

Recent Crisis in Egypt Livestock Industry due to Foot and Mouth Disease Virus

Asmaa A. Hegazy¹, Hoda A. Abd-Ellatieff¹, Emank. Bazh¹, Wael M. Goda¹ and Abdel-Rahman A. Abourawash¹

¹Department of Pathology and Parasitology, Faculty of Veterinary Medicine, Damanshour University, El-Beheira, Egypt

Corresponding author: Abdel-Rahman A. Abourawash Email: rawashaa@yahoo.com

Abstract: FMD is a highly contagious viral disease that threatens the livestock industry worldwide. The disease being endemic in many countries of Africa, Asia and South America. The disease was endemic in Egypt since 1950 and however that, the Egyptian farms have been still attacked by several outbreaks of FMDV during the last years resulted in a highly mortality rates in both young and adults. In order to investigate the morphopathological changes in animals that naturally infected and died due to FMDV infection. During 2016-2017, total of 500 pm tissues samples and 100 blood and 100 serum samples collected from 170 field samples, were analyzed. Pathological examination revealed presence of vesicular and erosive lesions developed on the tongue, interdental space of the feet, muzzle, and teats. Multi-focal areas of lympho-plasmocytic myocarditis were seen in the heart of all calves, and similar lesions were also observed in the hearts of 66 % of adults. Detailed pathological alterations in adult and young animals were described. Polymerase chain reaction (PCR) screening of tissue specimens using specific primers for FMDV was positive. The results provide a detailed description of the pathogenesis of the disease in cattle and buffalo livestock recently infected by FMDV outbreaks in Egypt.

Keywords: Foot-and-mouth disease virus; cattle; buffalo; pathological alterations, PCR

Date of Submission: 21-04-2018

Date of acceptance: 08-05-2018

I. Introduction

FMD, is a highly contagious vesicular disease, genus aphthovirus, Picornaviridae family, affecting a number of wild and domestic cloven-hoofed mammals (Samuel 2001; Grubman and de los Santos 2011). FMD is an enzootic disease, seen in most large areas of the world (Barker 1993). Vesicular lesions on the tongue, feet, snout, and teats, fever and lameness are the more characteristic lesions expressed by FMD infected animals (Donaldson 2000). The disease is notorious for its high morbidity and mortality especially in suckling animals but is not notable for high mortality rate in adults (Barker 1993). The mortality rate can exceed 50 % in young animals due to myocardial damage but it is about nearly 5 % in adult ruminants (Barker 1993). In calves, myocarditis is considered a fatal form of FMD as it is distinguished by hyaline degeneration, necrosis of muscle and can occur without developing the characteristic blister lesions noted in adult cattle (Barker 1993; Alexandersen 2003).

FMD being endemic in many countries of Africa, Asia and South America (Bayry J 2001). However, outbreaks recently occurred in countries that are normally free of FMD, including Japan, Korea, United Kingdom, France and the Netherlands (Gibbens 2001; Gibbens 2002; Muroga 2012). The disease is endemic in Egypt since 1950 (Zahran 1961), and cause a seasonal highly severe economic losses. Serotypes A and O of Foot-and-mouth disease Virus (FMDV) were the most prevalent serotypes circulating in Egypt, in addition to SAT2 serotype (EL-Bayoumy, Abdelrahman et al. 2014; Elhaig and Elsheery 2014; Abd El Moneim, Hafez et al. 2016). However, Vaccinations regime is regularly adopted by veterinary authorities; the Egyptian farms have been attacked by several outbreaks of FMDV during the last few years, which resulted in economic crisis to livestock industry as a result of highly mortality rates in both young and adult animals. So, in this study we aimed to catch the pathological alterations in cattle and buffaloes spontaneously infected with FMDV in correlation with the increased mortalities in adults; and identification of the causative FMDV strain by using RT-PCR technique based on serotype specific primers.

