

Bisbenzylisoquinoline Alkaloids From Genus *Triclisia* As Potential Anticancer Agents: A Review

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Abstract

Despite advancements in anticancer strategies such as radiotherapy, surgery, chemotherapy, and immunotherapy, cancer continues to be a global health burden. There were an estimated 20 million new cancer cases and 9.7 million deaths in 2022 with lung cancer leading in incidence and mortality. It ranks as the second most prevalent cause of death globally. Some of the challenges encountered in cancer treatment and management include toxicity of conventional treatments and multidrug resistance among others. There is therefore need for continued search for new drugs to counter these challenges and lessen the cancer burden. Plants are a potential source of compounds that can be developed into new anticancer agents. Bisbenzylisoquinoline (BBIQ) alkaloids from plants belonging to the family Menispermaceae are a good example of compounds that have been shown to possess anticancer activity. The genus *Triclisia* belongs to this family. The plants in this genus have found use in traditional medicine where they are used in the management and treatment of many conditions and ailments including cancer. The main purpose of this review is to highlight the anticancer potential of BBIQ alkaloids from plants in this genus. Information was collected from scientific databases and relevant books. A literature survey revealed that there are five species in the genus *Triclisia* from which a total of 24 BBIQ alkaloids have been isolated. These are: *Triclisia patens* Oliv, *T. dictyophylla* Diels, *T. gillettii* DeWild, *T. subcordata* Oliv. and *T. sacleuxii* (Pierre) Diels. Phaeanthine, 1,2-dehydroapateline, tetrandrine, cycleanine, fangchinoline, aromoline, pycmanilline and cocsuline are some of the BBIQ alkaloids from these plants that are reported to possess anticancer properties. Some of the compounds like tetrandrine and fangchinoline have been extensively studied while others remain unexplored. The BBIQ alkaloids that have shown promising activity *in vitro* and *in vivo* should be subjected to preclinical trials and possibly clinical trials to assess their safety and effectiveness. Combination therapy and structural modifications are also areas that need more focus. The unexplored alkaloids should be studied to discover their anticancer potential. All these will help reduce the relatively high morbidity and mortality associated with cancer globally.

Keywords: Menispermaceae, *Triclisia*, bisbenzylisoquinoline alkaloids, cancer, cytotoxicity.

Date of Submission: 06-04-2026

Date of Acceptance: 16-04-2026

I. Introduction

Cancer is a generic term for a large group of diseases that can affect any part of the body. Other terms used are malignant tumours and neoplasms. One defining feature of cancer is the rapid creation of abnormal cells that grow beyond their usual boundaries, and which can then invade adjoining parts of the body and spread to other organs; the latter process is referred to as metastasis. Widespread metastases are the primary cause of death from cancer¹. Despite advancements in anticancer strategies such as radiotherapy, surgery, chemotherapy, and immunotherapy, cancer continues to be a global health burden. According to International Agency for Research on Cancer (IARC), an agency of the WHO, there were an estimated 20 million new cancer cases and 9.7 million deaths in 2022 with lung cancer leading in incidence and mortality². Cancer ranks as the second most prevalent cause of death globally³. The global cancer burden is not evenly distributed. There is a clear correlation between socioeconomic development and cancer rates with poor countries bearing the greatest burden. This is due to factors such as lack of proper preventive measures, late diagnosis and high treatment costs. Asia leads in incidence, mortality and prevalence. Over 35 million new cancer cases are predicted in 2050, a 77% increase from the estimated 20 million cases in 2022². Challenges that continue to be encountered in cancer management and treatment globally include; multi-drug resistance (MDR), immune-related side effects, high costs, immunosuppressive tumour microenvironments and toxicity of conventional treatments^{4,5,6}.

Till 2019, approximately 84% of the anticancer drugs in clinical use were either of purely natural origin or with natural pharmacophore, semi-synthetically modified, or the ones that mimic the natural products or the botanical formulations^{7,8}. Bisbenzylisoquinoline (BBIQ) alkaloids are among the compounds that have been shown to possess anticancer properties with some of them in clinical use as anticancer agents. They have shown great promise in combating some of the challenges in cancer treatment like MDR and toxicity of

conventional treatments. This review highlights the anticancer potential of BBIQ alkaloids from plants in the genus *Triclisia*. It identifies the gaps and points out areas of further research that will enable full exploitation of the anticancer potential of these alkaloids. These alkaloids are a defining feature of the family Menispermaceae, they are abundant and display a wide variety of pharmacological activities. *Triclisia* is a genus in this family from which BBIQ alkaloids have been isolated.

II. Materials And Methods

Information on plants in the genus *Triclisia*, their constituent BBIQ alkaloids and their cytotoxic activities was collected from the scientific search engines: Web of science, Google Scholar, SciFinder, and PubMed. Relevant books were also used. The chemical structures of BBIQ alkaloids isolated from plants in the genus *Triclisia* were drawn using ChemDraw Professional 15.0 software.

III. Results And Discussion

The Genus *Triclisia*

There are about 20 species of the genus *Triclisia* known throughout globe with 12 in tropical Africa Region⁹. This review identified 5 *Triclisia* species from which compounds have been isolated. These are *Triclisia dictyophylla* Diels, *Triclisia patens* Oliv., *Triclisia gillettii* (DeWild), *Triclisia subcordata* Oliv. and *Triclisia sacleuxii* (Pierre) Diels. Various parts of all these plants have found use as sources of medicine for the treatment of various ailments in traditional medicine.

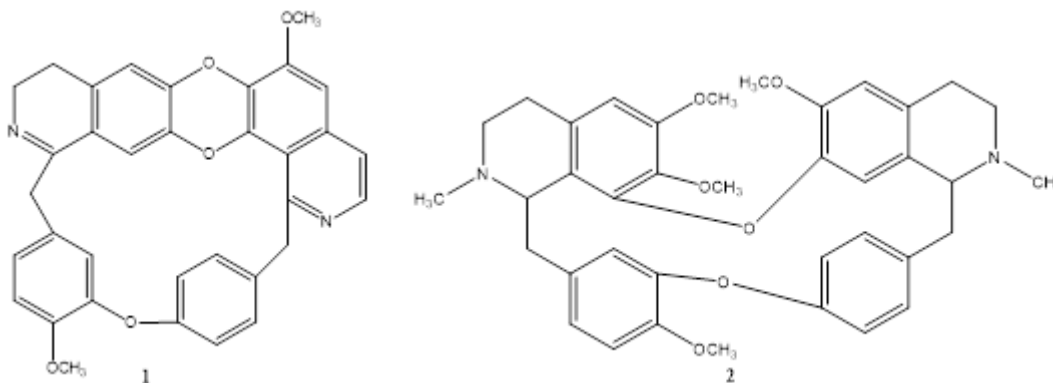
Bisbenzylisoquinoline (BBIQ) Alkaloids

BBIQ alkaloids are built up of two benzylisoquinoline (BI) units linked by ether bridges. In addition to this ether linkage, methylenoxy bridging or direct carbon-carbon bonding is also found between the two BI units. A variety of structural patterns arise in the BBIQ molecules due to differences in the number of aromatic oxygen substituents present, the number of ether linkages, the nature of ether bridges, viz., diphenyl ether or benzylphenyl ether and the sites on the two BI units at which the ether or the carbon-carbon bond originates¹⁰. Based on these differences, the BBIQ alkaloids are classified into groups and subgroups^{10, 11}.

