

A Review on the Phytoconstituents and Related Medicinal Properties of Plants in the Asteraceae Family

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Abstract: The Asteraceae family of plants is a widely distributed medicinal plant throughout the world and has been used since ancient time. Members of this family have been used traditionally as astringent, antipyretic, anti-inflammatory, hepatoprotective, diaphoretic in fevers, smooth muscle relaxant, nerve tonics, laxatives and for the treatment of wounds, bleedings, headache, pains, spasmodic diseases, flatulence, dyspepsia, dysentery, lumbago, leucorrhoea, hemorrhoids, gangrenous ulcer and disorders causing cachexia. Phytochemical investigations of the Asteraceae family have revealed that many components from this family are highly bioactive. There are many reports on the mentioned herbal effects. Although, the medicinal properties of plants belonging to the Asteraceae family are recognized worldwide, there is no review article mainly about the structures of the phytochemical constituents of the plant of the asteraceae family. The present paper reviews the medicinal properties alongside with peculiar phytoconstituent of various plants belonging to the Asteraceae family. Various medicinal effects of these plants may be due to the presence of a broad range of secondary bioactive metabolites such as flavonoids, phenolic acids, coumarins, terpenoids (monoterpenes, sesquiterpenes, diterpenes, and triterpenes) and sterols which have been frequently reported from the Asteraceae family.

Keywords: Antioxidant, Asteraceae, bioactive compounds, hepatoprotective, phytochemical

I. Introduction

Asteraceae plants are distributed throughout the world and most common in the arid and semi-arid regions of subtropical and lower temperate latitudes. These plants typically have hairy and aromatic leaves and flat clusters of small flowers on the top of the stem. Since these flowers have various colors, a number of species are popular garden plants. The majority of the Asteraceae family members are as the medicinal plants which have therapeutic applications. These comprise Achillea, Carthamus, Chromolaena, Emilia, and Pluchea.

There are no review papers on the different aspects of asteraceae as a noteworthy and medicinal family. Recently, Saaidnia et al published a review article mainly about the structures of phytochemical constituents and a brief section of biological properties of Achillea species. Literature reviews show that there are many reports on pharmacological, immunological, biological and other therapeutic activities of these valuable herbs belonging to the asteraceae family which are reviewed in this article.

1.1 Phytochemical Screening

Preliminary phytochemical screening revealed the presence of glycoside, flavonoids, tannins, mucilage, carbohydrate and reducing sugar in *Tridax procumbens* Linn. In *Baccharis racemosa*, the main phytoconstituents were sabinene, β -pinene, myrcene, limonene, δ -cadinene, nerolidol, viridifloral, α -muurolol and α -cadinol. Also, *Baccharis linearis* phytoconstituents were α - and β -pinene, myrcene, limonene, bicyclogermacrene, δ -cadinene, caryophyllene oxide, cubenol and α -cadinol. Yield on the *Baccharis ovata* oil was 2.82% and the main constituents were α -thujene, α - and β -pinene, sabinene, myrcene, limonene and terpinen-4-ol (figure 1) [1]. Leaves from another Argentinian *Baccharis* species, *Baccharis stenelia* were collected from the Lanos region of La Rioja for analysis of the essential oil composition. Spathulenol (figure 2) was the major component (29-34%) of the oil [2]. Characteristic diterpenic constituents from the *Baccharis* genus are the neo-clerodane type diterpenes, although kaurane and labdane derivatives have also been isolated. From *Baccharis gaudichaudiana*, new clerodane diterpenoids, gaudichanolides A and B (figure 3), and a new ent-clerodane diterpene named bacchariol (Figure 4) were isolated [3]. The first anti-spasmodic flavonoids, cynaroside I (figure 4) and cosmosiin II (figure 5) were isolated from *Achillea millefolium* L. [4].

The main constituents of the safflower *Carthamus tinctorius* L. are carthamin and carthamidin. Other constituents are safflor yellow, arctigenin, tacheloside, N-feruloyltryptamine, N-feruloylserotonin, steroids, flavonoids, and polyacetyles. Carthamin is responsible for producing water-insoluble red dye and carthamidin for water-soluble yellow colour dye [5]. The principle colouring component of marigold flower *Tagetes erecta* is lutein, a fat-soluble carotenoid, which is responsible for the yellow to orange colour of the dye [6].

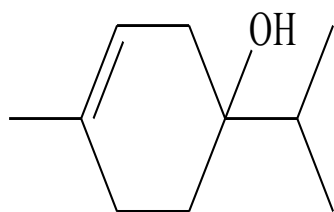


Fig. 1 Structure of terpinen-4-ol.

