out using 14 quantitative and 70 qualitative characters. Out of these, 8 quantitative characters of the 30 samples along with the same characters from previously reported genera of this family were used in Principle Component Analysis (PCA) in an attempt to group the samples in different clusters. In addition DNA sequence data corresponding to 80S ribosomal large subunit rRNA (28S) and 70S (mitochondrial) ribosomal small subunit rRNA (16S) was collected for the same 30 samples. Sequence data for the same rRNA molecules was also downloaded from NCBI site for 12 reported genera of Hymenochataceae. The sequences were aligned using MEGA (version 4) software and a phylogenetic tree was constructed using maximum parsimony (PAUP 4.0). Results indicated that clustering achieved using DNA sequence data differed from that using morphological data. Probable causes for this discrepancy could be that the evolution rate at phenotypic and molecular levels may differ and / or that the phenotype is determined by environmental factors in addition to genetic factors.

Keywords: Hymenochaetaceae, Phylogenetic analysis

I. Introduction

Family Hymenochaetaceae forms a cosmopolitan group of wood inhabiting fungi that metabolize lignin for their growth and reproduction. It consists of 28 genera and 400 species exhibiting saprophytic and parasitic features. It is characterised by xanthochroic reaction, yellow to deep brown trama, generative hyphae without clamp connection and frequent occurrence of setae. The description of hymenochaetacean genera is based on characters pertaining to morphology, anatomy, sexuality, nuclear behaviour, pigmentation, and ecology and is rather imprecise. In this work we have attempted to identify 30 samples collected from Pune and Switzerland on the basis of information available on 12 genera of Hymenochaetaceae using DNA sequence data (rDNA) as well as morphological and anatomical data (1,2,3,4,5).

II. Material and methods

25 samples were collected from Pune region such as Pune University, Empress Garden, Papel Seminary (Nagar road), Katraj Ghat, NDA reserved forest, Taljai, Khadakwasla, Sinhgad, Lohagad, Mulshi, Dongarwadi, Lonavala, Bhimashankar, and road side old avenue trees in Pune city. 5 samples of Hymenochaetaceae members were obtained from Dr. Valérie Hofstetter, Agroscope Changins-Wädenswil Research station Institute, Switzerland. Morphological studies were carried out using 11 quantitative characters (Table 1) of the 30 collected samples and other genera of this family reported in literature were subjected to Principle Component Analysis (PCA) in an attempt to group the samples in different clusters. Cladistic analysis was also done using the same morphological characters using Pearson correlation and single-linkage distance method (MINITAB software). In addition, DNA sequence data corresponding to 80S ribosomal large subunit rRNA (28S) and 70S (mitochondrial) ribosomal small subunit rRNA (16S) was collected for the same 30 samples (6,7,8). Sequence data for the same rRNA molecules was also downloaded from NCBI site for 12 reported genera of Hymenochataceae. The sequences were aligned using ClustalW method (MEGA 4 software) and a phylogenetic tree was constructed using maximum parsimony (PAUP 4.0).

III. Results and Discussion

The PCA analysis did not lead to distinct clusters for the samples. However the Cladistic analysis of morphological data showed distinct grouping between temperate and tropical genera (Fig 1). All temperate samples showed similarity with the type specimens chosen for the analysis. Of the tropical samples, 15 formed a group with the *Phellinus* type species: *P. laevigatus*, *P. tremulae* and *P. lundelli* while 8 samples formed an individual cluster, which did not include any type genera.

The dendrogram generated using 80S ribosomal large subunit (LSU) rRNA (28S) region showed 10 distinct clusters out of which 9 clusters included at least one type genus (Fig 2a). The dendrogram generated using 70S (mitochondrial) ribosomal small subunit (mitSSU) rRNA (16S) showed 8 clusters which included the type genera (Fig 2b). *Fomitopsis* and *Coltricia* formed outgroups. None of the samples clustered with *Hymenochaetae*, *Coltricia*, *Omnia* and *Porodaedalea*. The tropical samples showed similarity to either *Phellinus*, *Inonotus*, and *Fulvifomes* and the clustering based on morphological and molecular data differed (Table 2).