II. Materials and methods

2.1. Sample Collection and histopathological examination.

Clinical samples (blood, oral swabs, and post mortem tissues) were collected during FMD outbreak 2016 – 2017 from different localities in El-Beheira government. Clinical signs and all the history of diseased animals were recorded (**Table. 1**). Dead animals were immediately subjected to careful postmortem (PM) examination according to animal welfare abattoirs in Egypt and all PM findings were recorded. Suitable tissue specimens were collected from oral cavity, rumen, reticulum, omasum, abomasum, tongue, buccal tissue, dental pad, palatine tonsil, Lung, liver, spleen, heart (atria, ventricles, and interventricular septum), and lymph nodes (mandibular, parotid, retropharyngeal, tracheobronchial, mediastinal, mesenteric) for histopathological and microbiological examinations. A set of tissue specimens were collected and kept frozen at -80 °C until used for RNA extraction. Another set of the tissue specimens were immediately fixed in 10% neutral buffered formalin (2–4 days at room temperature) for histopathological examinations. Fixed specimens were routinely processed through dehydration in ascending grades of ethanol and then cleared in xylene and embedded in paraffin blocks. Paraffin sections were prepared at 4 – 5 μ thickness on glass slides. The sections were then stained with hematoxylin & eosin and histopathologically examined using the light microscope (Bancroft, Suvarna et al. 2012). Severity of gross and histopathological lesions between calves and adults were evaluated based on previously described grading systems (Cross 1998; Kleiner DE 2005; Cheng Z 2010; Gibson-Corley, Olivier et al. 2013). The grading scale included five scores: -ve=no lesions; + =mild; ++=moderate and +++=severe.

2.2. RNA extraction and cDNA synthesis

Total RNA extraction from (blood, oral swabs, and post mortem tissues) was done by using Qiagen, All Prep[®] DNA/RNA Mini kit, Germany; according to manufacturer's instructions. Total RNA samples were reverse-transcribed by using HiSenScript™ RH (-) cDNA Synthesis kit, NtRON Biotechnology, Korea, according to the manufacturer's instructions. The obtained cDNA was used for specific PCR by using specified primers for amplification of FMDV. The primers were synthesized by Sigma–Aldrich–Japan.

2.3. Conventional PCR analysis:

Conventional PCR for FMDV detection was performed using cDNA from collected samples using previously published specific primer (Reid, Ferris et al. 2000; Cottam, Haydon et al. 2006) for FMDV screening. All PCR reaction was carried out using an Eppendorf thermal cycler (**SENsQUEsTlabcyler**). The PCR amplification performed in a 25 μl volume containing 4 μl DNA, 2 μl dNTP, 1 μl of each primer (10 μmol), 2.5 μl 10× Ex Taq buffer, 0.25 μl Ex Taq polymerase (Takara, Kyoto, Japan), and 14 μl RNA, DNA free water.

The PCR condition for the universal primer that used for detection of all FMDV serotypes was as follows: 94°C for 5 min, one cycle; 94°C for 1 min, 55°C for 1 min, followed by 35 cycles at 72°C for 2 min with final extension at 72°C for 7 min, one cycle. The positive samples were used to amplify VP1 gene specific for serotype O by using the following condition: 94°C for 1 min, one cycle; 62°C for 1 min, followed by 35 cycles at 72°C for 2 min with final extension at 72°C for 7 min, one cycle.

III. Results

3.1. Clinical signs

Various clinical signs with various degrees in relation to age, species were exhibited by infected animals. Young animals in both cattle and buffalo species in comparison to the adults exhibited severe clinical signs ended by mortality (**Table. 1**). Clinical signs include; fever which ranged from (39C – 42C) with severe salivation, vesicular lesions of the mouth, feet, tongue, snout and teats. Mouth lesions vary from an elevated area of hydropic degeneration on the upper surface of the tongue to red area of sub mucosal hemorrhage oral commissar, and severe ulceration that result from ruptured vesicles are found in upper lips, tips of tongue, dental pad, and upper third of the tongue. Lameness due to the foot lesion which represented by severe vesicular lesions in the digits and coronary bands ended by severe ulceration in interdigital space. Debilitating effects, rather than high mortality rates especially in young animals were also recorded (**Table. 1**).