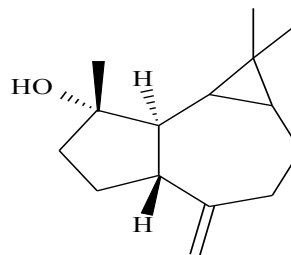


Fig. 2 Structure of spathunol

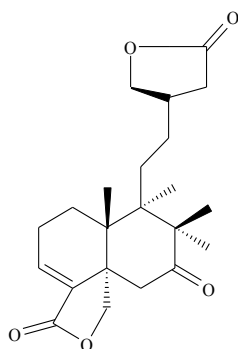


Fig.3 Gaudichanolide

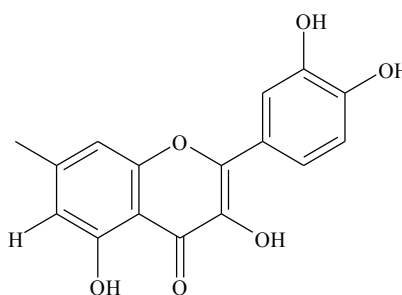


fig. 4 Cyniriside 1

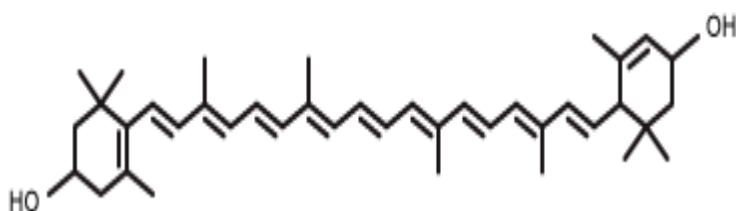


Fig. 6 structure of lutein

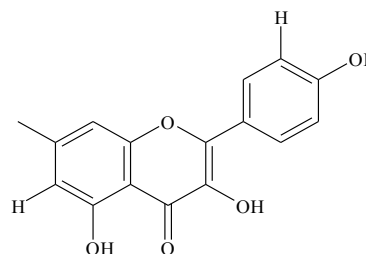


Fig 5 cosmosiin

II. Medicinal Properties Of Asteraceae Family

2.1. Wound healing activity

Aqueous extract of the flowers of *Achillea kellerensis* applied topically, has shown significant wound healing activity in rats. The wound sizes of the test compared to control groups were reduced faster [7]. This is due to the activity of flavonoids. Extracts from the leaves of *Chromolaena odorata* have been shown to be beneficial for treatment of wounds. In traditional usage, the leaf is ground into a paste and is applied topically on affected places to heal wounds [8]. The aqueous extract and the decoction from leaves of this plant have been used throughout Vietnam for the treatment of soft tissue wounds and burns for decades. A product made from *Chromolaena* named eupolin have already been licensed for use in Vietnam for soft tissue burns and wounds.

2.2. Protective activity

The protective activity of natural antioxidants in biological systems has received attention. Some medicinal plants have proved free radical scavenging or antioxidant activities [9]. The infusions of *Achillea* species were tested on antioxidant enzyme systems of erythrocytes and *Achillea falcata* L. was the most effective one against CAT (catalase), GPx (glutathione peroxidase) and SOD (superoxide dismutase) enzyme systems of erythrocytes. Among the plant infusions, highest activities on leucocyte enzymes were by *Achillea crithmifolia* and *Achillea nobilis* L. subspecies *neilreichii* on CAT, by *Achillea millefolium* subspecies *annonica* on SOD, by *Achillea retifolia* Willd. on GPx and by *Achillea nobilis* subspecies *sipylea* on LPO (lactoperoxidase). Therefore, *Achillea* species may be of potential sources of natural antioxidants for treatment or prevention of related diseases [10].

The influence of the extracts of *A. alexandri-regis* on hydroxyl and superoxide radicals' quantity in different *in vitro* systems have been determined. The ethyl acetate extract exhibited hydroxyl radical scavenging activity in all tested biological systems (liver homogenate, hemolyzed blood, serum and post mitochondrial liver fraction), whereas butanol extract reduced hydroxyl radicals significantly only in the post mitochondrial liver fraction (a

homogenate of liver cells remaining after sedimentation of the mitochondrial fraction by centrifugation). Both extracts affected only hemolysed blood [11]. The hydroalcoholic extract of *A. santolina* L. was studied on various in vitro antioxidative systems and it has been reported that the extract prevented formation of thiobarbituric acid reactive substances in Fe²⁺-ascorbate induced lipid peroxidation in rat liver tissue. Free radical induced protein oxidation has also been suppressed significantly by high concentration (1000 µg/ml) of the extract [12]. Ethanol extracts of eight wild samples of *A. ligustica* and one sample of cultivated *A. millefolium* were evaluated for radical scavenging activities including DPPH test. The TEAC (the concentration of a Trolox solution having an antioxidant capacity equivalent to that of the diluted hydroalcoholic extract) were in the range of 4.18 and 12.3 mM. The ability of the extracts to inhibit non-enzymatic lipid peroxidation using an in vitro system of linoleic acid oxidation has been investigated. Five of the nine extracts had a protective effect at the lowest tested amount (5 µg). Protection on CaCo-2 intestinal cells against TBH-induced toxicity was also investigated and two of the tested ethanolic extracts of *A. ligustica* showed protection against the oxidative stress. The antioxidant capacity and cytoprotective activity of *A. collina* infusions against oxidative stress were investigated by chemical (DPPH and Folin-Ciocalteu assay) and biological assays (in vitro model of cytotoxicity and lipid peroxidation in PC12 cells line) and it has been shown that the infusions of leaves had the highest antioxidant and cytoprotective activity, where antioxidant capacity was significantly correlated with the total phenolic content but not with the cytoprotective profile [13].

2.3. Estrogenic activity

A crude extract of the aerial parts of *A. millefolium* has shown estrogenic activity [14]. Evaluation of the isolated and identified compounds from this plant indicated that luteolin and apigenin were the most important estrogenic compounds among tested compounds. Many phytoestrogens appear to have a stronger binding affinity with β estrogen receptors than estradiol [15].