IV. Figures and Tables

Table 1

Number	Quantitative Characters
1	Area of basidiocarp in square cm
2	no of pores / mm
3	Width of Contextual skeletal hyphae in μm
4	Width of Hymenial skeletal hyphae in μm
5	mean length of basidiospore in μm
6	mean breadth of basidiospore in μm
7	context layer height in mm
8	margin width in mm
9	no. of strata
10	length of pore tube in mm
11	length of setae in μm

Table 2

Number	Genus in Hymenochaetaceae	Distribution of samples no.		Geographical location
		Molecular	Morphological	
1	<i>Fomitopsis</i>	173	173	Switzerland
2	<i>Fuscoporia</i>	58	58	Switzerland
		21	-	Pune
3	<i>Fomitiporia</i>	62	62	Switzerland
4	<i>Coltricia</i>	-	-	-
5	<i>Inonotus</i>	49		Pune
		50		Pune
		51	51	Pune
		53	53	Pune
			59	Switzerland
6	<i>Phylloporia</i>	59	-	Switzerland
7	<i>Phellinus igniarius</i>	55	55	Switzerland
8	<i>Phellinus</i>	-	2, 7, 8, 10, 11, 12, 17, 21, 26, 29, 36, 43, 45, 145, 153	Pune
9	<i>Fulvifomes</i>	1, 2, 11, 26, 28, 29, 36, 45, 145, 153		