3.2. Gross pathology

The adult cattle and buffalo showed several gross lesions as vesicles, erosions and ulcerations in the mouth and interdigital space. Oral lesions consisted of vesicular, irregular erosions/ulcers measuring 1–4 cm in diameter usually located on the torus lingua and the anterior third of tongues and larger erosion on the gingiva and lip. Foot lesions of circumferential vesicles of the coronary band with partial undermining and detachment of the bulbs of the heel, coronary bands of all feet were eroded or ulcerated, and there was severe detachment of the soles. Similar ulcers found in ruminal pillars, abomasum and intestine. Severe hemorrhagic inflammation and

Table 1: Showing lesion scoring of clinical signs, gross pathology, histopathology and PCR results of animals naturally infected with FMD outbreak.

Criteria	Calves (2months-2 years)				Adult		
	Cattle calves		buffalo calves		cattle	buffalo	
Age	1_6 months	6months_2Y	1_6 months	6months_2Y	> 2 years	> 2 years	
Number	36	52	22	15	27	18	
Clinical signs	Fever	++	+++	+++	++	+++	+++
	Oral lesion	++	+++	+++	++	+++	++
	Foot lesion & lameness	-ve	+++	-ve	++	+++	+
	Sudden death	+++	+++	++	++	+++	++
Gross pathology	Erosion & ulcer along GIT	++	+++	+++	++	+++	++
	Myocardial necrosis	+++	+++	++	++	+++	++
	Myocardial hemorrhage	++	+++	+	++	+++	++
Histopathology of organs	Oral cavity	++	+++	+++	++	+++	++
	GIT	+++	+++	+++	++	+++	++
	Liver	+	++	++	++	+++	++
	Heart	+++	+++	+++	+++	+++	++
	Foot	-ve	+++	-ve	++	+++	++
	Lung	+++	+++	++	++	+++	++
PCR results %	+ve % by using Universal primer for FMDV	78%	67%	45%	52%	81%	44%
	+ve % by using Specific primer for serotype (O)	72%	60%	42%	45%	75%	41%

-ve = no lesions; += mild; ++ = moderate; +++ = severe.
Sudden death + = 25%, ++ = >25-50%, +++ = >50- 75%

myocardial hemorrhage range from small petechial to sever hemorrhage with various degree of myocardial necrosis was also observed grossly (Fig 1A, B, C, and D).

Young calves have all the above mentioned lesions in oral, digestive and feet but with increased severity, represented by the increase of the size and distribution of vesicular, erosive, ulcerative lesions in that sites. Pronounced myocardial hemorrhage with the characteristic tiger heart appearance were the most pathognomonic gross lesions in all of the dead calves in the form of yellowish to grayish streaking bands in myocardium (**Table. 1**).

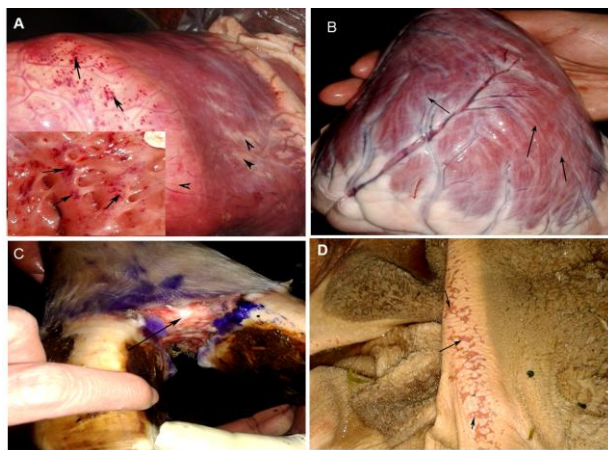


Fig.1. (A) Heart of calf 6 months old naturally infected by FMD, showing petechial hemorrhage at epicardium (arrows) with myocardium degeneration (arrowheads), the inset picture showing petechial hemorrhage in endocardium (arrows). (B) Heart of 13 months of age showing several yellow streaks (arrows) of myocardium degeneration and necrosis “tiger heart”. (C) Severe ulceration with hemorrhage of hoof (arrows). (D) Focal area of ulcers of ruminal pillars (arrows).