2.4. Anti-diabetic activity

The effect of *A. santolina* (hydro alcoholic extract) on blood glucose level, serum NO (nitric oxide) concentration and the oxidative stress in rat pancreatic tissue have been evaluated [16]. *Baccharis floribunda* H.B.K. (Niquitao) is used in Venezuela as decoctions or infusions of leaves and stems for skin infections, diabetes and rheumatism. *Baccharis gaudichaudiana* D.C. (*Chilcamelosa*) is used in Paraguay as a folk medicine for the treatment of diabetes and as a tonic and for the relief of gastrointestinal ailments [2].

2.5. Antispermatic effect

Ethanolic (intraperitoneally) and hydroalcoholic extracts (orally) of *A. millefolium* were administered to Swiss mice to evaluate the effect on spermatogenesis. The extract treated animals had an increased number of metaphases in the germ epithelium which should be due to substances stimulating cell proliferation [17].

2.6. Antioxidant activity

The nitric oxide scavenging activity of the *Chromolaena odorata* extract was demonstrated. Quantitative determination of the total phenolic content shows that the extract contains an appreciable amount of phenolic compounds and may be responsible for the observed antioxidant potential [18].

The fresh juice and the methanolic extract of *Emilia sonchifolia* had potent inhibitory effects on hydroxyl radical formation and superoxide radical generation in vitro. The results indicate that pre-treatment with *Pluchea lanceolata* attenuates cadmium chloride induced oxidative stress and genotoxicity by altering antioxidant enzymes and reducing chromatid breaks and micronuclei formation [19]. Extracts from *Pluchea indica* were screened for flavonoid content, total phenolics, and antioxidant activity. *Pluchea indica* Less. Extracts inhibited linoleic acid oxidation and had the DPPH, ABTS, and ferric cyanide antioxidant capacities. Therefore, the plant may contribute to dietary antioxidant intake [20]. The aqueous ethanol extracts of *Pluchea arabica* showed the inhibition of DPPH radical at 89–93%, after 15 min of incubation at a test concentration of 50 g/ml. The total antioxidant capacity as gallic acid equivalents of 1790 mg/g of ethanol extracts were obtained for *P. crispata* in the phosphomolybdenum assay [21].

2.7. Trypanocidal Activity

Four compounds active against the epimastigote forms of *Trypanosoma cruzi*, the causative agent of Chagas' disease, were isolated from the hexane extract of *Pluchea quiroc* L. (Compositae). The flavone casticin, identified on the basis of its spectroscopic characteristics [22].

2.8. Antiulcer activity

The methanolic fraction of root extract was found to possess significant antiulcer activity in different experimental animal models. The extract also afforded significant protection to chemically-induced duodenal

lesion in guinea pigs [23]. Significant enhancement of healing process in acetic acid-induced chronic gastric lesions was also observed in the *P. indica* extract-treated animals [20]. *Baccharis grisebachii*, the aerial parts of this shrub, in the Andean provinces in Argentina, are recommended in infusion to treat gastric ulcers, as a digestive, local antiseptic and cicatrizing [2]. Reviewing literature reveals that *A. millefolium* showed effectiveness in protecting the gastric mucosa against acute gastric lesions induced by ethanol and indomethacin and in healing chronic gastric lesions induced by acetic acid ($ED_{50} = 32 \text{ mg/kg}$, orally) [24]. Oral administration (30, 100 and 300 mg/kg) of the hydroalcoholic extract inhibited ethanol-induced gastric lesions by 35, 56 and 81%, respectively. Oral treatment with this extract (1 and 10 mg/kg) reduced the chronic gastric ulcers induced by acetic acid by 43 and 65%, respectively, and promoted significant regeneration of the gastric mucosa after ulcer induction denoting. The effects of aqueous ethanol extract of *A. wilhelmsii* on rat's gastric acid output in basal, vagotomized (VX), and vagal-stimulated conditions have been investigated. The extract showed a reduction in the acid output in vagal-stimulated condition at doses of 1 and 2 mg/kg, which were not statistically significant [25].

2.9. Gastro intestinal disease

Emilia sonchifolia root juice is used to treat diarrhea by the Nepalese, the Chinese make use of the leaf tea to treat dysentery and in Africa it is consumed as vegetable for the treatment of stomach ailments [26]. *Baccharis conferta* is used in the region of Veracruz, Mexico, to treat stomachaches. It is also recommended for use as a laxative and for stimulating urination [2]. *Baccharis gaudichaudiana* D.C. (*Chilcamelosa*) is used in Paraguay as a folk medicine for the treatment of diabetes and as a tonic and for the relief of gastrointestinal ailments. *Baccharis heterophylla* is ethnomedically used in the state of Queretaro, Mexico, for alleviating gastrointestinal disorders. The plant is used in the form of infusions or decoctions of the aerial parts. *Baccharis vaccinioides* (Chilco) is one of the most popular remedies from the Chiapas region of Mexico for gastrointestinal disorders. The plant is used in the form of infusions or decoctions of the whole plant [2].