There are about 500 natural products belonging to this large alkaloid class¹¹. The number is likely higher than this considering that this figure was given about six years ago. These alkaloids mostly occur in the Menispermaceae, Berberidaceae, Lauraceae, and Ranunculaceae plant families, which preferably grow in tropical and subtropical regions¹²⁻¹⁶. A good number of plants from these families have been used in traditional medicine to treat and manage various ailments and conditions¹⁷⁻²⁰. Formulations containing parts of these plants have also been used for centuries as an arrow poison in South America²¹. Most of these traditional uses can be attributed partly to the presence of BBIQ alkaloids in various parts of these plants. This is supported by reports on the pharmacological activities of these alkaloids isolated from numerous plants in these families. There are five species in the genus *Triclisia* from which a total of 24 BBIQ alkaloids have been isolated. These are: *Triclisia sacleuxii* (Pierre) Diels, *T. subcordata* Oliv., *T. gillettii* DeWild, *T. patens* Oliv. and *T. dictyophylla* Diels.

T. sacleuxii (Pierre) Diels

T. sacleuxii is a climbing plant producing stems 10 - 15 metres long²¹. It is found in Angola, Central African Republic, Congo, Gabon, Kenya, Mozambique and Tanzania²². BBIQ alkaloids have been isolated from the roots and leaves of this plant. From the roots gasabiimine (1), phaeanthine (2), 1,2-dehydroapateline (3), N-methylapateline (4), 1,2-dehydrotelobine (5) and *O*-methylcoccoline (6) have been isolated²³⁻²⁵. Lindoldhamine (7)²⁵ and isotetrandrine (8) have been isolated from the leaves²⁶.



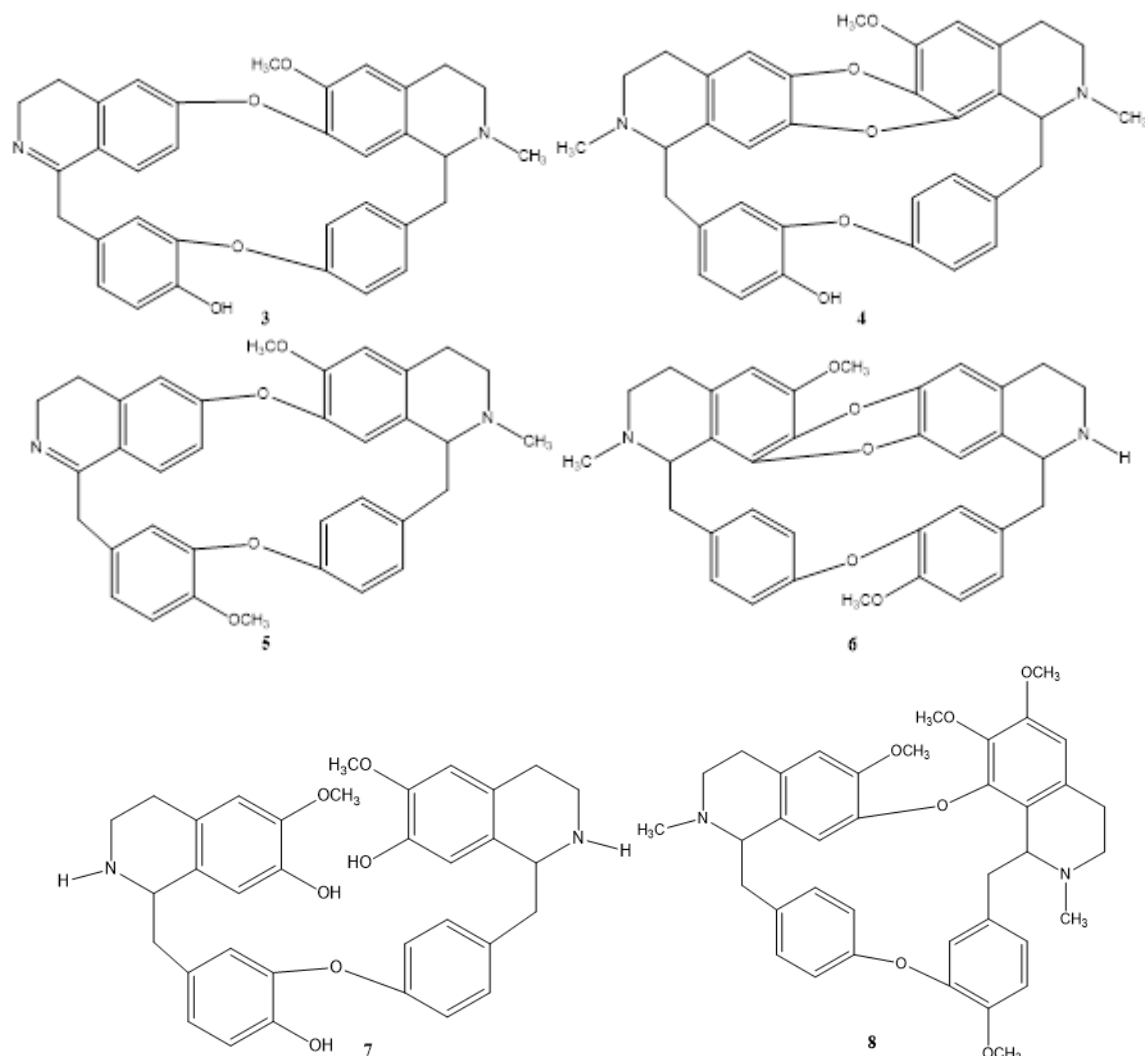
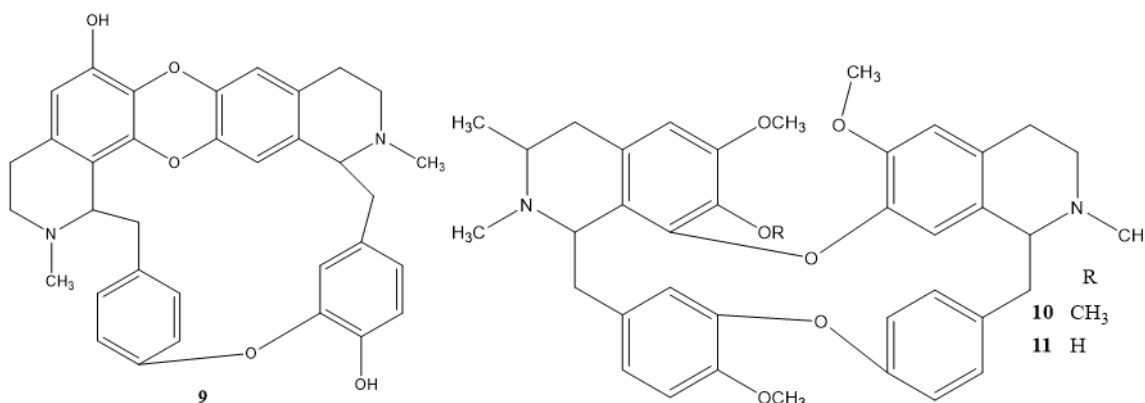