3.3. Histopathological findings:

Histopathological examination of young and adult livestock was recorded. Oral cavity, foot (coronary band), omasum, and abomasum and rumen lesions revealed typical FMD lesions consisting of vesicle formation at the stratum spinosum, characterized by focal hydropic and ballooning degeneration ended by vesicular formation of varying sizes, edema, cell necrosis, and focal infiltrates of neutrophils (Fig 2 A). Intraepithelial bullae formation with superficial epithelial caps was also detected. Acanthosis with hyperkeratosis of overlying and adjacent epithelium. Erosions were seen with detached keratinocytes, amorphous eosinophilic material, neutrophils. Basal epithelium was usually preserved, but some lesions extended through the basement membrane with marked mixed inflammatory cell infiltrates in the submucosa or dermis with subsequent sloughing of necrotic tissue leading to ulcers formation. Focal myositis in the tongue, oral skin, omasum, abomasum and rumen characterized by myofiber hyaline degeneration and zenker's necrosis accompanied by sarcolemmal proliferation. The heart of adult animals showed mild to severe myocarditis (Fig 1 B,C,D), while the heart of the young calves is severely affected especially in younger ages in the form of moderate to severe myocarditis (Table. 1). Severe degeneration and necrosis of myocardium with complete lysis of some muscle fibers and replacement of these fibers by inflammatory cells (Fig 2 A), were also detected in many examined cases. Inflammatory edema with pronounced infiltration with lymphocytic cell and some macrophages, myocardial hemorrhage and vasculitis were also present. The liver showed mild to moderate hydropic degeneration of hepatocytes with varying degree of coagulative necrosis and periportal inflammation (Fig 2 B,C). The lung showed variable degree (ranged from mild to severe) of various types of pneumonia (lymphocytic, hemorrhagic, serous, fibrinous pneumonia and bronchopneumonia). All that forms of pneumonia were seen alone or mixed with each others as serohemorrhagic or serofibrinous (Fig 2 D).

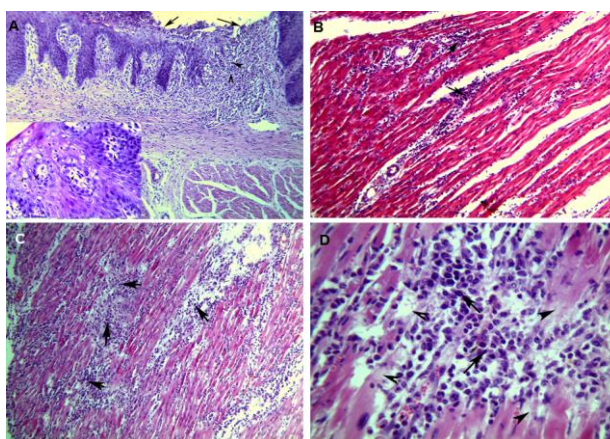


Fig.2.(A) ulcerative dermatitis of adult cow naturally infected by FMDV (arrows) with leukocytic cell infiltrations (arrowheads), the inset picture showing leukocytic cell infiltration. H&E, Bar=100 µm. (B) Mild myocarditis of myocardium with mild lymphocytic cell aggregations (arrows). H&E, Bar=100 µm. (C) Moderate myocarditis with increased number of lymphocytic cell aggregation (arrows) between degenerated muscles H&E, Bar=100 µm. (D) Higher magnifications of (C)

showing lymphocytes and mononuclear cells (arrows) with necrosed myocardial muscle (arrowheads) H&E, Bar=50 μ m.