2.10. Cytotoxicity effect

The main constituents are safflower (*Carthamus tinctorius* L.) carthamin and carthamidin. The chief constituent carthamin has uterine stimulating, coronary dilating and hypotensive properties. It also has the cytotoxic, antigenic and anti-platelet activity [27]. There are some reports about the anti-proliferative activity of the isolated constituents from *A. falcata* and *A. clavennae* L. These compounds have been found to decrease keratinocyte cell viability significantly. Statistical analysis confirmed an enhanced potency of the β -OH isosecotanaphthalide over the α , β -OH diastereoisomeric mixture. The enhancement of the lipophilicity of the molecule resulted in the highest potency [28]. The aerial part of *A. clavennae* was used for isolation of the phytoconstituents and the antiproliferative activity of the compounds was tested to HeLa, K562 and Fem-X human cancer cell lines. Guaianolides, 9 α -acetoxycartecanin and apressin showed significant cytotoxic effects in all tested cell lines. A bisabolene, inducumenone exhibited a moderate activity. The most active compound was a flavonol, centaureidin, which was already known as cytotoxic agent [29]. Cytotoxicity of the *Chromolaena* extract was also studied. These studies have demonstrated the presence of some compounds in the extract such as acetin (5, 7-dihydroxy-4'-methoxy flavone) and luteolin (5, 7, 3', 4'-tetrahydroxyflavone) which expressed activity against human small cell lung cancer and human breast cancer [30].

2.11. Immunosuppressive activity

The immunosuppressive potential of 50% ethanolic extract (PL) of *Pluchea lanceolata* and its bioactive chloroform fraction (PLC) was investigated with basic models of immunomodulation. The findings revealed that *P. lanceolata* causes immunosuppression by inhibiting Th1 cytokines [31]. The aqueous extract of *A. talagonica* was studied on humoral antibody responses in BALB/c mice and albino rabbits. Intraperitoneal administration of the extract to mice, prior to immunization with sheep red blood cells, resulted in a significant dose dependent decrease in haemagglutinating antibody (HA) titer in rabbits after intrascapular injection of the extract, a significant decrease in typhoid-H antibody (anti-HD) titer was found, but no change was observed in secondary response [32]. Methanol and aqueous methanol (80% and 50% v: v) extracts of *A. talagonica* have been examined to find its immunosuppressive components. Guided by anti-SRBC (sheep red blood cells) assay, active principles were isolated by chromatographic methods and identified as choline, quercetin and caffeoylglucoside. These compounds compared to the control groups decreased anti-SRBC titer significantly. Alongside these compounds, 3'-methoxy luteolin and proline has been also reported from this plant [33]. Methanol extract and some other fractions of *A. millefolium* were studied on humoral immunity in BALB/c mice by microhaemagglutination test. Only two fractions showed a significant decrease in the anti-SRBC titer of mice. The immunological properties may be related to presence of glycosylated derivatives of caffeic acid, because caffeic acid glucoside was isolated and identified from the active fractions. Some known compounds including, luteolin 7-O-glucoside and apigenin 7-O-glucoside have also been reported from this species [34]. Effects of the essential oils of *A. talagonica* and *A. millefolium* have been studied on humoral immune

responses in BALB/c mice. The oil isolated from *A. millefolium* ssp. *millefolium* possessed a high percentage of sesquiterpenes (55.4%) in which bisabolol was the main compound. The volatile oil of *A. millefolium* decreased the anti-SRBC antibody titer, but the oil of *A. talagonica* was not effective. High percentage of sesquiterpenes and presence of proazulene in *A. millefolium* together with the lack of these compounds in *A. talagonica* could account for the different immunological effects of these plants [35].

2.12 Biological effects

Ethyl acetate extract of *A. talagonica* showed toxicity in BST (brine shrimp lethality test) and on the basis of results only 5-hydroxy 3', 4', 6, 7-tetra methoxy flavone XXVII (Scheme 1) showed toxicity (LC₅₀=15 µg/ml) against *Artemia salina* larvae [33]. It is reported that the essential oil of *A. biebersteinii* exhibited antimicrobial activity against 8 bacteria, 14 fungi and one yeast namely *C. albicans*, whereas methanolic extract was inactive [36]. The antimicrobial activity of the essential oil of *A. ligustica* was evaluated by the broth micro-dilution method on 6 microbial strains and it showed to be effective against *Streptococcus mutans* [36]. In another report, antibacterial activity of the extracts (hexane: ether: methanol = 1:1:1) of the aerial parts of *A. clavennae*, *A. holosericea* Sm., *A. lingulata* and *A. millefolium* were evaluated against five bacteria (*S. aureus*, *E. coli*, *K. pneumoniae*, *P. aeruginosa* and *Salmonella enteritidis*) and two fungi (*A. niger* and *C. albicans*) and it was found that the extracts of all four species possessed a broad spectrum of antimicrobial activity against all tested strains [37].

