

Antilipidaemic Drugs

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Abstract

Everyone has some degree of coronary atherosclerosis. CHD is most common in affluent nations. Diets high in fats, elevated serum cholesterol levels, lack of exercise, emotional stress, hypertension and smoking all contribute to the production of this disease.

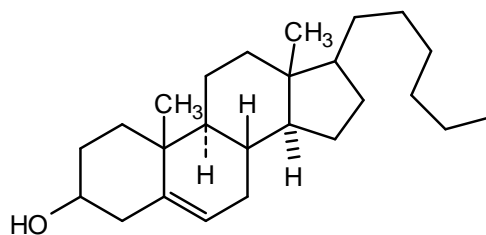
Keywords: CHD, Atherosclerosis, Hypertension

I. INTRODUCTION

Coronary Heart Disease (CHD) is the leading cause of death in the United States. More than 650,000 people die each year from CHD in this country and about 20% of the population will develop the first symptoms before age 60^{1,2}. Although nearly everyone has some degree of coronary atherosclerosis the underlying cause of CHD is most common in affluent nations. Diets high in fats, elevated serum cholesterol levels, lack of exercise, emotional stress hypertension and smoking all contribute to the production of this disease³⁻⁹.

DRUGS OF HISTORICAL INTEREST

During the mid-1900s, considerable interest arose in plant sterols as serum cholesterol-lowering agents. Mixtures of plant sterols, including, soybean sterols and β -sitosterol (22, 23- dihydrastigmasterol) (1) are among the preparations that are absorbed to a limited extent from the intestinal tract and compete for the absorption of both endogenous and exogenous cholesterol^{10,11}.

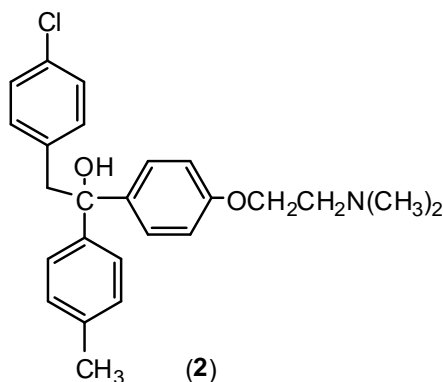


β -SITOSTEROL (1)

In animals (1) also increases the rate of cholesterol and lipid metabolism¹².

TRIPARANOL (2)

Triparanol (2) was a potent inhibitor of cholesterol biosynthesis^{13,14}.

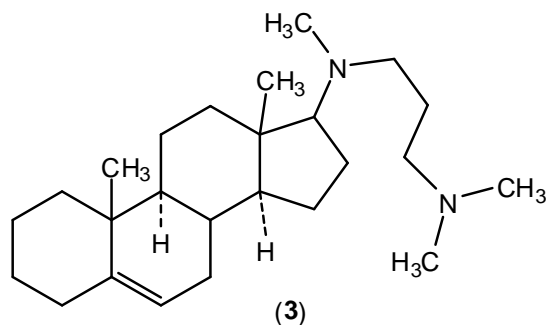


(2)

DIAZACHOLESTEROLS

It was originally suggested that diazacholesterols and related analogs, which look sufficiently like cholesterol, would bind to negative feedback enzymes because of electron rich nitrogen. Such binding would mimic cholesterol binding and inhibit cholesterol biosynthesis. Compounds such as 20, 25-diazacholesterol (3)