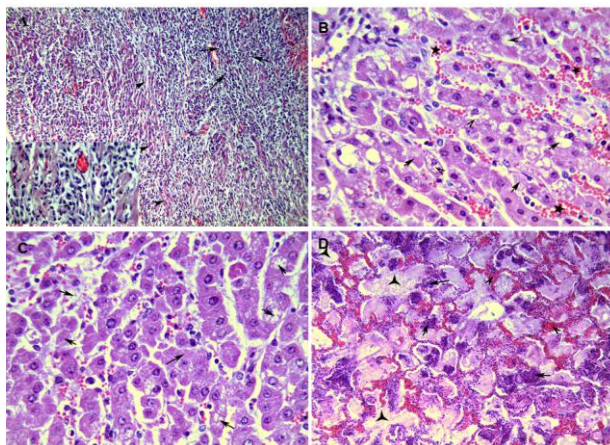


Fig.3. (A) Severe myocarditis with heavy aggregation of lymphocytes and mononuclear cells (arrows) which replaced the necrosed myocardial muscle (arrowheads), inset picture showing leukocytic cells and necrosed muscle.H&E, Bar=100 μ m.(B) Liver of naturally infected buffalo by FMDV showing hydropic degeneration (arrowheads) with congestion of blood sinusoids(black stars) and mild coagulative necrosis of hepatocytes (arrows).H&E, Bar=50 μ m. (C) Liver of cow naturally infected by FMDV showing focal areas of coagulative necrosis (arrows).H&E, Bar=50 μ m. (D) Lung of naturally infected calve by FMDV showing congestion of perialveolar capillaries (arrowheads) with various forms of pneumonia as lymphocytic pneumonia (arrows) and fibrinous pneumonia (black stars).H&E, Bar=100 μ m.

3.4. RT-PCR results

Tissues, serum and Blood samples were analyzed by PCR for the presence of viral RNA. The percentage of positivity in each species was summarized in (Table 1).

IV. Discussion

a wide variety of cloven-hoofed domesticated and wild animal species were naturally infected by FMDV, resulting in foot and mouth disease, an acute febrile disease characterized by vesicular lesions of the oral cavity and upper digestive tract, including tongue, soft and hard palate, inner sides of cheeks, feet and teats (Grubman 2004). From previous reports, FMD usually resolves without the need for treatment despite of the extensive lesions caused by the virus in a short duration in the individual animal and is seldom lethal in adults (Arzt 2011). However, recently in the last outbreaks, FMD considered one of the most feared livestock diseases not only due to the highly contagious nature, or wide dissemination but due to the adverse significant economic impact on the livestock industry represented by a highly mortality rates in both young and adults. For that reasons, we tried to provide a comprehensive overview of FMD pathological alterations in young calves, cattle, and buffalo spanning from the earliest studies to recently acquired insights that lead to the high mortality rates in both young and adults in the last outbreaks. In addition to characterization of the field isolates of FMDV and its correlation with the apparently increased virulence of virus based upon increased mortalities in adults with pathological findings.

Hydropic degeneration with vesicle formation followed by erosion and ulcers of various epithelial sites including the mouth, feet, teats, omasum, abomasum and pillars of the rumen were observed not only during the viraemic phase but it often extends beyond the period of viraemia (Alexandersen 2005; Arzt 2009). Ruptured vesicles in oral cavity, feet, teats and skin were the most prominent gross lesions seen in the dead animals at necropsy. The degree of severity based upon the size and dissemination of lesions varied from mild to moderate in adults to high severity in young calves. Gross lesions were accompanied with the microscopic lesions exhibited by the affected animals in most of the cases (Seibold 1963; Arzt 2009). Absence of the gross lesions not indicative of absence of the virus as the virus can be detected even in the absence of macroscopic lesions (Brown 1995; Murphy and Alexandersen 2010).