Fig. 1: Chemical structures of BBIQ alkaloids from *T. sacleuxii* (Pierre) Diels: Gasabiimine (1), phacanthine (2), 1,2-dehydroapateline (3), N-methylapateline (4), 1,2-dehydrotelobine (5) and *O*- methylcoccoline (6), lindoldhamine (7) and isotetrandrine (8)

***T. subcordata* Oliv.**

This is a slender woody twiner with brownish stems 6–15 ft. long. It has green flowers which later turn brown. Its fruits are orange²⁷. It is found in Benin, Burkina Faso, Ghana, Ivory Coast, Nigeria and Togo²⁸.

Early phytochemical studies in the 1970s revealed the presence of only three BBIQ alkaloids: tricordatine (9)²⁹, fangchinoline (10)³⁰ and tetrandrine (11)³¹ from this plant species. Additional BBIQ alkaloids from the roots have since been reported. These include cycleanine (12)³², isochondodendrine (13) and 2'-norcoccoline (14)³³.



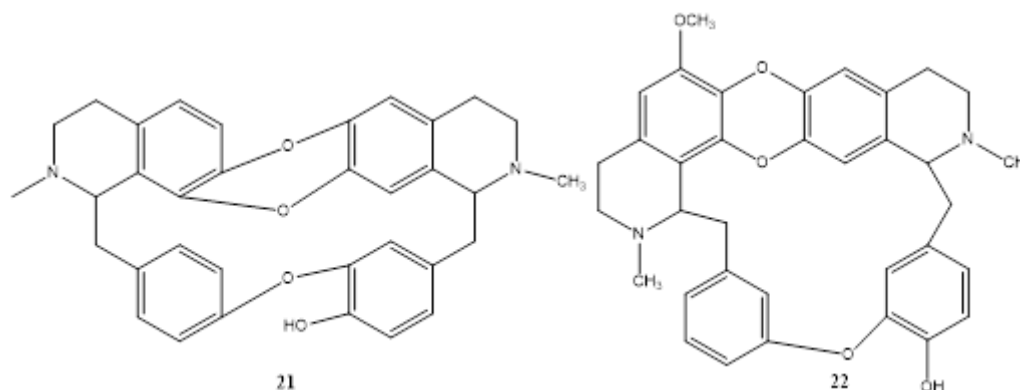


Fig. 3: Chemical structures of stebisimine (15), gilletteine (16), isogilletteine-N-oxide (17), obamegine (18), trigilletimine (19), pycmanilline (20), cocsuline (21) and trigilletine (22)

T. patens Oliv.

T. patens Oliv., is a dioecious liana with a stem up to 6 cm in diameter and a length of up to 12 m. The leaves alternate, are whole, and have pinnate veins. The fruit is made up of spheroid, flattened drupes and the flowers are unisex³⁸. It is a native of Burkina Faso, Ghana, Guinea, Guinea-Bissau, Ivory Coast, Liberia, Senegal and Sierra Leone³⁹.

Phaeanthine (2), trigilletimine (19), pycnamine (20), cocsuline (21), aromoline (23) and *N,N'*-dimethylphaeanthine (24) have been isolated from its leaves^{30,38,40,41}. From the stem, phaeanthine (2) and trigilletimine (19) have been isolated^{31,42}. Trigilletimine (19) has also been isolated from the roots³¹.

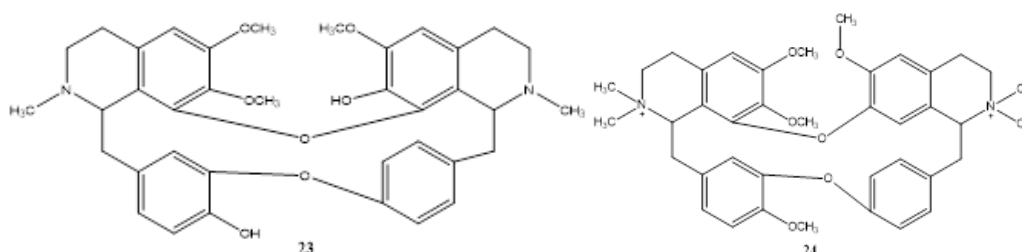


Fig. 4: Chemical structures of aromoline (23) and *N,N'*- dimethylphaeanthine (24)

T. dictyophylla Diels

T. dictyophylla Diels is a climbing plant or scrambling shrub of the lowland dense rain forest with stems that can be up to 30 m long. The plant occurs from Liberia to Central Africa, Democratic Republic of Congo, Angola and Tanzania³⁴. Phaeanthine (2), *N,N'* – dimethylphaeanthine (24), trigilletimine (19) and cocsuline (21) and have been isolated from the whole plant^{20,41,43}.

Cytotoxic BBIQs from *Triclisia* Species

Most of the BBIQ alkaloids isolated from plants in the genus *Triclisia* have been reported to possess cytotoxic activity hence they have anticancer potential. Some of the alkaloids like phaeanthine (2), isotetrandrine (8), fangchinoline (10), tetrandrine (11), cycleanine (12) and isochondodendrine (13) have been extensively studied. Others have been studied to a small extent while some remain unexplored.

Phaeanthine (2)

This BBIQ alkaloid has potential as an anticancer agent against different cancers according to various studies. For example, it has been observed to suppress the growth of cervical cancer (HeLa) cells with an IC₅₀ value of 8.11 μM⁴⁴. Its mechanism of action involves the activation of the intrinsic mitochondrial pathway; ultimately leading to apoptosis. It has also been observed that 2 induces several molecular changes. It reduces the levels of mitogen-activated protein kinase (Mcl-1) while simultaneously increasing B-cell lymphoma 2 (Bcl-2) associated X protein (BAX) and Bcl-2 homology domain (BH3) interacting domain death agonist (BID) expression. Furthermore, it elevates the cytoplasmic concentration of cytochrome c and leads to the activation of cleaved caspases 9 and 3⁴⁴. These are executioner caspases which play central roles in inducing apoptosis⁴⁵. 2 is also reported to possess cytotoxic activity against human lung fibroblasts, MCR-5 cells^{24,37} and KB (carcinoma of the human nasopharynx) cells^{46,47}.