Activity of nineteen Jordanian plants against multidrug-resistant *E. coli* has been reported. The methanolic extract of *A. santolina* (one of 19 species) was combined with antibiotics of different classes (chloramphenicol, neomycin, doxycycline, cephalexin and nalidixic acid) and tested against both the standard and resistant strain of *E. coli*. The results showed that the activity of all tested antibiotics especially doxycycline on the resistant strain was enhanced when it was used in combination with plant material. The enhanced activity of cephalexin against the standard strain has been reported to be higher than resistant strain [38]. The minimum inhibitory concentration (MIC) of the methanol extract of *A. millefolium* is reported as 50 µg/ml [39]. In another study, the ethyl acetate extracts of *A. talagonica* and *A. tenuifolia* showed a moderate activity against the epimastigotes of *T. cruzi* [40]. *A. fragrantissima* showed the highest antiviral activity (among these species) against POLIO in a concentration dependent manner at complete non-toxic concentration range (10–100 µg/ml) and the highest detected antiviral activity was recorded at Rf of 10⁻⁶. It seems that the interesting antiviral activity of *A. fragrantissima* against POLIO may be attributed to essential oil content which has been traditionally used as an antiseptic agent [41]. A new ionone glucoside, biebersteininide, together with four known compounds 6-epiroseoside, ascaridole, strictic acid and centipedic acid were reported from the aerial parts of *A. biebersteinii*. The compounds were reported for the first time from *A. biebersteinii*. Also, antifungal activity was observed from the compounds [42].

2.13 Antispasmodic activity

The antispasmodic effects of *Achillea* species might be due to the flavonoid constituents of the plant. Galangin, quercetin and eupatilin, which are found commonly in *Achillea*, are reported to cause a potent relaxation of the ileum guinea-pig ileum [43]. The effect of *A. millefolium* hydro-alcoholic extract on the contractile responses of the isolated guinea-pig ileum at five concentrations ranging from 0.05 to 5 mg/ml has been reported. Changes in contraction of tissues were monitored using force displacement transducer amplifier connected to physiograph. Each segment served as its own control. Results showed that the contractile response was inhibited by extract in a dose-dependent manner (EC₅₀ = 1.5 mg/ml). Those results demonstrated that in vitro evaluation of *A. millefolium* extract resulted in inhibition of electrical induced contractions of the guinea-pig ileum [44].

2.14 Anti-inflammatory activity

Baccharis coridifolia D.C. (Mio-Mio, Romerillo) is used externally as an anti-inflammatory [2]. The aqueous extract and dichloromethane extract of *Pluchea sagittalis* produced a total inhibition of 50.85% and 41.16% respectively. This study shows that the aqueous and dichloromethane extract of *Pluchea sagittalis* had an anti-inflammatory effect in the carrageenan-foot oedema test [45]. The methanol extracts of *Emilia sonchifolia* leaves found to inhibit carrageenan induced oedema indicating its Anti-inflammatory activity. [26] The influence of the methanol fraction of *Pluchea indica* Less root extract (PIRE) was evaluated in vivo for anti-inflammatory activity. PIRE produced significant anti-inflammatory activity against glucose oxidase induced paw oedema [46]. The anti-inflammatory and antinociceptive activities of the ethanolic extract (EE) from aerial parts of *Pluchea quitoc* DC. (Asteraceae) were evaluated in mice and rats. Oral treatment with the EE (1–2 g/kg, p.o.) decreased the paw oedema induced by carrageenan in rats, showed antinociceptive effects on the tail-flick test and on acid-induced writhing in mice, and inhibited both phases of pain (neurogenic and inflammatory) of the formalin test in rats. Topical application (EE 1.25, 2.5 and 5.0 mg) inhibited the ear oedema induced by croton

oil in mice. The results support the folkloric use of the plant in inflammatory processes [47]. The ethanolic extract of *Pluchea lanceolata* exhibited significant anti-inflammatory activity, which was further investigated after fractionation. The result showed that activity was localized in the hexane fraction, from which 1-taraxasterol acetate was isolated which proved to be one of the active constituents. Taraxasterol acetate, isolated from hexane fraction, accounted for only part of the activity of that fraction. It is obvious that there are other active substances present in the hexane fraction which need to be isolated and characterized [48]. Neolupenol, a pentacyclic triterpene isolated from *Pluchea lanceolata* flowers, was studied to determine its anti-inflammatory activity against carrageenin-induced rat-paw edema. The degree of edema inhibition was found to increase with dose as well as time interval and was found to be maximum at 300 min. Neolupenol, when administered at 100 mg/kg, p.o. was found to exhibit 70% edema inhibition which was greater than that of the reference compound, ibuprofen (50mg/kg, p.o., 65% inhibition, and 300 min) [49].

Besides the alkaloids, as the noteworthy active anti-inflammatory compounds, sesquiterpenes are introduced as another effective group of the secondary metabolites. The methylene chloride - methanol extract of aerial parts of *A. coarctata* was investigated by chromatographic analysis and resulted in isolation of two new guaiane acid derivatives, 1 α ,6 α ,8 α -trihydroxy-5 α ,7 β H-guaia-3,10(14),11(13)-trien-12-oic acid and 1 α ,6 α ,8 α -trihydroxy-5 α ,7 β H-guaia-3,9,11(13)-trien-12-oic acids, in addition to three known compounds, ligustolide-A, arteludovicinolide-A and austriacin XL [50]. They also reported that the compounds enhanced the proliferation of beneficial macrophages significantly and compounds exhibited anti-inflammatory properties (Hegazi et al., 2008). Another article has reported that chromatographic separation on dichloromethane extract of *A. clypeolata* resulted in one guaiane 4,10,11-trihydroxyguaiane, four eudesmanes 4(15)-eudesmene-1 β ,11-diol, clypeotriol, 3-epi-clypeotriol, cryptomeridiol, one diterpene sugereoside and two phenolics centaureidin and scopoletin XLVII. The compounds XLI and XLVI have been reported for the first time [51].