Death due to FMD outbreaks is a fairly common feature of FMD epizootics in young livestock, as calves, piglets and lambs, (Alexandersen 2005). Death is often attributed to myocarditis (Donaldson 1984; Gulbahar and Kabak 2007). However, the death in adults is accompanied with degeneration of the myocardium which is a rare manifestation known as 'malignant FMD', (Shimshony 1986; Arzt and Rieder 2011) which became not a rare nowadays. In our study we found that FMD caused high mortality in calves in addition to

increased mortality percentage in adults in contrary to earlier reports as FMD didn't result in high mortality in adult animals, than neonates (Alexandersen 2003).

RT-PCR analysis revealed positive of FMDV from all samples collected from animals showing clinical signs. Histological examinations revealed Zenker's necrosis with myositis in the examined muscles of tongue, cheeks, omasum, abomasum and ruminal muscles as previously reported (Shimshony 1988), which is inconsistent with myotropic nature of FMDV (Arzt and Rieder 2011). The heart showed myocardial necrosis with hyaline degeneration, and intense mononuclear cell infiltration (Gunes 2005; Karapinar 2010). In contrary to earlier reports, many myocarditis cases have been detected in calves aged from 2 months to 2 years old and in some adult animals aged more than 2 years (Gulbahar 2007; Karapinar 2010; Aslani 2013; Kaya 2013). In which most of myocarditis were reported in calves aged one week to three months old. Several types of Pneumonia (serous, hemorrhagic, fibrinous, serohemorrhagic, serofibrinous and serohemorrhagic) and bronchopneumonia were predicted especially in young calves from 1 month to 2 years old age, with severe myocarditis and myocardial necrosis and these results were supported with the detection of high quantities of viral RNA, infectious, and viral antigen in pulmonary tissues either in the viraemic or priviraemic stage (Burrows 1981; Arzt 2010).

In conclusion, severe myocarditis with myocardial degeneration and necrosis, were the most characteristic pathological findings of FMD during the last outbreak of FMDV infection in EL-Beheira governorate/ Egypt. Severe pneumonia was also not uncommon findings. The risk of myocarditis in FMD is not directly related only to the age but could also be attributed to increased pathogenicity of the recently isolated field strain circulating among cattle in Egypt or emerging of new viral strains due to legal and illegal trade in animals and animal products importing.