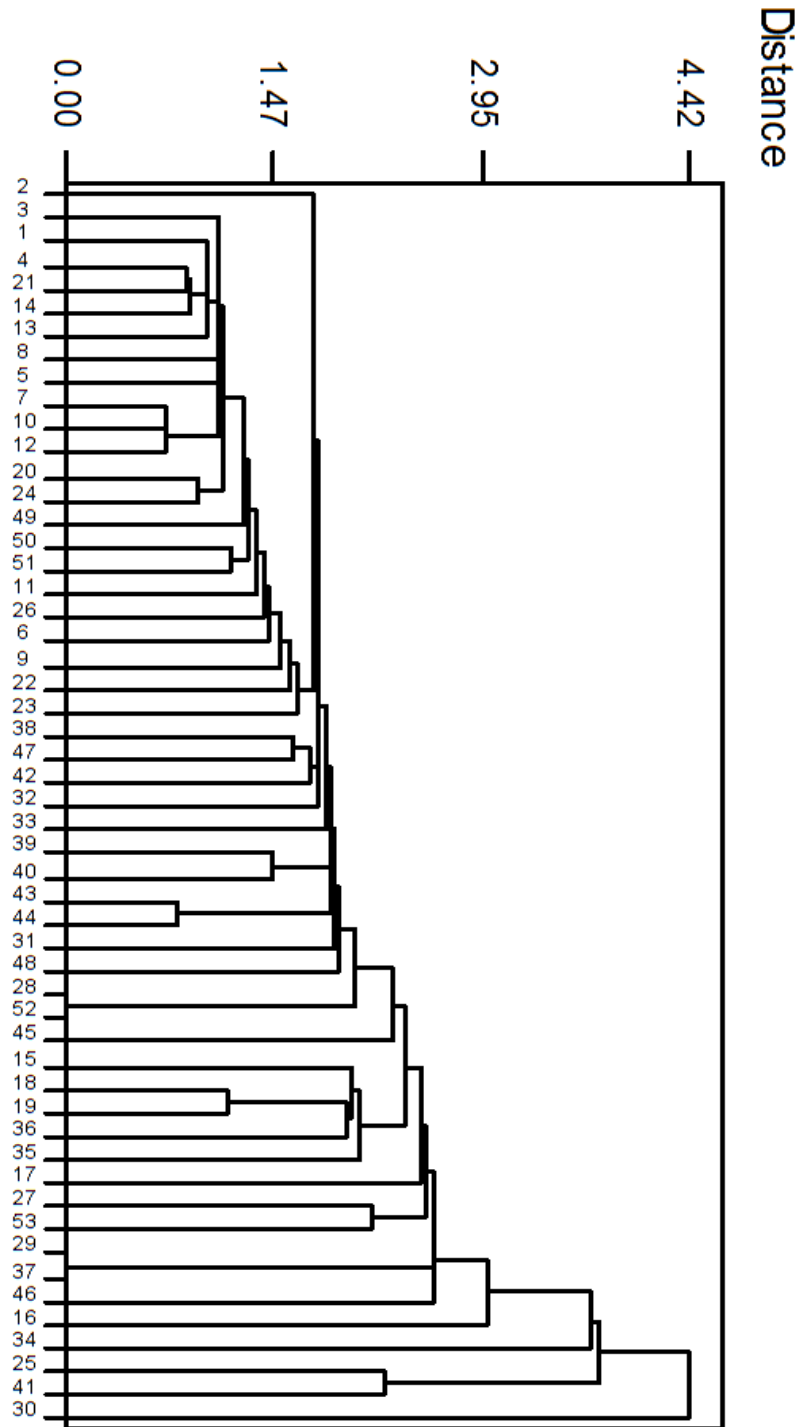


Fig 1. PCA analysis using Morphological character data

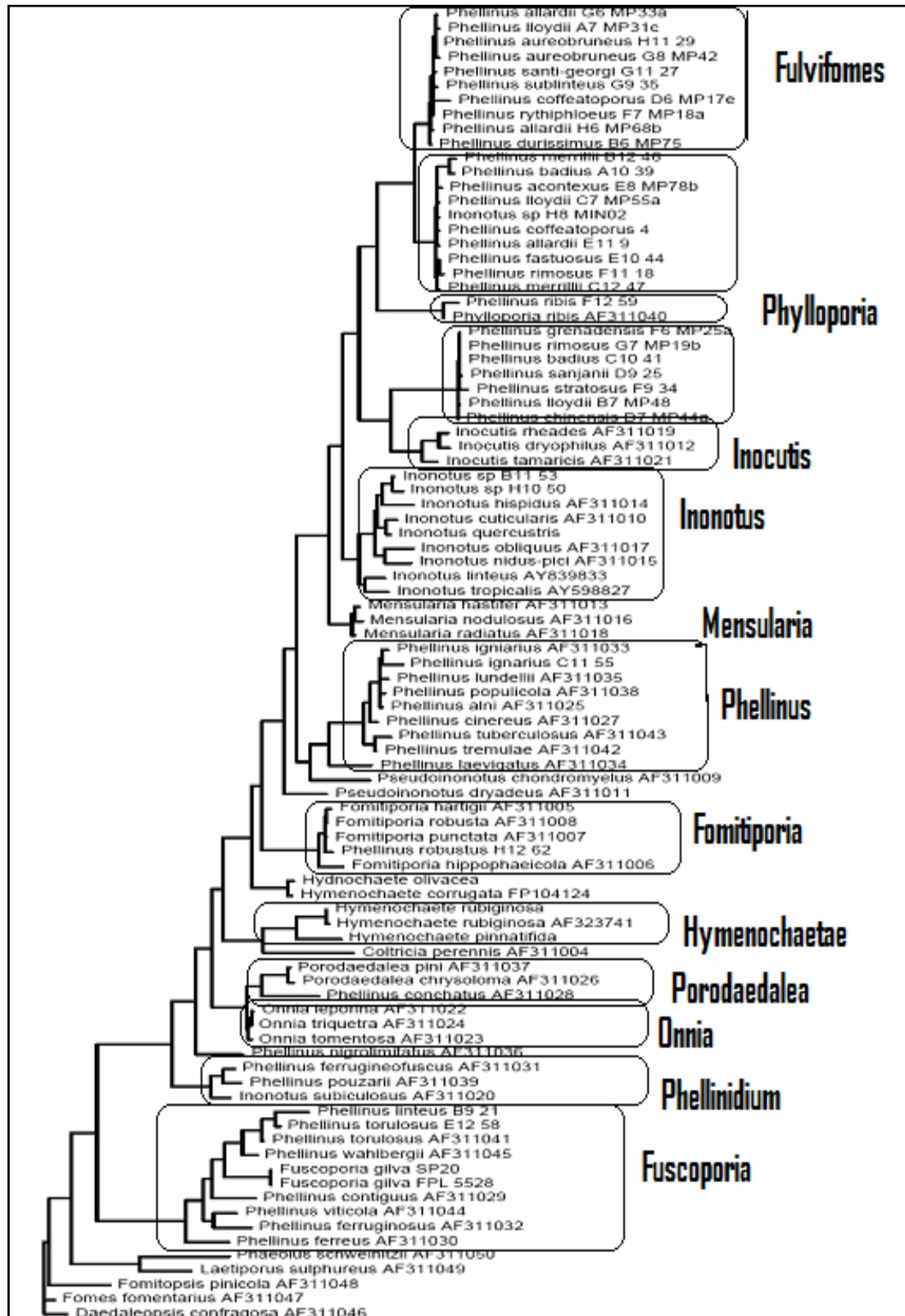


Fig 2a: Dendrogram generated using 80S ribosomal large subunit (LSU) rRNA (28S)