Isotetrandrine (8)

The earliest report on the anticancer potential of this alkaloid was reported in 1976⁴⁸. Their study revealed that **8** has cytotoxic effects *in vitro* and antitumour effects *in vivo*. It was cytotoxic against cultured HeLa and HeLa-S₃ cells with ED₅₀ values of 5.8 and 10 µg/ml respectively. It had antitumour effects against both ascites type and solid type tumours in mice. This alkaloid also has cytotoxic effects against human oral epidermoid carcinoma (KB) cells with ED₅₀ value of 10600 nM⁴⁶. In addition, results from a study on P-glycoprotein-mediated multidrug resistance in human breast cancer doxorubicin-resistant (MCF-7/DOX) cells suggest that it might become a candidate of effective MDR reversing agent in cancer chemotherapy⁴⁹.

Fangchinoline (10)

Various reports reveal that this alkaloid inhibits the proliferation of different types of cancer cells. For example, it inhibits the proliferation of MDA-MB-231 and MCF-7 breast cancer cells, with IC₅₀ values of 25.2 and 34.6 µmol/L, respectively⁵⁰. In another study by Liu *et al.*⁵¹, it was cytotoxic against MDA-MB-231 cells with IC₅₀ value of 12.3 µmol/L, comparable to that of vincristine (13.1 µmol/L). The antiproliferation effect of **10** towards MDA-MB-231 cells is also reported by Wang *et al.*⁵², where its IC₅₀ values at 24 and 48 h were 14.3 and 10.5 µg/mL, respectively. Flow cytometry showed that MDA-MB-231 cell death was 16.4% and 28.0% after 24 and 48 h treatment with 20 µg/mL of **10**, respectively.

A study by Bao *et al.*⁵³ revealed that **10** significantly inhibits the growth of conjunctival melanoma (CM) cells *in vitro* and *in vivo*. This is a rare and fatal ocular tumour with poor prognosis. In this study, it exhibited *in vitro* anti-proliferative activity against four CM cell lines: CM-A516, CRMM1, CRMM2 and CM2005.1 with IC₅₀ values of 5.67, 2.68, 3.60 and 7.40 µM respectively. It also exhibited *in vivo* antitumour effects where the tumour volume of the treated group (at a dose of 50mg/Kg/day), was less by 27% compared to the control group. In addition, it increased the *in vivo* antitumour efficacy of cisplatin and reduced its dosage and toxicity. Mechanistic experiments in this study indicated that **10** suppressed the homologous recombination (HR)-directed DNA repair by binding with far upstream element binding protein 2 (FUBP2) and downregulating the expression of HR factors BRCA1 and RAD51.

In a study by Guo *et al.*⁵⁴, **10** effectively suppressed proliferation and invasion of A549 lung cancer cell line by inhibiting the phosphorylation of focal adhesion kinase (FAK) at Tyr397 position and its downstream pathways. Furthermore, it has antiproliferation activity against multiple myeloma⁵⁵, bladder cancer cells, T24 and 5637⁵⁶, hepatocellular carcinoma cells (HepG2 and PLC/ PRF/5 cells)⁵⁷, leukemia^{58,59}, gastric cancer⁶⁰, glioblastoma⁶¹, pancreatic cancer⁶² and prostate cancer cells⁶³. The molecular mechanisms of **10** against these cancers have been widely studied. It has been observed that it inhibits cancer proliferation by interfering with migration and angiogenesis, primarily through the downregulation of mesenchymal marker proteins such as fibronectin, vimentin, Snail, and Twist⁶⁴.

Tetrandrine (11)

This is one of the most studied BBIQ alkaloid from *Triclisia* species. There are numerous reports on its anticancer activity both *in vitro* and *in vivo* against a wide range of cancers of the breast, liver, pancreas, blood (leukemia), colon, lung, bladder, cervix, prostate, ovaries, etc^{33,65-70}. It has also been used in clinical trials in combination with the conventional drugs gemcitabine and cisplatin to treat lung cancer without any side effects^{71,72}. Many experiments have demonstrated that **11** has a number of anti-tumor mechanisms. It can inhibit the growth of tumor cells by inducing apoptosis, sensitizing radiotherapy, and protecting against potentially lethal injury caused by radiation. It can also alleviate radiation damage by inhibiting inflammatory reactions⁷¹. It has been used clinically in China for the treatment of lung cancer and leukemia⁷³.

Cycleanine (12)

In a study by Uche *et al.*³³, this alkaloid has anticancer activity on four ovarian cancer cell lines (Ovcar-8, A2780, Igrov-1, and Ovcar-4) with IC₅₀ values of 1.0, 7.6, 14 and 7.2 µM respectively. It causes apoptosis as shown by activation of caspases 3/7 and cleavage of poly (ADP-ribose) polymerase to form poly (ADP-ribose) polymerase-1 by using western blot analysis. Uche *et al.*⁷⁴ also reports the cytotoxic activity of **12** against the A2780 ovarian cancer cell line with IC₅₀ of 7.56 µM in and 44% viability at dose of 10µM. In addition, **12** has cytotoxic effect against human oral epidermoid carcinoma (KB) cells⁴⁶, human colon carcinoma (HCT-116) cells⁷⁵, HeLa cells^{44,48}, KB cells, SiHa cells and A549 cells⁷⁶ and Vero cells⁷⁷.

Isochondodendrine (13)

This BBIQ alkaloid is reported to possess weak to very good cytotoxic activity by a number of studies. In a study by Marshall *et al.*⁴⁷, it exhibited a very weak cytotoxic activity against KB (carcinoma of the human nasopharynx) cells with IC₅₀ value >421 µM. It has moderate cytotoxic activity against human colon carcinoma (HCT-116) cells with CC₅₀ values of 29 µM⁷⁵. Despite this very weak and moderate cytotoxicity, this alkaloid

has shown similar potency to the clinically used drug carboplatin against ovarian cancer *in vitro*³². When assayed against four ovarian cancer cell lines (Ovcar-8, A2780, Igrov-1, and Ovcar-4) it displayed very high potency with IC₅₀ values of 8.0, 3.5, 9.0 and 17 µM respectively. It acted by inducing apoptosis in ovarian cancer cells by activating caspases 3/7 amongst other mechanisms.

Other bisbenzylisoquinoline alkaloids from *Triclisia* species with cytotoxic activity

There are some BBIQ alkaloids from *Triclisia* species that are reported to have cytotoxic activity against one or two cancer cell lines only. This is a category that requires more focus. These include **3**, **4** and **5**, which possess cytotoxic activity against human fetal lung fibroblasts, MCR-5 cells with IC₅₀ values of 3.61, 1.25 and 3.59 µM respectively²⁴.

Other BBIQ alkaloids that require more studies include **9**, **14**, **18**, **21** and **23**. They have cytotoxic effects against human oral epidermoid carcinoma (KB) cells⁴⁶. In addition to cytotoxicity against human oral epidermoid carcinoma (KB) cells, **14**, **18**, **21** and **23** have been shown to exhibit activity against other cancer cell lines. For example, according to a study by Uche *et al.*³², **14** possesses anticancer activity on four ovarian cancer cell lines (Ovcar-8, A2780, Igrov-1, and Ovcar-4) with IC₅₀ values 1.9, 2.9, 0.8 and 6.2 µM respectively. It was found to be more potent than carboplatin in all four cancer cell lines and was observed to act by inducing apoptosis in the ovarian cancer cells. **16**, **18**, **19**, **20**, **21** and **23** possess cytotoxic activity against KB (carcinoma of the human nasopharynx) cells⁴⁷. The cytotoxicity of **20** is further confirmed in a study by Kanyanga *et al.*³⁷ where it is reported to be cytotoxic against MRC-5 cells (human lung fibroblasts).