V. Reference

- [1] Abd El Moneim, A. A., M. H. Hafez, et al. (2016). "Pathological and Molecular Investigations on Foot and Mouth Virus Outbreaks Among Cattle Herds in Dakahlia Governorate, Egypt." *Zagazig Veterinary Journal (Zag. Vet. J.)* **44**(2).
- [2] Alexandersen, S., Z. Zhang, A. I. Donaldson, A. J. M. Garland (2003). "The pathogenesis and diagnosis of foot-and-mouth disease." *J. Comp. Pathol* **129**, 1-36.
- [3] Alexandersen, S. M., N (2005). "Foot-and-mouth disease: host range and pathogenesis." *Curr Top Microbiol Immunol* **288**, 9-42.
- [4] Arzt, J., Baxt, B., Grubman, M. J., Jackson, T., Juleff, N., Rhyan, J., and E. Rieder, Waters, R. & Rodriguez, L. L. (2011). "The pathogenesis of foot-and-mouth disease II: viral pathways in swine, small ruminants, and wildlife; myotropism, chronic syndromes, and molecular virus-host interactions." *Transbound Emerg Dis* **58**, 305-326.
- [5] Arzt, J., D. A. Gregg, A. Clavijo, and L. L. Rodriguez, (2009). "Optimization of immunohistochemical and fluorescent antibody techniques for localization of Foot-and-mouth disease virus in animal tissues" *J. Vet. Diagn. Invest* **21**, 779-792.
- [6] Arzt, J., J. M. Pacheco, and L. L. Rodriguez, (2010). "The early pathogenesis of foot-and-mouth disease in cattle after aerosol inoculation: identification of the nasopharynx as the primary site of infection." *Vet Path* **47**, 1048-1063.
- [7] Arzt, J., Juleff, N., Zhang, Z. & Rodriguez, L. L. (2011). "The pathogenesis of foot-and-mouth disease I: viral pathways in cattle." *Transbound Emerg Dis* **58**, 291-304.
- [8] Aslani, M. R., M. Mohri, A. R. Movassaghi (2013). "Serum troponin I as an indicator of myocarditis in lambs affected with foot and mouth disease." *Vet. Res. Forum* **4**, 59-62.
- [10] Bancroft, J. D., K. S. Suvarna, et al. (2012). *Bancroft's Theory and Practice of Histological Techniques E-Book*, Elsevier Health Sciences.
- [11] Barker, I. K., A. Van Dreumel, N. Palmer (1993). "The alimentary system. In: Pathology of Domestic Animals, 4th ed. (Jubb, K. V. F., P. C. Kennedy, N. Palmer, Eds.) Academic Press, San Diego, CA" **pp.141-144**.
- [12] Bayry J, K. S. (2001). "Foot and mouth disease: a revised policy is required." *J Clin Microbiol* **39**:3808.
- [13] Brown, C. C., H. J. Olander, and R. F. Meyer, (1995). "Pathogenesis of foot-and-mouth disease in swine, studied by in-situ hybridization" *J. Comp. Pathol* **113**, 51-58.
- [14] Burrows, R., J. A. Mann, A. J. Garland, A. Greig, and D. Goodridge, (1981). "The pathogenesis of natural and simulated natural foot-and-mouth disease infection in cattle." *J. Comp. Pathol* **91**, 599-609.
- [15] Cheng Z, D. D., Zhao L, Wang HL, Doherty TM, Bresee C, Frykman PK (2010). "Murine model of Hirschsprung-associated enterocolitis. I: Phenotypic characterization with development of a histopathologic grading system. ." *J Pediatr Surg* **45**:475-482. [PubMed: 20223308].
- [16] Cottam, E. M., D. T. Haydon, et al. (2006). "Molecular epidemiology of the foot-and-mouth disease virus outbreak in the United Kingdom in 2001." *Journal of Virology* **80**(22): 11274-11282.
- [17] Cross, S. (1998). "Grading and scoring in histopathology." *Histopathology* **33**(2): 99-106.
- [18] Donaldson, A. I., Ferris, N. P. & Wells, G. A. (1984). "Experimental foot-and-mouth disease in fattening pigs, sows and piglets in relation to outbreaks in the field." *Vet Rec* **115**, 509-512.
- [20] Donaldson, A. I., F. Sellers (2000). "Foot-and-mouth disease. In: Diseases of Sheep. (Martin, W. N., I. D. Aitken, Eds) " Blackwell Science, Oxford, United Kingdom" **pp. 254-258**.
- [21] EL-Bayoumy, M. K., K. A. Abdelrahman, et al. (2014). "Molecular Characterization of Foot-and-Mouth Disease Virus Collected from Al-Fayoum and Beni-Suef Governorates in Egypt." *Global Veterinaria* **13**(5): 828-835.
- [22] Elhaig, M. M. and M. N. Elsheery (2014). "Molecular investigation of foot-and-mouth disease virus in domestic bovids from Gharbia, Egypt." *Tropical animal health and production* **46**(8): 1455-1462.
- [23] Gibbens, J. C., and J. W. Wilesmith (2002). "Temporal and geographical distribution of cases of foot-and-mouth disease during the early weeks of the 2001 epidemic in Great Britain" *Vet. Rec* **151**:407-412.
- [24] Gibbens, J. C., C. E. Sharpe, J. W. Wilesmith, L. M. Mansley, E. Michalopoulou, J. B. M. Ryan, and M. Hudson. (2001). "Descriptive epidemiology of the 2001 foot-and-mouth disease epidemic in Great Britain: the first five months" *Vet. Rec* **149**:729-743.

