

Micro Analysis of Medicinal Compounds using Ammonium Metavanadate Reagent: Study of Alcohols and Sugars

Kumari Anita Sinha^{1,*}

¹(PG Department of Chemistry, Magadh University, Bodh Gaya, Bihar 824234)
(At-Janakpur, PO-Buniyadganj, Gaya, Bihar)

Abstract: The current study depicts that the determination of butane 1,3 diol, butane 2,3 diol, triethylene glycol, methanol, ethanol, n-propanol, n-butanol, 3-pentanol, propane 1,2 diol, and tetraethylene glycol with the use of V(v) reagent. The study also shows that the oxidizing capacity of ammonium metavanadate (v) with certain organic compounds, the oxidation of certain sugars was studied using an accurate and quantitative procedure has been described for the determination of certain mono and disaccharides on micro scale.

Background: Drugs are substances or products with a definite physiological activity that are used or intended to be used to modify or explore physiological systems or pathological states for the benefit of the recipient. It is used in the prevention, diagnosis, treatment or cure of diseases in man or other animals. The determination of these biologically active functional groups is important for the quantitative determination of drugs. Physiological activity of a compound is associated with particular structural unit or particular functional groups. These functional groups are biologically active.

Materials and Methods: Aliquots containing 1 to 5 mg of glucose were taken in flask and 1 ml of 0.3 N, V(v) reagent and 5 ml of 10 N sulphuric acid was added to it. The reaction was carried out on a boiling water bath for 5, 10, 15, 20, 25, 30, 35, 40, 45 minutes. After the prescribed reaction time contents were cooled at room temperature and the unconsumed V(v) reagent was determined. The unconsumed V(v) reagent was titrated against 0.025 N ferrous I ammonium sulphate using N-phenyl anthranilic acid as an indicator. A blank experiment was also run under identical condition, using all the reagents except the sample. The recovery of the sample was calculated.

Results: In the view of the reactions discussed both sugars and alcohols were studied using the reagent. It has been found that sugars containing aldehydic group are oxidised to formic acid only. In case of Ketonic sugars also formic acid is the end product but ketonic group appear to be oxidized to carbon dioxide. Polymers of Ethylene glycol like triethylene glycol and tetra ethylene glycol consumes 8 equivalent of V(v) reagent to give corresponding dicarboxylic acids.

Conclusion: The current study depicts that the determination of butane 1,3 diol, butane 2,3 diol, triethylene glycol, methanol, ethanol, n-propanol, n-butanol, 3-pentanol, propane 1,2 diol, and tetraethylene glycol with the use of V(v) reagent. The study also shows that the oxidizing capacity of ammonium metavanadate (v) with certain organic compounds, the oxidation of certain sugars was studied using an accurate and quantitative procedure has been described for the determination of certain mono and disaccharides on micro scale.

Key Word: Medicinal Compounds, Alcohols, Sugars, Ammonium Metavanadate Reagent.

Date of Submission: 31-10-2020

Date of Acceptance: 12-11-2020

I. Introduction

Drugs are substances or products with a definite physiological activity that are used or intended to be used to modify or explore physiological systems or pathological states for the benefit of the recipient. It is used in the prevention, diagnosis, treatment or cure of diseases in man or other animals. The determination of these biologically active functional groups is important for the quantitative determination of drugs. Physiological activity of a compound is associated with particular structural unit or particular functional groups. These functional groups are biologically active.

Lindberg and Missionry^{1,2} employed sodium borohydride as the reductant for determining carbohydrates. The sample was dissolved in water and treated with solution of sodium borohydride. After the reaction is completed the residual borohydride is determined by measuring the hydrogen evolved upon the addition of acid. Peat, Whelan and Roberts' determined the degree of polymerization of reducing oligosaccharides, by means of sodium borohydride.