2.15 Venom Neutralizing Capacity

The methanol root extracts of *Pluchea indica* (Less) were explored for the first time for neutralization of snake venom (*Viperurus sellii*) activity. The *P. indica* root extracts significantly neutralized the viper venom-induced lethality and haemorrhagic activity in albino rat and mouse. Venom-induced coagulant and anticoagulant activity was also antagonized by both the extracts. No precipitating bands were observed between the plant extract and polyvalent snake venom antiserum. These observations confirmed that certain Indian medicinal plants like *Pluchea indica* possess significant snake venom neutralizing capacity and need further examination for their active constituents [23].

The neutralization of viper and cobra venom by β -sitosterol and stigmasterol isolated from the root extract of *P. indica* Less. (Asteraceae) was evaluated in experimental animals. Cobra venom-induced lethality, cardiotoxicity, neurotoxicity, respiratory changes and PLA2 activity were also antagonized by the active component. It potentiated commercial snake venom antiserum action against venom-induced lethality in male albino mice. The active fraction could

antagonize venom-induced changes in lipid peroxidation and superoxide dismutase activity. This study suggests that β -sitosterol and stigmasterol may play an important role, along with antiserum, in neutralizing snake venom induced actions [52].

III. Conclusion

Some of these traditional and folk usages have been evaluated showing the potential medicinal use of the plant. The medicinal properties of Asteraceae are worldwide recognized and the plant is included in the national Pharmacopoeias of countries such as Germany, Czech Republic, France and Switzerland. As it is reviewed in this paper, antioxidant and protective activity of the Asteraceae is reported frequently. This might be due to high content of flavonoids and phenolics in these plants. It is noteworthy that oxidative stress is produced under diabetic condition and plant like *Achillea millefolium* (Asteraceae) is considered for high hypoglycemic activity. Among the medicinal properties of *Achillea*, their cytotoxic and antiulcer effects are important especially when the species contain immunomodulatory constituents. The activity of these plants against different bacteria, fungi and parasites might be due to the presence of a broad range of secondary active metabolites such as flavonoids, phenolic acids, coumarins, terpenoids (monoterpenes, sesquiterpenes, diterpenes, triterpenes) and sterols which have been isolated. Finally, presence of anti-inflammatory compounds such as sesquiterpenes and alkaloids is another reason for importance of these plants as the potential source of medicinal compounds and drugs in future.

