

## Longevity of Survival of *Trichoderma harzianum* on Sucrose added Cakes

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**Abstract:** Tree borne oilseeds (TBOs) namely Neem cake, Jatropha cake, Mahua cake and Karanja cake were evaluated for mass culturing of *Trichoderma harzianum* showed that Neem cake is the best substrate in vitro while Jatropha cake in vivo. The antagonist could be stored for more than 3 months but an increase in number of viable propagules up to 4 months was measured from all the substrates except Karanja (up to 105 days) when mixed with Sucrose; a kind of sugar. Among all substrates Neem cake found maintaining high population at 30 days of inoculation with *T. harzianum* in the form of colony forming units (CFUs) per gram of substrate.

**Keywords:** Tree born oilseeds (TBOs); sucrose; *Trichoderma harzianum*; self-life; longevity; viability etc.

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### I. Introduction

Since antagonists are present as low population in native soils, further augmentation of their density to reach a higher stability through artificial application is necessary. Various agricultural wastes and by-products for multiplication of bio-control agents have been used by several workers [1], [2], [3] and [4].

*Trichoderma* as an effective antagonist against a range of plant pathogens won considerable scientific attention [5] and [6]. Different organic media like Neem cake, coir pith, farmyard manure, and decomposed coffee pulp also have been suggested for its multiplication [7]. Therefore, a study was conducted to enhance the conidial yield of *T. harzianum* on these substrates using nutritional supplements.

### II. Materials And Methods

#### 2.1 Sources and maintenance of culture

##### 2.1.1 Collection of soil samples and isolation of bio-control agent

To isolate the bio-control agent i.e., *Trichoderma harzianum* from rhizospheric soil, samples were collected from fields of Crop Research Centre (CRC) of university. For isolation, 1g of soil sample was placed in a 250 ml conical flask containing 100 ml of sterilized distilled water (SDW) and mixed thoroughly. Different dilutions of working samples were prepared by diluting the stock solution. 1 ml of last serial dilution i.e.,  $10^{-6}$  was spread on Trichoderma Selective Medium (TSM) for isolation of *T. harzianum* [8]. The plates were incubated for 7 days at  $28 \pm 2^\circ\text{C}$  and after incubation, pure culture was grown; it was initially hyaline but turned green as conidia produced [9].

##### 2.2 Maintenance of the culture

The fungus was sub-cultured in the PDA slants and allowed to grow at  $28 \pm 2^\circ\text{C}$  temperature. The culture thus obtained was stored in refrigerator at  $5^\circ\text{C}$  for further study and was sub cultured periodically.

##### 2.3 Screening of different oil cakes for mass multiplication of *T. harzianum*

###### 2.3.1 Collection of oil cakes and other substrates

Cakes of Neem, Jatropha, Mahua and Karanja were collected from local agricultural product-processing units and farms. Materials were cleaned and crushed (in heavy pestle & mortar) to prepare a coarse powder (particles of approximately 1 mm diameter). Cakes were mixed with sterilized water (SDW) to maintain 25% moisture (10: 2.5, w/v) and autoclaved at  $1.1 \text{ kg/cm}^2$  for 20 minutes. The flasks were allowed to cool at room temperature prior to inoculation. Flasks containing substrates were inoculated with 3-4 days old actively growing culture of *T. harzianum* (2-3 bits of about 5mm size) under aseptic conditions in laminar flow. The flasks were shaken thoroughly once a day, and incubated at  $28 \pm 2^\circ\text{C}$  for 30 days. For each treatment, three replicates of flasks were maintained and arranged in a completely randomized manner (CRD).

## 2.4 Determination of population dynamics

Population dynamics were determined by serial dilution plate technique. Growth and development of *T. harzianum* were monitored after each 15 days interval up to 120 days under laboratory conditions. 1g substrate of each cake was taken from each flask for colony forming units (CFUs) count, using PDA medium.

## 2.5 Colony Forming Units (CFUs)

### 2.5.1 Serial dilution and CFUs counting

One gram of sample (substrate where *Trichoderma* was being grown) was suspended in 10 ml distilled water to make microbial suspension diluted 10 times, it will give 1: 10 conc. or  $10^{-1}$  dilution of original sample, i.e. the original sample has been diluted to 1/10th. Similarly it was prepared up to  $10^{-6}$  on dilution of the original sample. Finally 1ml of microbial suspension from last serial dilution i.e.,  $10^{-6}$  was added to sterile Petri dishes (triplicate in completely randomized manner) and incubated for 5 days at  $28 \pm 2^{\circ}\text{C}$ . The density of cells, spores/conidia of *T. harzianum* can be measured in laboratory by Plate Dilution technique [10].