IV. Conclusion

Plants in the genus *Triclisia* are a rich source of BBIQ alkaloids with anticancer properties. Some of these alkaloids like **2**, **8**, **10**, **11**, **12** and **13** have been extensively studied and their mechanisms of action established while others remain unexplored. Studies have shown that some of these alkaloids like **10** and **13** have efficacies that are comparable to conventional drugs against some cancers while others like **8** are potential candidates as multidrug resistance reversing agents in cancer chemotherapy by employing combination therapy. In addition, some of them reduce toxicity and increase the efficacy of anticancer drugs in clinical use. However, there are challenges that have been observed with these alkaloids. For example, **11**, which is in clinical use, has low bioavailability.

It is also worth noting that **14** which has not been so widely studied has been shown to be more potent than a clinically used drug against ovarian cancer. This means that it may be useful as a starting point in the development of new drugs against ovarian cancer. Future research should focus on structural modifications to improve the potency of these alkaloids. There is also need for more comprehensive studies on less explored and unexplored alkaloids to uncover their full therapeutic potential. Studies to solve the challenge of bioavailability should be carried out. In addition, extensive preclinical and clinical trials of BBIQ alkaloids like **10** are necessary.

References

- [1]. https://www.who.int/health-topics/cancer#tab=tab_1. Accessed 27/10/2025).
- [2]. Ferlay, J., Ervik, M., Lam, F., Laversanne, M., Colombet, M., Mery, L., Piñeros, M., Znaor, A., Soerjomataram, I., & Bray, F. Global Cancer Observatory: Cancer Today. Lyon, France: International Agency For Research On Cancer. Available From: <https://gco.iarc.who.int/today>. Accessed [27/10/2025].
- [3]. Sung H, Ferlay J, Siegel RL, Laversanne M, Soerjomataram I, Jemal A, Bray F. Global Cancer Statistics 2020: Globocan Estimates Of Incidence And Mortality Worldwide For 36 Cancers In 185 Countries. *Ca: A Cancer Journal For Clinicians*. 2021;71(3): 209-249. <https://doi.org/10.3322/caac.21660>.
- [4]. Azmal M, Miah Mm, Prima Fs, Paul Jk, Haque Asnb, Ghosh A. Advances And Challenges In Cancer Immunotherapy: Strategies For Personalized Treatment. *Seminars In Oncology*. 2025;52(3):152345. <https://doi.org/10.1016/j.seminoncol.2025.152345>.
- [5]. Nikolaou M, Pavlopoulou A, Georgakilas Ag, Kyrodimos E. The Challenge Of Drug Resistance In Cancer Treatment: A Current Overview. *Clinical & Experimental Metastasis*. 2018;35(4):309-318.
- [6]. Saha M, Sarkar A. Review On Multiple Facets Of Drug Resistance: A Rising Challenge In The 21st Century. *Journal Of Xenobiotics*. 2021;11(4):197-214. <https://doi.org/10.3390/jox11040013>.
- [7]. Islam Mr, Islam F, Nafady Mh, Akter M, Mitra S, Das R, Urmee H, Shohag S, Akter A, Chidambaram K, Alhumaydhi Fa. Natural Small Molecules In Breast Cancer Treatment: Understandings From A Therapeutic Viewpoint. *Molecules*. 2022;27(7):2165. <https://doi.org/10.3390/molecules27072165>
- [8]. Newman D J, Cragg Gm. Natural Products As Sources Of New Drugs Over The Nearly Four Decades From 01/1981 To 09/2019. *Journal Of Natural Products*. 2020;83: 770–803. <https://doi.org/10.1021/acs.jnatprod.9b01285>
- [9]. Troupin G. Menispermaceae. In W.B. Turrill, E. Milne-Redhead (Eds.), *Flora Of Tropical East Africa* (Pp. 32). Royal Botanic Gardens, 1956.
- [10]. Guha Kp, Mukherjee B, Mukherjee R. Bisbenzylisoquinoline Alkaloids-A Review. *J. Nat. Prod*. 1979;42: 1-84. <https://doi.org/10.1021/Np50001a001>
- [11]. Weber C, Opatz T. Bisbenzylisoquinoline Alkaloids. *The Alkaloids: Chemistry And Biology*. 2019;81: 1–114. <https://doi.org/10.1016/Bs.alkal.2018.07.001>
- [12]. Barbosa-Filho Jm, Da-Cunha Evl. Alkaloids Of The Menispermaceae. *The Alkaloids: Chemistry And Biology*. 2000;54:1–190. [https://doi.org/10.1016/S0099-9598\(00\)54002-4](https://doi.org/10.1016/S0099-9598(00)54002-4)