Various other titrimetric methods²³⁻²⁵ have been proposed from time to time for the determination of alcohols. Dusic²⁶ determined diols by using NaO₄ solution as oxidant excess 10, was determined potentiometrically with N₂H₄H₂S₀, at platinum electrode. Singh et al oxidised methanol and glycerol with

excess amount of permanganate in alkaline solution. The excess reagent was titrated potentiometrically with sodium formate in presence of BaCl₂. Beck² determined ethylene glycol and glycerol with trivalent copper by potentiometric titration. Stognushko et al developed a potentiometric method for the analysis of phosphorylated product at higher alcohols. A suitable method has been suggested for the determination of Crotyl alcohol with chloramine-T by Naidu et al Haring et al³² evolved a method for the determination of allyl alcohol with manganese pyrophosphatic. Various metal ions³³⁻³⁴ were also employed for the determination of alcohols. Oxidation of secondary alcohol as well as allyl alcohol³⁶ with PFC and Ni(iv) respectively are carried out. Various Methods developed for the determination of alcohols, with different inorganic reagents. In the present work a new titrimetric method has been described for the micro determination of some monohydric and dihydric alcohols with the use of ammonium metavanadate as an oxidising reagent. This is quick, easy and accurate procedure and is of general applicability. The accuracy of the method is within +1% is most of the cases.

Komers et al proposed a method for the determination of small amount of MeOH and EtOH in aqueous solution. The method based on combination of stripping and gas chromatography technique. Normal C₇-C₁₆ aliphatic alcohols mixture was analysed by gas Chromatography' at 190° on a column packed with 10% polyethyleneglycol. Other chromatographic techniques like liquid chromatography and gas chromatography were also employed for the determination of monohydric and dihydric alcohols. Glycerol was determined spectrophotometrically in its industrial liquors by measuring the absorbance at 580 nm of the violet colour several other spectrophotometric method' are also available in the literature for the determination of alcohols. Barkat evolved a titrimetric method using N-bromosuccinimide as oxidant for the determination of primary and secondary alcohols. Kruse et al developed an oxidimetric method for differentiating between three types of saturated alcohols using Nhaloimides. A similar type of method has also been proposed by Severin. It has been claimed that the method is rapid and accurate compared to the methods proposed by Lucas and Ritter. Litter as well as Jones studied the reaction. Kinetics of oxidation of alcohols using pentavalent vanadium as oxidant and excess vanadate was back titrated with Mohr's salt or with Fe (II) sulphate solution.

Colorimetric methods are also available for the determination of sugars. The first procedures were developed by Floin and Wu⁴ in 1919 which is based in the conversion of cupric ions to the cuprous sulphate by the presence of reducing sugars. The cuprous ions subsequently reduce phosphotungstic acid to a blue complex which is measured calorimetrically. Dearing⁵ described a micro method for the estimation of cellulose Wahba and Coworkers⁶ converted glucose to glucosazone and measured the yellow colour of its solution. Shallenberger and Mooros⁷ developed the colour with a reagent containing copper sulphate and arsenomolybdates and measured the solution at 500mg Various phenolic compounds have been recommended for the colorimetric determination of carbohydrates. Dubois⁸ used phenol for the determination of reducing sugars. Tillmans⁹ determined the reducing sugars with the use of thymol. Sorenson, Fisher and Lindh 10-12 used resorcinol and phloroglucinol for the determination of reducing sugars.

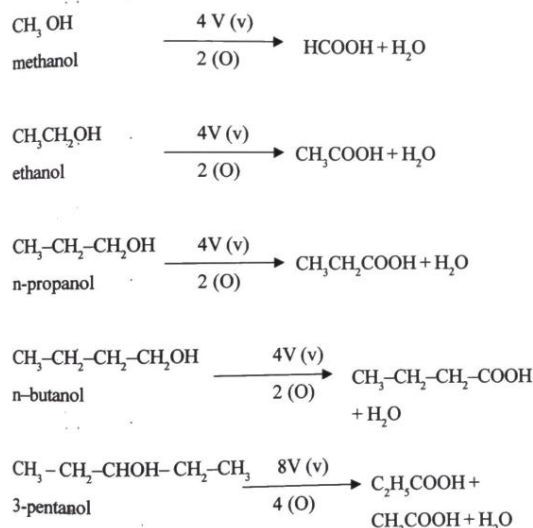
Rebega³⁸ and Rlbega and coworkers³⁹ used cuprithiosalicylate as oxidising agent and have determined sucrose, fructose and glucose empirically. Celsi and SarraillW° oxidised glucose, sucrose and lactose with Cu(14) in the mixture of K₂CO₃ and KCNS. The precipitated CuSCN was estimated using Fe(III) and Ag(I). Defrates and castles estimated reducing sugars with Fehling's solution using electrometric end point detector. Potassium ferricyanide has been widely used in determining sugar^{42, 48}. According to Blom and Rosted⁴⁹ this method of estimation so sugars with ferricyanide is much inferior to cuprimetry method in as much as ferricyanide procedures, side reactions and it attacks other organic substances present in the sugar. Lately there have been good deal of attempts to regularise the oxidation of sugars with ferricyanide better results have been claimed when sugar is treated against a boiling alkaline Solution of hexacyanoferrate (111)⁵⁰.