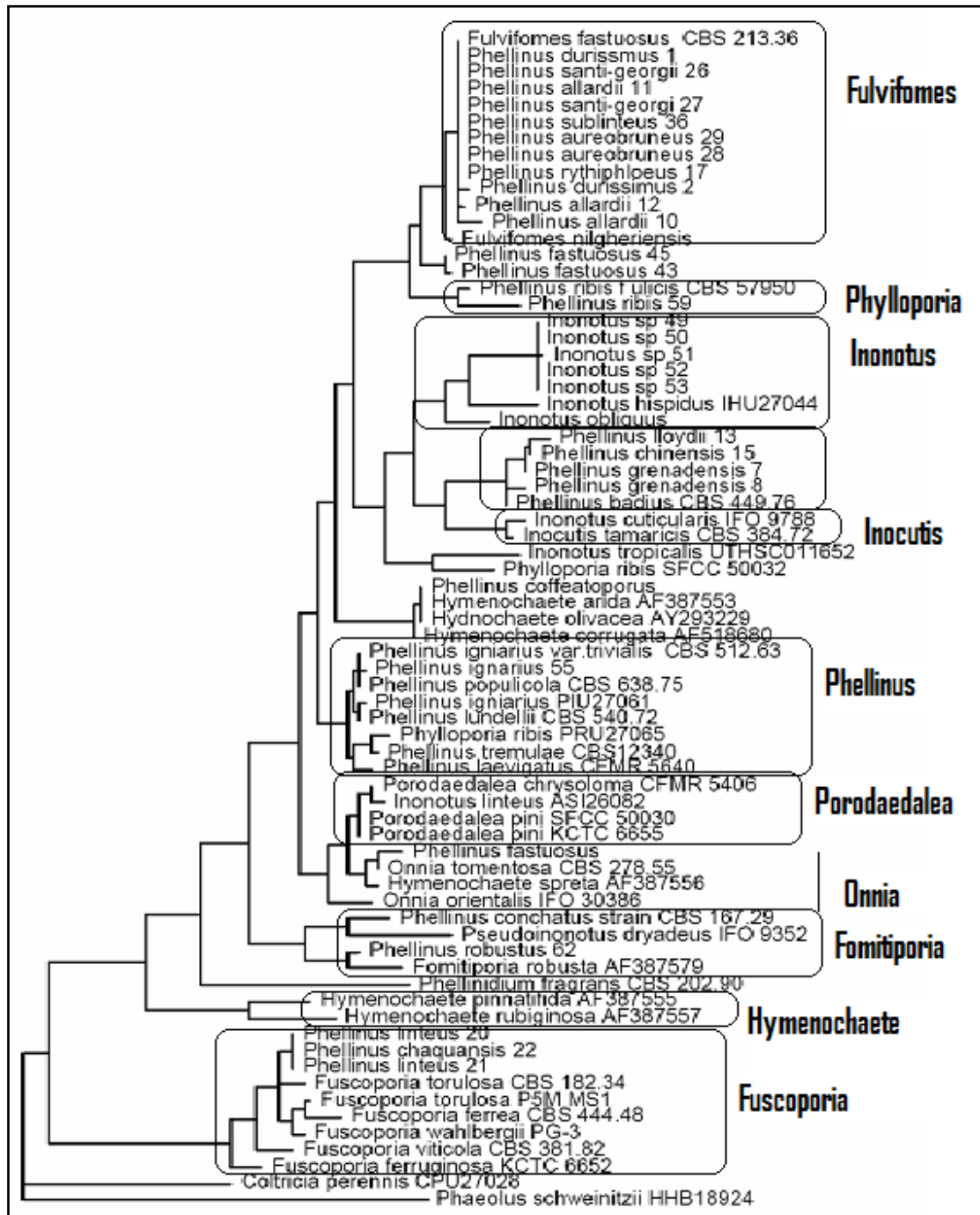


Fig 2b: Dendrogram generated using 70S (mitochondrial) ribosomal small subunit (mitSSU) rRNA (16S)