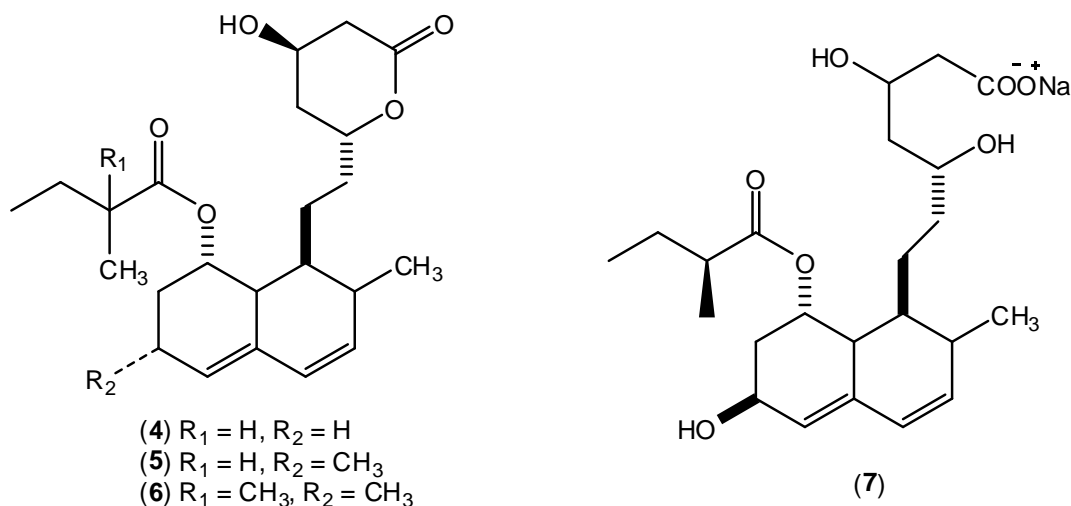
however, generally inhibit reduction of 7, 8- and 24, 25- double bonds¹⁵ and *in vivo*, HMG-CoA reductase activity is stimulated even though serum cholesterol levels are reduced¹⁶.



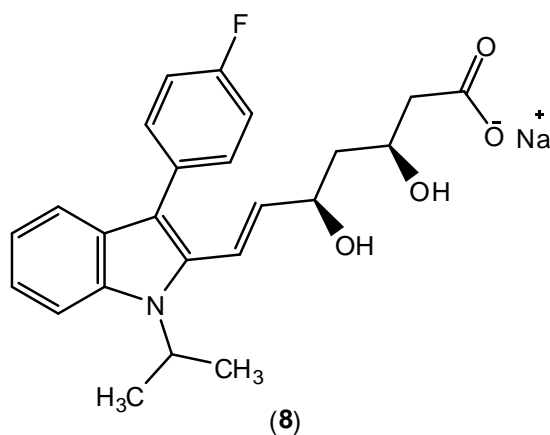
DRUGS OF CLINICAL SIGNIFICANCE

Statins (or vastins) represents a new class of lipid-lowering compounds that inhibit HMG-CoA reductase, the first and rate-limiting step in cholesterol biosynthesis in cells^{17, 18}.

Mevastatin (4) and lovastatin (5) were isolated as fungal metabolites from cultures of *Penicillium brevicompactum* and *Aspergillus terreus* respectively^{19, 20}. Other closely related analogs include pravastatin (6) and simvastatin (7).



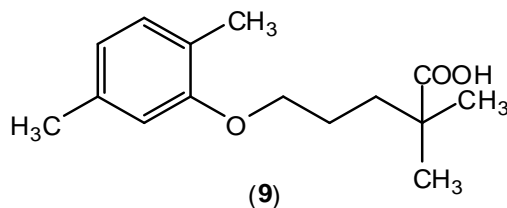
Fluvastatin sodium (8) is the most recently approved statin in the United States²¹.



It is a white pale yellow, hygroscopic, powder that is freely soluble in water ethanol, and methanol. It is supplied as 20mg and 40mg capsules for oral administration.

FIBRATES**GEMFIBROZIL (9)**

It was introduced in 1981 after the statins, and is the second most frequently presented antilipidaemic drugs. This drug is a white solid that is nearly insoluble in water and acidic solutions, and soluble at greater than 1% in dilute base²².

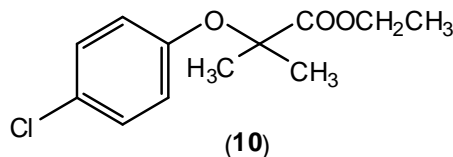


Overall gemfibrozil (9) effectively lowers plasma triglycerides, VLDL cholesterol and VLDL ApoA-I concentrations in **type II a** and **II b** hyperlipoproteinemic patients and produces a greater reduction in LDL cholesterol levels in type II a patients^{23, 24}.