References

- [1]. Tapia, A., Rodríguez, J., Theoduloz, C., Lopez, S., Feresin, G. E. and Schmeda-Hirschmann, G. Preliminary phytochemical activity of the *Pluchea* species. *Journal of Ethnopharmacology*(2004);**95**, 155.
- [2]. Maria J. A., and Paulina B. Baccharis (Compositae): a review update. *marigold petals. Journal of Food Science*(2007); **41**,163-164.
- [3]. Guo, Y., Li, Y., Xu, J., Watanabe, R., Oshima, Y., Yamakuni, T. and Ohizumi, Y.. Preliminary Phytochemical and Antimicrobial screening of *Bucharrisgaudichaudiana* *Journal of Natural Product*(2006);**69**: 274.
- [4]. Falk, A. J., Smolenski, S. J., Bauer, L. and Bell, C. L.. Isolation and identification of three new flavones from *Achillea millefolium* L. *Journal of Pharmaceutical Sciences*(2000);**64**: 1838-1842.
- [5]. Kizil, S., Çakmak, S., Kirici, M. and Inan, A. Comprehensive study on safflower (*carthamustinctorius* L.) In semi-arid conditions. *Journal of biotechnology*(2008);**12**: 23
- [6]. Philip, T., and Berry, W. Process for the purification of lutein-fatty acid esters from *tegetes erecta*. *Phytotherapy Research*(2000);**9**:145.
- [7]. Pirbalouti, A.G., Koohpayeh, A., Karimi, I. The wound healing activity of flower extracts of *Punicagranatum* and *Achillea kellerensis* in Wistar rats. *Acta Pol Pharm.*(2000);**67**: 107-110.
- [8]. Ayyanar, M. and Ignacimuthu, S. Herbal medicines for wound healing among tribal people in Southern India: Ethnobotanical and Scientific evidences. *International Journal of Applied Research in Natural Products*(2009);**2**(3): 29-42.
- [9]. Mantle, D., Eddeb, F. and Pickering, A.T. Comparison of relative antioxidant activities of British medicinal plant species in vitro. *Journal of Ethnopharmacology*(2000);**72**: 47-51
- [10]. Konyalioglu, S. and Karamenderes, C. The protective effects of *Achillea* L. species native in Turkey against H₂O₂-induced oxidative damage in human erythrocytes and leucocytes. *J Ethnopharmacol*(2005);**102**: 221-227
- [11]. Kundakovic, T., Mimica, T., Dukic, N. and Kovacevic, N. Free radical scavenging activity of *Achillea alexandri-regis* extracts. *Fitoterapia*(2005);**76**: 574-576.
- [12]. Ardestani, A. and Yazdanparast, R. Antioxidant and free radical scavenging potential of *Achillea santolina* extracts. *Food Chemistry*(2007);**104**: 21-29.
- [13]. Giorgi, A., Bombelli, R., Luini, A., Speranza, G., Cosentino, M., Lecchini, S. and Cocucci, M. Antioxidant and cytoprotective properties of infusions from leaves and inflorescences of *Achillea collina* L. *Ethnopharmacology*(2009);**104**: 21-29
- [14]. Schulz, V., Hansel, R. and Tyler, V.E. In: *Rational Phytotherapy: A Physician's guide to Herbal Medicine*: Springer(2001); **12** P 294.
- [15]. Innocentia, G., Vegetob, E., Dall-Acqua, S., Cianab, P., Giorgetta, M., Agradi, E., Sozzib, A., Ficoc, G. and Tomec, F. In vitro estrogenic activity of *Achillea millefolium* L. *Phytomedicine*(2007);**14**: 147-152.
- [16]. Yazdanparast, R., Ardestani, A. and Jamshidi, S. Experimental diabetes treated with *Achillea santolina*: Effect on pancreatic oxidative parameters. *Journal of Ethnopharmacology*(2007);**112**: 13-18.
- [17]. Montanari, T., De Carvalho, J.E. and Dolder, H. Antispermatic effect of *Achillea millefolium* L. in Mice. *Contraception*(2005);**58**: 309-313.
- [18]. Alisi, C. S. and Onyeze, G.O. Nitric oxide scavenging ability of ethyl acetate fraction of methanolic leaf extracts of *Chromolaena odorata* (Linn.). *African Journal of Biochemistry Research*(2008);**2** (7): 145-150.
- [19]. Jahangir, T.T.H., Khan, L., Prasad, S., and Sultana B.. Anti oxidant activity of the *Pluchea* genus *Journal of Pharmacy and Pharmacology*(2005); 57 119
- [20]. Andarwulan, N.R., Batari, D.A., Sandrasari, B., Bolling, H. and Wijaya, A. Total phenolics and Antioxidant activity of *Pluchea indica*. *Food Chemistry*(2010); **12**: 34
- [21]. Marwah, R.G., Fatope, M.O., Mahrooqi, R.A., Varma, G.B. and Abadi, H. Total phenolics and Antioxidant activity of *Pluchea Arabica*. *Food Chemistry*(2007);**101**: 465.
- [22]. Zani, C.L., Alves, T.M., Oliveira, A.B.D., Murta, S.M.F., Ceravolo, R. I and Romanha, I J. Phytoconstituent and antimicrobial activity of *Pluchea aquitoc* *Phytotherapy Research* (1994); **13**, 375.
- [23]. Alam, M.I., Auddy, B. and Gomes, A. Preliminary phytochemical and antimicrobial activity of *Pluchea indica*. *Phytotherapy Research*(2000);**10**: 58
- [24]. Cavalcanti, A.M., Baggio, C.H., Freitas, C.S. Rieck, L., De Sousa, R.S., Da Silva-Santos, J.E., Mesia-Vela, S., and Marques, M.C.A. Safety and antiulcer efficacy studies (Darul(2006); **17**: 37-41.
- [25]. Niazmand, S., Khooshnood, E. and Derakhshan, M. Effects of *Achillea wilhelmsii* on rat's gastric acid output at basal, vagotomized, and vagal-stimulated conditions. *Pharmacognosy Mag*(2010);**6**: 282-285.
- [26]. Muko, K.N. and Ohiri, F.C. A Preliminary study on the anti-inflammatory properties of *Emilia sonchifolia* leaf extract. *Fitoterapia*(2000);**71** (1): 65-8
- [27]. Chengaiah, B.K., Mallikarjuna, R.A., Mahesh, M., Alagusundaram, C. and Madhusudhana, M. Medicinal importance of natural dyes. *International Journal of Pharmaceutical Technology Research* (2010); **2**(1):144-154.
- [28]. Ghantous, A., Nasser, N., Saab, I., Darwiche, N. and Saliba, N.A. Structure-activity relationship of seco-tanaphthalides isolated from *Achillea falcata* for inhibition of HaCaT cell growth. *European Journal of Medicinal Chemistry*(2009); **44**: 3794-3797
- [29]. Trifunovic, S., Vajs, V., Juranic, Z., Zizak, Z., Tesevic, V., Macura, S. and Milosavljevic, S. Cytotoxic constituents of *Achillea clavennae* from Montenegro. *Phytochemistry* (2006);**68**: 887-893.
- [30]. Suksamran, A., Chotipong, A., Suavansri, T., Boongird, S., Timsuksai, P., Vimuttipong, S. and Chuaynugul, A. Antimycobacterial activity and cytotoxicity of flavonoids from the flowers of *Chromolaena odorata*. *Arch Pharm Res.*(2004);**5**(27): 507-511.
- [31]. Mandeel, Q. A. and Taha, A. Preliminary phytochemical screening of *Pluchea lanceolata*. *Journal of Pharmaceutical Biology*(2005);**43**(4), 340.
- [32]. Rezaeiipoor, R., Saeidnia, S. and Kamalinejad, M. Immunosuppressive activity of *Achillea atalagonica* on humoral immune responses in experimental animals. *Journal of Ethnopharmacology*. (2000); **65**: 273-276.
- [33]. Saeidnia, S., Yassa, N., Rezaeiipoor, R., Shafiee, A., Gohari, A.R., Kamalinejad, M., and Goodarzy, S. Immunosuppressive principles from *Achillea atalagonica*, An endemic species of Iran. *Darul*(2009); **17**: 37-41.
- [34]. Yassa, N., Saeidnia, S., Pirouzi, R., Akbaripour, M. and Shafiee, A. Three phenolic glycosides and immunological properties of *Achillea millefolium*. *Darul*(2007);**15**: 49-52.
- [35]. Saeidnia, S., Yassa, N. and Rezaeiipoor, R. Comparative investigation of the essential oils of *Atalagona Boiss* and *A. millefolium* L., Chemical composition and immunological studies. *Journal of Essential Oil Research* 2004; **16**: 262-264.
- [36]. Maggi, F., Bramucci, M., Cecchini, C., Coman, M.M., Cresci, A., Cristalli, G., Lupidi, G., Papa, F., Quassinti, L., Sagratini, G. and Vittori, S. Composition and biological activity of essential oil of *Achillea ligustica* All. (Asteraceae) naturalized in central Italy: Ideal candidate for anti-carcinogenic formulations. *Fitoterapia*(2009);**80**: 313-319.