- [13]. Kessler Pja. In K.Kubitzki, J.G. Rohwer & V. Bittrich (Eds.), Flowering Plants—Dicotyledons: Magnoliid, Hamamelid And Caryophyllid Families (Pp. 402-418). Springer, 1993.
- [14]. Loconte, H. In K. Kubitzki, J.G. Rohwer, & V. Bittrich V. (Eds.), Flowering Plants—Dicotyledons: Magnoliid, Hamamelid And Caryophyllid Families (Pp. 147-152). Springer, 1993.
- [15]. Tamura, M. In K. Kubitzki, J.G. Rohwer, V. Bittrich (Eds.) Flowering Plants— Dicotyledons: Magnoliid, Hamamelid And Caryophyllid Families (Pp.563-583). Springer, 1993.
- [16]. Van Der Werff H, Richter Hg. Toward An Improved Classification Of Lauraceae. *Annals Of The Missouri Botanical Garden*. 1996;83: 409–418. <https://doi.org/10.2307/2399870>
- [17]. Chen Kk, Chen Al. Alkaloids Of Han-Fang-Chi. *Journal Of Biological Chemistry*. 1935;109: 681–685. [https://doi.org/10.1016/S0021-9258\(18\)75199-5](https://doi.org/10.1016/S0021-9258(18)75199-5)
- [18]. Chhabra Sc, Mahunnah Rla, Mshiu En. Plants Used In Traditional Medicine In Eastern Tanzania. Iii. Angiosperms (Euphorbiaceae To Menispermaceae). *Journal Of Ethnopharmacology*. 1990;28: 255–283. [https://doi.org/10.1016/0378-8741\(90\)90078-8](https://doi.org/10.1016/0378-8741(90)90078-8)
- [19]. De Wet H, Van Wyk B-E. An Ethnobotanical Survey Of Southern African Menispermaceae. *South African Journal Of Botany*. 2008;74:2–9. <https://doi.org/10.1016/J.Sajb.2007.07.001>
- [20]. Semwal Dk, Semwal Rb, Vermaak I, Viljoen Aj. The Ethnobotanical, Phytochemical And Pharmacological Significance Of Cissampelos (Menispermaceae). *Journal Of Ethnopharmacology*. 2014;155: 1011–1028. <https://doi.org/10.1016/J.Jep.2014.06.054>
- [21]. Bisset Ng. Arrow And Dart Poisons. *Journal Of Ethnopharmacology*. 1989;25: 1–41. [https://doi.org/10.1016/0378-8741\(89\)90043-3](https://doi.org/10.1016/0378-8741(89)90043-3)
- [22]. Tropical Plants Database, Ken Fern. *Tropical.Theferns.Info*. 2025-06-18. <https://Tropical.Theferns.Info/Viewtropical.Php?Id=Triclisia+Sacleuxii>
<https://powo.science.kew.org/taxon/urn:lsid:ipni.org:names:581679-1> (Accessed On 5/4/2025).
- [23]. Murebwayire S, Diallo B, Luhmer M, Vanhaelen-Fastre R, Vanhaelen M. Alkaloids And Amides From *Triclisia Sacleuxii*. *Fitoterapia*. 2006;77:615–617. <https://doi.org/10.1016/J.Fitote.2006.09.007>
- [24]. Murebwayire S, Frederich M, Hannaert V, Jonville M-C, Duez P. Antiplasmodial And Antitrypanosomal Activity Of *Triclisia Sacleuxii* (Pierre) Diels. *Phytomedicine*. 2008;15:728–733. <https://doi.org/10.1016/J.Phytmed.2007.10.005>
- [25]. Murebwayire S, Ingkaninan K, Changwijit K, Frederich M, Duez P. *Triclisia Sacleuxii* (Pierre) Diels (Menispermaceae), A Potential Source Of Acetylcholinesterase Inhibitors. *Journal Of Pharmacy Pharmacology*. 2009;61:103–107. <https://doi.org/10.1211/Jpp.61.01.0014>
- [26]. Samita F, Ochieng Co, Owuor Po, Manguro Loa, Midiwo Jo. Isolation Of A New B-Carboline Alkaloid From Aerial Parts Of *Triclisia Sacleuxii* And Its Antibacterial And Cytotoxicity Effects. *Natural Product Research*. 2017;31(5): 529-536. <https://doi.org/10.1080/14786419.2016.1201666>
- [27]. <https://plants.jstor.org/compilation/triclisia.subcordata> Accessed On 5/4/2025).
- [28]. <https://powo.science.kew.org/taxon/urn:lsid:ipni.org:names:581682-1> Accessed On 5/4/2025)
- [29]. Tackie An, Dwuma-Badu D, Okarter T, Knapp Je, Slatkin Dj, Schiff Jr Pl. Trigilletine And Tricordatine: Two New Bisbenzylisoquinoline Alkaloids From *Triclisia* Species. *Phytochemistry*. 1973;12:2509-2511. [https://doi.org/10.1016/0031-9422\(73\)80465-0](https://doi.org/10.1016/0031-9422(73)80465-0)
- [30]. Tackie An, Dwuma-Badu D, Okarter T, Knapp Je, Slatkin Dj, Schiff Jr Pl. Constituents Of Some West African Medicinal Plants. Ii. The Isolation Of Alkaloids From Selected *Triclisia* Species. *Lloydia*. 1974;37: 1–5.
- [31]. Dwuma-Badu D, Ayim Jsk, Tackie An, Knapp Je, Slatkin Dj, Schiff Jr Pl. Additional Alkaloids Of *Triclisia Patens* Sand *Triclisia Subcordata*. *Phytochemistry*. 1975;14:2524–2545. [https://doi.org/10.1016/0031-9422\(75\)80390-6](https://doi.org/10.1016/0031-9422(75)80390-6)
- [32]. Uche Fi, Abed Mn, Abdullah Mi, Drijfhout Fp, Mccullagh J, Claridge Tw, Richardson A, Li W-W. Isochondodendrine And 2'-Norcocusline: Additional Alkaloids From *Triclisia Subcordata* Induce Cytotoxicity And Apoptosis In Ovarian Cancer Cell Lines. *Royal Society Of Chemistry Advances*. 2017;7(70):44154-44161. Doi: 10.1039/C7ra08032h
- [33]. Uche Fi, Drijfhout Fp, Mccullagh J, Richardson A, Li W-W. Cytotoxicity Effects And Apoptosis Induction By Bisbenzylisoquinoline Alkaloids From *Triclisia Subcordata*. *Phytotherapy Research*. 2016;30(9):1533-1539. <https://doi.org/10.1002/Ptr.5660>
- [34]. Troupin, G. Monographie Des Menispermaceae Africaines. Mémoires In 8. Académie Royale Des Sciences D'outre-Mer, Classe Des Sciences Naturelles Et Médicales, Nouvelle Série, 1962.
- [35]. Owusu Pd, Slatkin Dj, Knapp Je, Schiff Jr Pl. (1981). Constituents Of West African Medicinal Plants. Xxviii. Additional Alkaloids Of *Triclisia Gilletii*. *Journal Of Natural Products*. 1981;44(1): 61-66. <https://doi.org/10.1021/Np50013a011>
- [36]. Dwuma-Badu D, Ayim Jsk, Tackie An, Owusu Pd, Knapp Je, Slatkin Dj, Schiff Jr Pl. Gilletine, A New Bisbenzylisoquinoline Alkaloid From *Triclisia Gilletii*. *Heterocycles*. 1978;9: 995-1002. <https://doi.org/10.1007/Bf01945760>
- [37]. Kanyanga Rc, Munduku Ck, Lumpu Ns, Ehata Mt, Bool-Miting Fm, Kabangu Ok, Maya Bm, Cos P, Maes L, Vlietinck A., Tuenter E, Foubert K, Pieters L. Isolation And Structure Elucidation Of Two Antiprotozoal Bisbenzylisoquinoline Alkaloids From *Triclisia Gilletii* Stem Bark. *Phytochemistry Letters*. 2018;28:19-23. <https://doi.org/10.1016/J.Phytol.2018.09.008>
- [38]. Mosango, D.M. *Triclisia Patens* Oliv. In: G.H. Schmelzer, A. Gurib-Fakim (Eds.), *Plant Resources Of Tropical Africa Ii: Medicinal Plants I* (618-619). Backhuys Publishers, 2008.
- [39]. <https://powo.science.kew.org/taxon/urn:lsid:ipni.org:names:581677-1> (Accessed On 05/04/2025).
- [40]. Camacho Mr, Phillipson Jd, Croft Sl, Rock P, Sarah J, Marshall Sj, Schiff Jr Pl. In Vitro Activity Of *Triclisia Patens* And Some Bisbenzylisoquinoline Alkaloids Against *Leishmania Donovanii* And *Trypanosoma Brucei Brucei*. *Phytotherapy Research*. 2002;16: 432–436. <https://doi.org/10.1002/Ptr.929>
- [41]. Kronlund A, Kristiansson K, Sandberg F. The Occurrence Of Phaeanthine And N,N' -Dimethylphaeanthine In *Triclisia Dictyophylla* And *T. Patens*. A New Simple Method For Estimation Of Muscle Relaxant Effect. *Acta Pharmaceutica Suecica*. 1970;7: 279-284.
- [42]. Ekong R, Partridge Sj, Anderson Mm, Kirby Gc, Warhurst Dc, Russell Pf, Phillipson Jd. Plasmodium Falciparum: Effects Of Phaeanthine, A Naturally-Occurring Bisbenzylisoquinoline Alkaloid, On Chloroquine-Resistant And-Sensitive Parasites In Vitro, And Its Influence On Chloroquine Activity. *Annals Of Tropical Medicine & Parasitology*. 1991;85(2): 205-213. <https://doi.org/10.1080/00034983.1991.11812547>
- [43]. Spiff Ai, Zabel V, Watson H, Zemaitis Ma, Ateya Am, Slatkin Dj, Knapp Je, Schiff Jr Pl. Constituents Of Some West African Medicinal Plants Xxx. Tridictyophylline, A New Morphinan Alkaloid From *Triclisia Dictyophylla*. *Journal Of Natural Products*. 1981;44(2): 160-165. <https://doi.org/10.1021/Np50014a003>