No. of colonies CFUs = _____ Amount plated X dilution factor
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## 2.6 Statistical analysis

The data's were analyzed using ANOVA and treatment means differentiated using Fischer's completely randomized design (CRD) *in vitro* studies. Statistical analysis was conducted using general linear model procedures of SPSS version 16 [11] and [12].

## III. Results

### 3.1 Screening of different oil cakes added with sucrose for mass culturing of *T. harzianum*

#### 3.1.1 AT 15 DAI

Four oil cakes exhibited varying response toward sucrose, after 15 days of incubation. Neem cake ( $43.67 \times 10^6$  CFUs) amended with sucrose emerged out to be significantly superior in giving rise to higher population dynamics followed by Jatropha cake ( $40.67 \times 10^6$  CFUs), Mahua cake ( $37.00 \times 10^6$ ). Karanja cake was found significantly lower than Mahua cake, which supported only  $33.00 \times 10^6$  CFUs of *T. harzianum* (Table 2, Fig. 2).

#### 3.1.2 AT 30 DAI

At 30 DAI, the highest mean population of the *T. harzianum* was supported by Neem cake ( $67.67 \times 10^6$  CFUs), which was significantly higher than the mean population supported by Jatropha cake ( $57.00 \times 10^6$ ). Mahua cake was next best formulation, which supported  $51.67 \times 10^6$  CFUs of *T. harzianum* was significantly lower than Jatropha cake and followed by Karanja cake ( $49.00 \times 10^6$ ), which was found significantly lower to the Mahua cake (Table 2, Fig. 2).

#### 3.1.3 AT 45 DAI

A slight reduction in CFUs was recorded after 45 days when Neem cake supported  $60.67 \times 10^6$  CFUs whereas, mean population supported by Jatropha cake is  $50.00 \times 10^6$  followed by Mahua cake, which supported  $44.67 \times 10^6$  CFUs, while lowest population was supported by Karanja cake i.e.  $42.00 \times 10^6$  CFUs of *T. harzianum* and it was found significantly lower than Mahua cake (Table 2, Fig. 2).

#### 3.1.4 AT 60 DAI

After 60 days, highest mean population of the *T. harzianum* was supported by Neem cake ( $46.33 \times 10^6$ ), which was significantly higher than Jatropha cake ( $37.33 \times 10^6$ ), Mahua cake ( $32.00 \times 10^6$  CFUs) and Karanja cake which supported  $27.33 \times 10^6$  CFUs of *T. harzianum*. Karanja cake was found significantly lower than Mahua cake (Table 2, Fig. 2).

#### 3.1.5 AT 75 & 90 DAI

Neem cake found significant in supporting CFUs of *T. harzianum*, which supported  $36.67 \times 10^6$  and was significantly higher than Jatropha cake, Mahua cake and Karanja cake which supported  $27.00 \times 10^6$ ,  $22.67 \times 10^6$  and  $18.67 \times 10^6$  CFUs, while after 90 days of inoculation maximum no. of CFUs were recorded again in Neem cake ( $24.33 \times 10^6$ ) followed by Jatropha cake ( $21.67 \times 10^6$ ), Mahua cake ( $19.33 \times 10^6$ ). Least performing cake was Karanja cake with  $15.67 \times 10^6$  CFUs of *T. harzianum* and was found significantly lower than Mahua cake (Table 2, Fig. 2).

### 3.1.1 AT 105 &120 DAI

Even after 105 and 120 days of inoculation highest population dynamics of *T. harzianum* was exhibited by Neem cake (20.00 & 7.33x10<sup>6</sup>) found significantly higher than Jatropha cake (15.33 & 2.33x10<sup>6</sup>) while Mahua cake with supporting 12.67 & 1.00x10<sup>6</sup> CFUs and found significantly lower than Neem and Jatropha cake. Least population of *T. harzianum* was supported by Karanja cake (10.33 x10<sup>6</sup>) found significantly lower than others and didn't support any CFUs after 120 days (Table 2, Fig. 2).

## IV. Discussion

In order to find out an additional source of carbon which can support in enhancing the growth, sporulation and longevity of *T. harzianum* over a comparatively longer period, an experiment was conducted to test the efficacy of sucrose, as amended to four TBOs cakes i.e., Neem, Jatropha, Mahua and Karanja. It was interesting to note that addition of sucrose to these cakes resulted in increasing the survival of *T. harzianum* up to 120 days, whereas without addition survival was up to 105 days.

Thangavelu [13] reported that addition of jaggery (10% w/v) to the substrate i.e., dried banana leaves increased the multiplication of *T. harzianum* which survived up to more than 6 months on the stored substrate. The results obtained in present study in regards to CFUs enhancing effect of sucrose mixing to the de-oiled cakes are comparable with the previous work of Thangavelu [13] as jaggery is a raw form of sucrose which helped in enhancing the population of *T. harzianum*.