- [37]. Stojanovic, G., Radulovic, N., Hashimoto, T. and Palic, R. In vitro antimicrobial activity of extracts of four Achillea species. The composition of Achillea clavennae L. (Asteraceae) extract. Journal of Ethnopharmacology (2005); **101**:185-190.
- [38]. Darwish, R.M. and Aburjai, T.A. Effect of ethnomedicinal plants used in folkloric medicine in Jordan as antibiotic resistant inhibitors on Escherichia coli. BMC Complement Alternative Medicine (2010); **10**: 9.
- [39]. Mahady, G.B., Pendland, S.L., Stoia, A., Hamill, F.A., Fabricant, D., Dietz, B.M. and Chadwick, L.R. In vitro susceptibility of Helicobacter pylori to botanical extracts used traditionally for the treatment of gastrointestinal disorders. Phytotherapy Research (2005); **19**: 988-991.
- [40]. Gohari, A.R., Saeidnia, S., Hadjiakhoondi, A., Naghinejad, A. and Yagura, T. Trypanocidal activity of some medicinal plants against the epimastigotes of Trypanosoma cruzi. Journal of Medicinal Plants (2008); **7**: 44-48.
- [41]. Soltan, M.M. and Zaki, A.K. Antiviral screening of forty-two Egyptian medicinal plants. Journal of Ethnopharmacology (2009); **126**: 102-107.
- [42]. Mahmoud, A.A. and Al-Shihry, S.S. A new ionone glucoside and terpenoid constituents from Achilleaiebersteini and their antifungal activity. Natural Product Community (2006); **1**: 697-703.
- [43]. Hammad, H.M. and Abdalla, S.S. Pharmacological effects of selected flavonoids on rat isolated ileum: Structure-activity relationship. General Pharmacology (2007); **28**: 767-771.
- [44]. Babaei, M., Abarghoei, M.E., Akhavan, M.M., Ansari, R., Vafaei, A.A., Taherian, A.A., Mousavi, S. and Toussy, J. Antimotility effect of hydroalcoholic extract of yarrow (Achillea millefolium) on the guinea-pig ileum. Pakistan Journal of Biological Sciences (2007); **10**: 3673-3677.
- [45]. Perez, F.E., Marin, T. and Adzet, T. Preliminary phytochemical screening and antimicrobial activity of Pluchea sagittalis. Phytotherapy Research (2000); **9**: 145.
- [46]. Sen, T., Dhara, A.K., Bhattacharjee, S.S., Pal, S., Nag, A.K. and Chaudhuri, A. Preliminary phytochemical and antimicrobial activity of Pluchea indica. Phytotherapy Research (2002); **16**: 331.
- [47]. Barros, I.M., Lopes, I., Borges, O., Borges, C., Ribeiro, N. and Freire, M. Preliminary phytochemical screening and antimicrobial activity of Pluchea quito. Journal of Ethnopharmacology (2006); **14**: 34.
- [48]. Srivastava, V. N., Varma, J.S., Tandon, R.C. and Srimal, A. Anti-inflammatory activity of Pluchea lanceolata. International Journal of Crude Drug Research (2000); **28**(2), 135.
- [49]. Kaith, B.S. Preliminary phytochemical and antimicrobial activity of Pluchea lanceolata. International Journal of Pharmacognosy (2000); **34**(1), 73.
- [50]. Hegazy, M.E.F., Abdel-Lateff, A., Gamal-Eldeen, A.M., Turkey, F., Hirata, T., Pare, P.W. and Karchesy, J.K. Anti-inflammatory activity of new guaiane acid derivatives from Achillea coarctata. Natural Product Community (2008); **3**: 851-856.
- [51]. Werner, I., Mucaji, P., Presser, A. and Glasl, S. Sesquiterpenes and phenolic compounds from Achillea clypeolata. Z Naturforsch B (2007); **62**: 267-271.
- [52]. Gomes, A., Saha, A., Chatterjee, I. and Chakravarty, A.K. Preliminary phytochemical and antimicrobial activity of Pluchea indica. Phytomedicine (2007); **14**: 637.

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