- [44]. Valsan A, Meenu Tm, Murali Pv, Malgija B, Joseph Ga, Nisha P, Radhakrishnan Vk, Maiti Kk. Exploration Of Phaeanthine: A Bisbenzylisoquinoline Alkaloid Induces Anticancer Effect In Cervical Cancer Cells Involving Mitochondria-Mediated Apoptosis. *Acs Omega*. 2023;8(16):4799-14813. Doi: 10.1021/Acsomega.3c01023
- [45]. Iksen, Witayateeraporn W, Hardianti B, Pongrakhananon V. Comprehensive Review Of Bcl-2 Family Proteins In Cancer Apoptosis: Therapeutic Strategies And Promising Updates Of Natural Bioactive Compounds And Small Molecules. *Phytotherapy Research*. 2024;38(5):2249-2275. <https://doi.org/10.1002/Ptr.8157>
- [46]. Angerhofer Ck, Guinaudeau H, Wangpanich V, Pezzuto Jm, Cordell Ga. Antiplasmodial And Cytotoxic Activity Of Natural Bisbenzylisoquinoline Alkaloids. *Journal Of Natural Products*. 1999;62:59-66. <https://doi.org/10.1021/Np980144f>
- [47]. Marshall Js, Russell Fp, Wright Wc, Anderson Mm, Phillipson Dj, Kirby Cg, David, C, Warhurst Cd, Schiff Jr Pl. In Vitro Antiplasmodial, Antiamoebic, And Cytotoxic Activities Of A Series Of Bisbenzylisoquinoline Alkaloids. *Antimicrobial Agents Chemotherapy*. 1994;38: 96-103. <https://doi.org/10.1128/Aac.38.1.96>
- [48]. Kuroda H, Nakazawa S, Katagiri K, Shiratori O, Kozuka M, Fujitani K, Tomita M. Antitumor Effect Of Bisbenzylisoquinoline Alkaloids. *Chemical Pharmaceutical Bulletin*. 1976;24: 2413-2420. <https://doi.org/10.1248/Cpb.24.2413>
- [49]. Wang Tx, Yang Xh. Reversal Effect Of Isotetrandrine, An Isoquinoline Alkaloid Extracted From Caulis Mahoniae, On P-Glycoprotein-Mediated Doxorubicin-Resistance In Human Breast Cancer (Mcf-7/Dox) Cells. *Yao Xue Xue Bao= Acta Pharmaceutica Sinica*. 2008;43(5): 461-466.
- [50]. Xing Z, Zhang Y, Zhang X, Yang Y, Ma Y, Pang D. Fangchinoline Induces G1 Arrest In Breast Cancer Cells Through Cell-Cycle Regulation. *Phytotherapy Research*. 2013;27(12): 1790-1794. <https://doi.org/10.1002/Ptr.4936>
- [51]. Liu Y, Xia B, Lan J, Hu S, Huang L, Chen C, Zeng X, Lou H, Lin C, Pan W. Design, Synthesis And Anticancer Evaluation Of Fangchinoline Derivatives. *Molecules*. 2017;22(11):1923. <https://doi.org/10.3390/Molecules22111923>
- [52]. Wang B, Xing Z, Wang F, Yuan X, Zhang Y. Fangchinoline Inhibits Migration And Causes Apoptosis Of Human Breast Cancer Mda-Mb-231 Cells. *Oncology Letters*. 2017;14(5): 5307-5312. <https://doi.org/10.3892/Ol.2017.6831>
- [53]. Bao K, Li Y, Wei J, Li R, Yang J, Shi J, Li B, Zhu J, Mao F, Jia R, Li J. Fangchinoline Suppresses Conjunctival Melanoma By Directly Binding Fubp2 And Inhibiting The Homologous Recombination Pathway. *Cell Death & Disease*. 2021;12(4): 380. <https://doi.org/10.1038/S41419-021-03653-4>
- [54]. Guo B, Su J, Zhang T, Wang K, Li X. Fangchinoline As A Kinase Inhibitor Targets Fak And Suppresses Fak-Mediated Signaling Pathway In A549. *Journal Of Drug Targeting*. 2014;23(3): 266-274. <https://doi.org/10.3109/1061186x.2014.992898>
- [55]. Jung Yy, Ha Ij, Um Jy, Sethi G, Ahn Ks. Fangchinoline Diminishes Stat3 Activation By Stimulating Oxidative Stress And Targeting Shp-1 Protein In Multiple Myeloma Model. *Journal Of Advanced Research*. 2022;35:245-257. <https://doi.org/10.1016/J.Jare.2021.03.008>
- [56]. Fan B, Zhang X, Ma Y, Zhang A. Fangchinoline Induces Apoptosis, Autophagy And Energetic Impairment In Bladder Cancer. *Cellular Physiology And Biochemistry*. 2017;43:1003-1011. <https://doi.org/10.1159/000481698>
- [57]. Wang N, Pan W, Zhu M, Zhang M, Hao X, Liang G, Feng Y. Fangchinoline Induces Autophagic Cell Death Via P53/Sestrin2/Ampk Signalling In Human Hepatocellular Carcinoma Cells. *British Journal Of Pharmacology*. 2011;164(2b):731-742. <https://doi.org/10.1111/J.1476-5381.2011.01349.X>
- [58]. Wang Y, Chen J, Wang L, Huang Y, Leng Y, Wang G. Fangchinoline Induces G0/G1 Arrest By Modulating The Expression Of Cdkn1a And Cnd2 In K562 Human Chronic Myelogenous Leukemia Cells. *Experimental And Therapeutic Medicine*. 2013;5(4):1105-1112. <https://doi.org/10.3892/Etm.2013.924>
- [59]. Yang J, Hu S, Wang C, Song J, Chen C, Fan Y, Ben-David Y, Pan W. Fangchinoline Derivatives Induce Cell Cycle Arrest And Apoptosis In Human Leukemia Cell Lines Via Suppression Of The Pi3k/Akt And Mapk Signaling Pathway. *European Journal Of Medicinal Chemistry*. 2020;186:111898. <https://doi.org/10.1016/J.Ejmech.2019.111898>
- [60]. Tian F, Ding D, Li D. Fangchinoline Targets Pi3k And Suppresses Pi3k/Akt Signaling Pathway In Sgc7901 Cells Retraction In/10.3892/Ijo. 2023.5547. *International Journal Of Oncology*. 2015;46(6): 2355-2363.
- [61]. Guo B, Xie P, Su J, Zhang T, Li X, Liang G. Fangchinoline Suppresses The Growth And Invasion Of Human Glioblastoma Cells By Inhibiting The Kinase Activity Of Akt And Akt-Mediated Signaling Cascades. *Tumor Biology*. 2016;37(2):2709-2719. <https://doi.org/10.1007/S13277-015-3990-1>
- [62]. Lee Hs, Safe S, Lee So. Inactivation Of The Orphan Nuclear Receptor Nr4a1 Contributes To Apoptosis Induction By Fangchinoline In Pancreatic Cancer Cells. *Toxicology And Applied Pharmacology*. 2017;332:32-39. <https://doi.org/10.1016/J.Taap.2017.07.017>
- [63]. Wang C-D, Huang J-G, Gao X, Li Y, Zhou S-Y, Xu Yan X, Zou A, Chang J-L, Wang Y-S, Yang G-X, He G-Y. Fangchinoline Induced G1/S Arrest By Modulating Expression Of P27, PcnA, And Cyclin D In Human Prostate Carcinoma Cancer Pc3 Cells And Tumor Xenograft, Bioscience, Biotechnology, And Biochemistry. 2010;74(3):488-493. <https://doi.org/10.1271/Bbb.90490>
- [64]. Valsan A, Omanakuttan Vk, Radhakrishnan Kv, Maiti Kk. A Comprehensive Appraisal Of Bisbenzylisoquinoline Alkaloids Isolated From Genus *Cyclea* For Anticancer Potential. *Journal Of Biochemical And Molecular Toxicology*. 2025;39(2):P.E70137. <https://doi.org/10.1002/Jbt.70137>
- [65]. He Bc, Gao Jl, Zhang Bq, Luo Q, Shi Q, Kim Sh, Huang E, Gao Y, Yang K, Wagner Er, Wang L. Tetrandrine Inhibits Wnt/B-Catenin Signaling And Suppresses Tumor Growth Of Human Colorectal Cancer. *Molecular Pharmacology*. 2011;79(2):211-219. <https://doi.org/10.1124/Mol.110.068668>
- [66]. Jang Bc, Lim KJ, Paik Jh, Cho Jw, Baek Wk, Suh Mh, Park Jb, Kwon Tk, Park Jw, Kim Sp, Shin Dh. Tetrandrine-Induced Apoptosis Is Mediated By Activation Of Caspases And Pkc-Δ In U937 Cells. *Biochemical Pharmacology*. 2004;67(10):1819-1829. <https://doi.org/10.1016/J.Bcp.2004.01.018>
- [67]. Meng Lh, Zhang H, Hayward L, Takemura H, Shao Rg, Pommier Y. Tetrandrine Induces Early G1 Arrest In Human Colon Carcinoma Cells By Down-Regulating The Activity And Inducing The Degradation Of G1-S-Specific Cyclin-Dependent Kinases And By Inducing P53 And P21cip1. *Cancer Research*. 2004;64(24):9086-9092. <https://doi.org/10.1158/0008-5472.Can-04-0313>
- [68]. Ng Lt, Chiang Lc, Lin Yt, Lin Cc. Antiproliferative And Apoptotic Effects Of Tetrandrine On Different Human Hepatoma Cell Lines. *The American Journal Of Chinese Medicine*. 2006;34(01):125-135. <https://doi.org/10.1142/S0192415x06003692>
- [69]. Qiu W, Su M, Xie F, Ai J, Ren Y, Zhang J, Guan R, He W, Gong Y, Guo Y. Tetrandrine Blocks Autophagic Flux And Induces Apoptosis Via Energetic Impairment In Cancer Cells. *Cell Death & Disease*. 2014;5(3): E1123-E1123.
- [70]. Wan J, Liu T, Mei L, Li J, Gong K, Yu C, Li W. Synergistic Antitumor Activity Of Sorafenib In Combination With Tetrandrine Is Mediated By Reactive Oxygen Species (Ros)/Akt Signaling. *British Journal Of Cancer*. 2013;109(2):342-350.
- [71]. Liu W, Zhang J, Ying C, Wang Q, Yan C, Jingyue Y, Zhaocai Y, Yan X, Heng-Jun S, Lin J. Tetrandrine Combined With Gemcitabine And Cisplatin For Patients With Advanced Non-Small Cell Lung Cancer Improve Efficacy. *International Journal Of Biomedical Science: Ijbs*. 2012;8(1): 28-35.