Bean and Wilcoxson [14] found the best growth of *Helminthosporium sativum* and *H. dictyoides* on sucrose. Whereas, Chandwani and Munjal [15] reported that maltose was the best source of carbon followed by sucrose and lactose for *H. sativum*, *H. oryzae*, *H. carbonum* and *H. gramineum* that supports the finding of present investigation.

Microencapsulation with sugars, such as sucrose, molasses or glycerol, significantly ( $P < 0.05$ ) increased the survival percentages of conidia after drying. Xixuan and Custis [16] suggested that microencapsulation of conidia with 2% sucrose solution resulted in the highest survival percentage of *T. harzianum* (7.5 x 10<sup>10</sup> CFUs) supports current findings of research performed. Result shown in present investigation found comparable with work of Sriramet *al.* [17] and they declared that the addition of glycerol at 3 and 6% extended the shelf-life (with viability of >2 x 10<sup>6</sup> CFU g<sup>-1</sup>) for 7 and 12 months, respectively compared to 4–5 months shelf-life in formulations derived without the addition of glycerol.

Aamir *et al.* [18] also carried a study to assess the effects of temperatures (20°C or 30°C) and sugars (dextrose or sucrose) on conidium germination and bioactivity as fresh conidia, or after 6 months of storage supported the findings of present study.

## V. Conclusion

Neem cake was found to be superior among all the cakes at the entire DAIs followed by Jatropha cake. Mahua was next to the Jatropha in supporting the population of *T. harzianum* and the minimum CFUs were supported by Karanja cake up to 105 days after incubation without addition of Sucrose during the course of investigation (Table 1, Fig. 1). Once sucrose is added a positive effect in enhancing the CFUs and longevity of survival, has been noticed. Similarly without sucrose Jatropha and Mahua cakes couldn't support the *T. harzianum* population after 105 days, while with sucrose even these two cakes could support the population up to 120 days (Table 2, Fig. 2).

Sucrose is main constituent of several semi-synthetic or synthetic culture media for fungal isolation and further sub-culturing, thus, its role is well justified for fungal growth promotion. Sucrose being the disaccharide might have supplied comparatively little more amount of carbon than either mono-saccharides could have helped in increasing the longevity of *T. harzianum* in Jatropha and Mahua cakes from 105 to 120 days and also in increasing the population dynamics.

**Table 1** Screening of different oil cakes up to 105 days after incubation (DAI) without added sugar

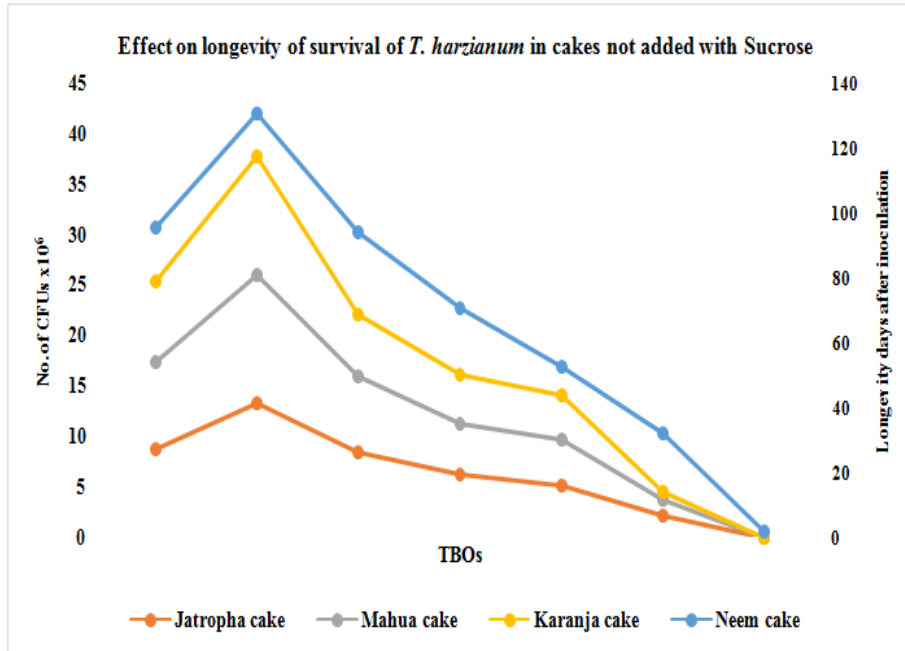
S.N.	Treatments	CFUs x10 <sup>6</sup>						
		15 DAI	30 DAI	45 DAI	60 DAI	75 DAI	90 DAI	105 DAI
1	Neem cake	30.67 <sup>a</sup>	42 <sup>a</sup>	30.33 <sup>a</sup>	22.67 <sup>a</sup>	17 <sup>a</sup>	10.33 <sup>a</sup>	0.67 <sup>ab</sup>
2	Jatropha cake	27.33 <sup>bs</sup>	41.67 <sup>b</sup>	26.33 <sup>b</sup>	19.67 <sup>b</sup>	16 <sup>b</sup>	6.67 <sup>b</sup>	0 <sup>ab</sup>
3	Mahua cake	26.67 <sup>bs</sup>	39.33 <sup>c</sup>	23.67 <sup>c</sup>	15.67 <sup>cs</sup>	14.33 <sup>cs</sup>	5.0 <sup>c</sup>	0 <sup>ab</sup>
4	Karanja cake	24.67 <sup>c</sup>	36.33 <sup>d</sup>	18.67 <sup>d</sup>	15 <sup>cs</sup>	13.67 <sup>cs</sup>	2.33 <sup>d</sup>	0 <sup>ab</sup>