Hass and Lynch²² developed a method for the determination of carbohydrates. Mayer and Isbe¹²³ employed radioactivity and determined end groups in carbohydrates. Browne²⁴ determined carbohydrates by biological methods. Figueiredo²⁵ have developed an improved complexometric method for the analysis of reducing sugars. Amongst the various oxidimetric methods for the determination of sugars, cuprimetry has been of great use from early times. Divalent copper complexed with tartarate or citrate in alkaline medium known as Fehling's solution²⁶⁻²⁹ and Benedict's solution respectively.

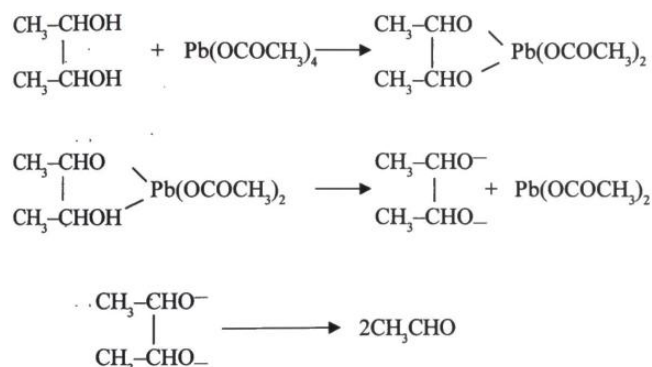
II. Material and Methods

The unconsumed V(v) reagent was titrated against 0.025 N ferrous I ammonium sulphate using N-phenyl anthranilic acid as an indicator. A blank experiment was also run under identical condition, using all the reagents except the sample. Recovery of the sample was calculated by following expression.

$$\text{mg of the sample} = \frac{M \times N (B - S)}{n}$$



In the above oxidation reaction primary alcohols after oxidation with V (v) gives corresponding carboxylic acid and secondary alcohol produces a mixture of acids. Aldehydes and Ketones are produced in case of 1, 2-glycol as well with certain reagents, notably periodic acid and lead tetraacetate. The reaction affords one of the few ways in which a carbon - carbon single bond can be cleaved readily various theory have been proposed to explain the cleavage, one of which postulates that the glycol is changed to 1, 4-Zwitter ions, which is turn breaks down to form two carboxyl groups. According to this theory the cleavage of 2,3 butane diol with lead tetraacetate⁴⁸ may be depicted as follows.



IV. Conclusion

The current study depicts that the determination of butane 1,3 diol, butane 2,3 diol, triethylene glycol, methanol, ethanol, n-propanol, n-butanol, 3-pentanol, propane 1,2 diol, and tetraethylene glycol with the use of V(v) reagent. The study also shows that the oxidizing capacity of ammonium metavanadate (v) with certain organic compounds, the oxidation of certain sugars was studied using an accurate and quantitative procedure has been described for the determination of certain mono and disaccharides on micro scale.

References

- [1]. Leenson, J:A. Sergeev G.B. et al, Z. Anal. Khim; 32 (5) (Russ) (1977).
- [2]. Barakat, M. S. and M. F. A., El-Wahab, J. Amer. Chem. Soc., 75, 573 (1953).
- [3]. Reynold, C. Fuson, J. Org. Chem; 21,478(1956)
- [4]. Rajni Mohanty K; Das, M; and Das, A., Indian J. Chem. Sec A. 37, 34 (1998).
- [5]. Naidu H. M. K. et al, Indian J.Chem. 13(a) 976-7 D. S. (Eup) (1975).
- [6]. Lotan, Noah, Ehrlich et al Microchem. J. 20(4) 534-8, (1975) Eng.
- [7]. Dusic, Z., Inst. Anal. Chem. Fac. Pharm. Arh Farm. 25 (4), 233-40 (1975)
- [8]. Singh, B. Singh A. et al, Indian Chem. Soc; 33, 778 (1956)
- [9]. Beck, G, Mikrochemic, 38, 152 (1951)
- [10]. Beck, G., Mikrochemic, 40, 258 (1953)lindberg. It. & Missionry, A. Svcsnk, Papperstidn; 55, 13 (1952)
- [11]. lindberg, 13. & Theander, Ibd; 57, 83 (1954)
- [12]. Peat, S.; Whelan, W. J. & J. Chem. Soc., 2258 (1956)
- [13]. Floin, O. & Wu, M. J. Biochem., 38, 110 (1919)
- [14]. Dearing, G. G. Nautre; 179, 579 (1957)
- [15]. Wahba, N.;Hanna, S. Analyst, 81, 27 (1957).