- [72]. Xiao-Hui C, Shuai W, Bao-An C. Research Advances On The Pharmacological Effects Of Tetrandrine. Chinese Journal Of Natural Medicines. 2011;9(6): 473-480. <https://doi.org/10.3724/Sp.J.1009.2011.00473>
- [73]. Liu C, Xiao P. Biological Activities And Domestic Resources Of Bisbenzylisoquinoline Alkaloids. Yaoxue Tongbao. 1983;18: 287-292.
- [74]. Uche F, Li Ww, Richardson A, Greenhough Tj. Anti-Ovarian Cancer Activities Of Alkaloids From Triclisia Subcordata Oliv. (Menispermaceae). Planta Medica. 2014;80 - Pf7. Doi: 10.1055/S-0034-1382585
- [75]. Otshudi, A.L., Apers, S., Pieters, L., Claeys, M., Pannecouque, C., De Clercq, E., Van Zeebroeck, A., Lauwers, S., Frederich, M., & Foriers, A. (2005). Biologically Active Bisbenzylisoquinoline Alkaloids From The Root Bark Of Epinetrum Villosum. Journal Of Ethnopharmacology. 2005;102: 89-94. <https://doi.org/10.1016/J.Jep.2005.05.021>
- [76]. Bala M, Kumar S, Pratap K, Verma Kp, Padwad Y, Singh B. Bioactive Isoquinoline Alkaloids From Cissampelos Pareira. Natural Products Research. 2019;33(5):622-627. Doi: 10.1080/14786419.2017.1402319
- [77]. Nawawi A, Ma Cm, Nakamura N, Hattori M, Kurokawa M, Shiraki K, Kashiwaba N, Ono M. Anti-Herpes Simplex Virus Activity Of Alkaloids Isolated From Stephania Cepharantha. Biological And Pharmaceutical Bulletin. 1999;22(3): 268-274. <https://doi.org/10.1248/Bpb.22.268>