\*Numbers followed by the same letter are not significantly different (P = 0.05)

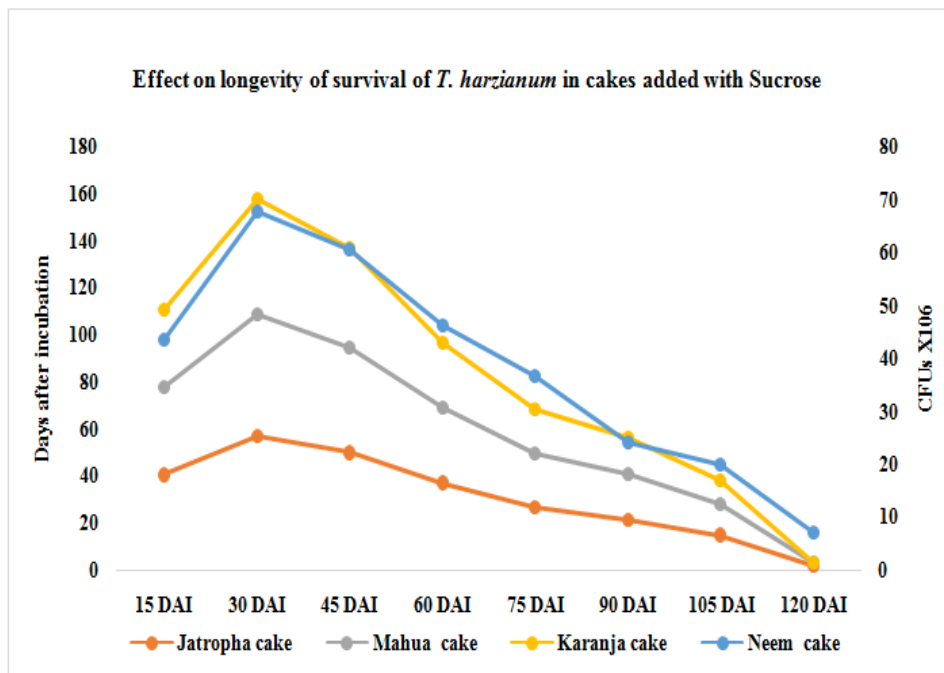
**Table 2** Screening of different oil cakes up to 120 days after incubation (DAI) added with Sucrose

S.N.	Treatments	CFUs x10 <sup>6</sup>							
		15 DAI	30 DAI	45 DAI	60 DAI	75 DAI	90 DAI	105 DAI	120 DAI
1.	Neem cake	43.67 <sup>a</sup>	67.67 <sup>a</sup>	60.67 <sup>a</sup>	46.33 <sup>a</sup>	36.67 <sup>a</sup>	24.33 <sup>a</sup>	20.00 <sup>a</sup>	7.33 <sup>a</sup>
2.	Jatropha cake	40.67 <sup>b</sup>	57.00 <sup>b</sup>	50.00 <sup>b</sup>	37.33 <sup>b</sup>	27.00 <sup>b</sup>	21.67 <sup>b</sup>	15.33 <sup>b</sup>	2.33 <sup>b</sup>
3.	Mahua cake	37.00 <sup>c</sup>	51.67 <sup>c</sup>	44.67 <sup>c</sup>	32.00 <sup>c</sup>	22.67 <sup>c</sup>	19.33 <sup>c</sup>	12.67 <sup>c</sup>	1.00 <sup>cs</sup>
4.	Karanja cake	33.00 <sup>d</sup>	49.00 <sup>d</sup>	42.00 <sup>d</sup>	27.33 <sup>d</sup>	18.67 <sup>d</sup>	15.67 <sup>d</sup>	10.33 <sup>d</sup>	0.00 <sup>cs</sup>

\*Numbers followed by the same letter are not significantly different (P = 0.05)



**Fig. 1** Screening up to 105 days of incubation into different TBOs not added with Sucrose



**Fig. 2** Screening up to 120 days of incubation into different TBOs added with Sucrose

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