- [16]. Singh, R., D. Phil. Thesis A. U. (1979).
- [17]. Paparia, Migue, *Quint Anal*; 30 (2), 93 (1976)9
- [18]. Bondak, M.I., Lyutvort, S.G., Malikov, V.T., Pyatin, S., Skidan, Yu.A. (Vennits Polybelinic Institute) USSR 544, 917
- [19]. Churasek, J., *J. Chromatogr. Libr*; 3 (1975)
- [20]. El-Aggar, A.M. et al. *J. Chem.* 19 (41, 499-503) (1970) (Eng.)
- [21]. Srivanasan, M. et al, *Z. Anal. Chem*; 282(2), 143 (1976) (Eng.)
- [22]. Kawati, Shohai et al. *J. Technol.Rep. Kansai Univ.*18, 47-52 (1972) (Eng.)
- [23]. Stognushko, D. P. et al, *Z. Anal. Khim*; 33(1) 142-5 (1978) (Russ)
- [24]. Naidu, H. M. K. and Mahadevappa, D. S., *Curr. Sci*; 45(6) 216-18 (1976) (Japan)
- [25]. Land, Haring and William, A. W., *J. Chem. Soci*; 35, 111 (1960)
- [26]. Kartha, K.P. R., Aloui, M. and Robert, A. F., *Tetrahedron*; 37, 8807 (1996).Ritter, F. et al, *J. Chem. Educ*; 30, 395 (1953)
- [27]. Litter, J. S. and Watters, W.A., *J. Chem. Soc*; 1299, 301, 4046 (1959).
- [28]. Jones, J. R. and Wafters, W. A., *J. Chem. Soc*; 1629, 2068 (1962).
- [29]. Srivastava, S and Gupta, V., *J. Indian Chem. Soc*; 83, 1103 (2006).
- [30]. Swann, S. and Xanthakos T. S, *J. Amer. Chem. Soc.* 53 (1928)
- [31]. Drummond, A. Y. and Waters, W. A., *J. Chem. Soc*; 3119, 3456 (1954), 217 (1955), 4312 (1957) 2129(1958)
- [32]. Rama Krishnan, P. S.and Chockalingam, *J. Indian Chem. Soc*; 70, 583 (1993).
- [33]. Tuwar,S. M., bewoor, S. T. Raju, J. R., Nandi 69, 651 (1992)
- [34]. Dutta, S Karki, R. Strivanda, P.G. and Udupa, N. I, *J. Pharma Sci*; 62, 381 (2000)
- [35]. Kande, R. B., Kasture, A.V., and Wakodher, A., *Ind. J. Pharm. Sci*;64, 24 (2002)
- [36]. Kumar, H.M.S, Anjaneyulu et al, *J. Indian Chem. Soc*; 84, 189 (2007).
- [37]. Iloukhani, H; Zarie F., *Asian J. Chem.* 17(4) 2437 (2005).
- [38]. Niknam, Khodabakhsh, *Asian J. Chem.* 17(4) 2513 (2005).
- [39]. SerVi, S. and Acar, A., *Molecules*; 7 104 (2002) Reaction of Organic Compounds 212 (1962)
- [40]. Reynold, C. Fuson, *Reaction of Organic Compounds*; 227 (1962)
- [41]. Severin, M. et al, *Bull, Rech Argon. Cemblaix*, 1(2), 289; C.A. 66, 176188 (1966)

Kumari Anita Sinha. "Micro Analysis of Medicinal Compounds using Ammonium Metavanadate Reagent: Study of Alcohols and Sugars." *IOSR Journal of Applied Chemistry (IOSR-JAC)*, 13(11), (2020): pp 58-62.