V. Conclusion (11 Bold)

Results indicated that clustering achieved using DNA sequence data differed from that using morphological data. *Fulvifomes* was classified as a sub-genus of *Phellinus* based on morphology, anatomy, pigmentation, and karyology (9-14). Later they were identified as two distinct genera on the basis of molecular data as well as the presence / absence of setae (13). Our data showed a distribution of 15 samples under *Phellinus* on the basis of morphological data, while molecular data showed the distribution of 10 of these same samples under *Fulvifomes*. Discrepancy in the conclusions drawn from morphological and molecular data could have arisen due to different rates of evolution at phenotypic and molecular levels and / or the role of environmental factors in determining the phenotype besides genetic factors.

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References

- [1]. Dai Y. C. (1995) Changbai wood-rotting fungi 3. The genus *Phellinidium* (basidiomycetes) and a new species, *P. aciferum*. *Ann Bot Fennici* 32:63–73.
- [2]. Dai Y. C. (1999) *Phellinus* sensu lato (Aphyllphorales, Hymenochaetaceae) in East Asia. *Acta Bot Fennica* 166:1–115.
- [3]. Fiasson J. L. and Niemela T. (1984) The Hymenochaetales: a revision of the European poroid taxa. *Karstenia* 24:14–28.
- [4]. Kirk, P.M. Cannon, P.F. David, J.C. and Stalpers J.A. (2001) *Dictionary of The Fungi*. 9th Edition, CAB International. Pp 655.
- [5]. Larsen, M. J. and Cobb-Pouille, L. A. (1990) *Phellinus* (Hymenochaetaceae). A survey of the world taxa. *Fungiflora*, Oslo, Norway pp 186.
- [6]. Aristóteles, G. N. Clarice, L.L. and Rosa, T. G. (2005) DNA extraction from frozen field collected and dehydrated herbarium fungal basidiomata: performance of SDS and CTAB-based methods. *Biotemas* 18 (2): 19 - 32, 2005.
- [7]. Bruns, T.D. Fogel, R. and Taylor J.W. (1990) Amplification and sequencing of DNA from fungal herbarium specimens. *Mycologia* 82: 175-184.
- [8]. Lamrood P.Y. (2004) Studies of some medicinally important fungi, Ph.D. Thesis, Department of Botany, University of Pune, Pune, 411007 pp 490
- [9]. Fischer M. and Binder M. (2004) Species recognition, geographic distribution and host-pathogen relationships: a case study in a group of lignicolous basidiomycetes, *Phellinus* s.l. *Mycologia*, 96(4), 799–811.
- [10]. Fischer M. and Wagner T. (1999) RFLP analysis as a tool for identification of lignicolous basidiomycetes: European polypores. *European Journal of Forest Pathology* 29, 295–304.
- [11]. Niemela T, Wagner, T. Fischer, M and Dai Y-C. (2001) *Phellopilus* gen. nov. and its affinities within *Phellinus* sensu lato and *Inonotus* sensu lato (basidiomycetes). *Ann Bot Fennici* 38:51–62.
- [12]. Wagner T. and Fischer M. (2001) Natural groups and a revised system for the European poroid Hymenochaetales (Basidiomycota) supported by nLSU rDNA sequence data. *Myc Res* 105:773–782.
- [13]. Wagner T. and Fischer M. (2002) Classification and phylogenetic relationships of Hymenochaete and allied genera of the Hymenochaetales, inferred from rDNA sequence data and nuclear behaviour of vegetative mycelium. *Myc Prog* 1:93–104.
- [14]. White, T.J., Bruns, T., Lee, S. and Taylor, J.W. (1990) Amplification and direct sequencing of fungal ribosomal RNA genes for phylogenetics. In *PCR protocols: A guide to methods and application*. Innis, M.A., D.H. Gelfand, J. J. Sninsky, and T.J. White (eds.) pp. 315-322, Academic Press Inc., San Diego.