- [25] Gibson-Corley, K. N., A. K. Olivier, et al. (2013). "Principles for valid histopathologic scoring in research." *Veterinary pathology* **50**(6): 1007-1015.
- [26] Grubman, M. J. and T. de los Santos (2011). *Aphthovirus*. The Springer Index of Viruses. C. Tidona and G. Darai. New York, NY, Springer New York: 1281-1286.
- [27] Grubman, M. J. B., B. (2004). "Foot-and-mouth disease." *Clin Microbiol Rev* **17**, 465-493.
- [28] Gulbahar, M. Y., Davis, W. C., Guvenc, T., Yarim, M., Parlak, U. & and Y. B. Kabak (2007). "Myocarditis associated with foot-and-mouth disease virus type O in lambs." *Vet Pathol* **44**, 589-599.
- [29] Gulbahar, m. Y., w. C. Davis, t. Guvenc, m. Yarim, u. Parlak, y. B. Kabak (2007). "Myocarditis associated with foot-and-mouth disease virus type o in lambs." *Vet. Pathol* **44**, 589-599.
- [30] Gunes, v., h. M. Erdogan, m. Cital, k. Ozcan. (2005). "Assay of cardiac troponins in the diagnosis of myocardial degeneration due to foot-and-mouth disease in a calf." *Vet. Rec* **156**, 714-715.
- [31] Karapinar, t., d. O. Dabak, t kuloglu, h. Bulut (2010). "High cardiac troponin I plasma concentration in a calf with myocarditis " *Can. Vet.* **151**, 397-399.
- [32] Kaya, a., s. Kozat, c. Ozkan, s. Yildirim, y. Akgul, o. Akgul (2013). " Serum homocysteine levels in calves with foot and mouth disease." *JAVA* **12**, 1357-1361.
- [33] Kleiner DE, B. E., Natta MV, Behling C, Contos MJ, Cummings OW, Ferrell LD, Liu Y, Torbenson MS, Unalp-Arida A, Yeh M, McCullough AJ, Sanyal AJ. (2005). " for the Nonalcoholic Steatohepatitis Clinical Research Network. Design and Validation of a Histological Scoring System for Nonalcoholic Fatty Liver Disease. ." *Hepatology* **41**:1313-1321.
- [34] Muroga, N., Hayama, Y., Yamamoto, T., Kurogi, A., Tsuda, T., Tsutsui, T., (2012). "The 2010 foot-and-mouth disease epidemic in Japan " *J. Vet. Med. Sci* **74**, 399-404.
- [35] Murphy, C., J. B. Bashiruddin, M. Quan, Z. Zhang, and S. and Alexandersen (2010). "Foot-and-mouth disease viral loads in pigs in the early, acute stage of disease." *Vet. Rec* **166**, 10-14.
- [36] Reid, S. M., N. P. Ferris, et al. (2000). "Primary diagnosis of foot-and-mouth disease by reverse transcription polymerase chain reaction." *Journal of Virological Methods* **89**(1-2): 167-176.
- [37] Samuel, a. R., n. J. Knowles (2001). "Foot-and-mouth disease virus: cause of the recent crisis for the UK livestock industry " *Trends. Genet* **17**, 421-424.
- [38] Seibold, H. R. (1963). "A revised concept of the lingual lesions in cattle with foot-and-mouth disease." *Am. J. Vet. Res* **24**, 1123-1130.
- [39] Shimshony, A. (1988). " Foot and mouth disease in the mountain gazelle in Israel 7:917-923." *Rev Sci Tech Off Int Epiz.*
- [40] Shimshony, A., Orgad, U., Baharav, D., Prudovsky, S., Yakobson, B., Bar Moshe, B. & Dagan, D. (1986). " Malignant foot-and-mouth disease in mountain gazelles." *Vet Rec* **119**, 175-176.
- [41] Zahran, G. (1961). "Foot-and-mouth disease virus. I. Propagation of 3 immunologic types of virus in chicks." *American Journal of Veterinary Research* **22**: 518-526.
- [42]