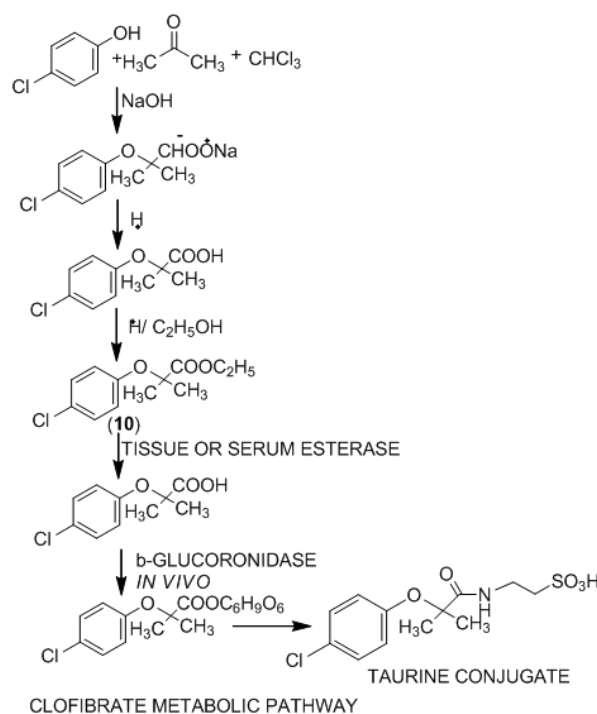
Gemfibrozil (9) is rapidly and well absorbed with a peak plasma level occurring 1 to 2 hours after dosing²⁵. It is extensively metabolized in humans and experimental animals²⁶.

CLOFIBRATE (10)

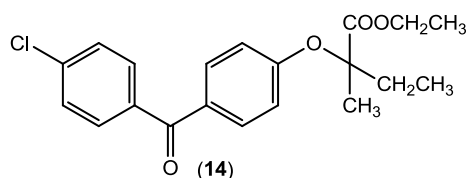
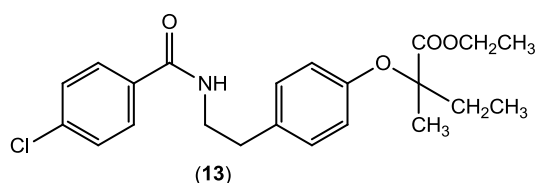
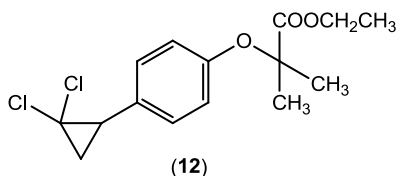
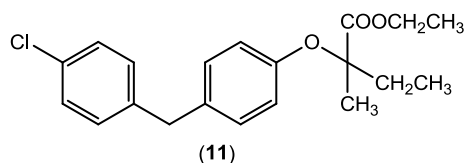
It was the first fibrate introduced in the United States. It is nearly colourless, oily liquid, insoluble in water and miscible with organic solvents²⁷.



Serum cholesterol lowering by clofibrate (10) in animals is related to changes in liver cholesterol through blockade of HMGCoA reductase²⁸. Serum cholesterol catabolism remains unchanged in human after clofibrate treatment²⁹, however, acidic and neutral sterol excretion into bile is increased by its treatment³⁰.



Other fibrates which have got clinical applications are beclobrate (**11**), bezafibrate (**12**), Ciprofibrate (**13**) and fenofibrate (**14**).



II. CONCLUSION:

ANTIATHEROSCLEROSIS DRUGS IN DEVELOPMENT

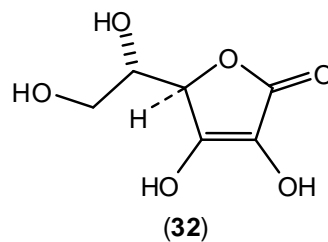
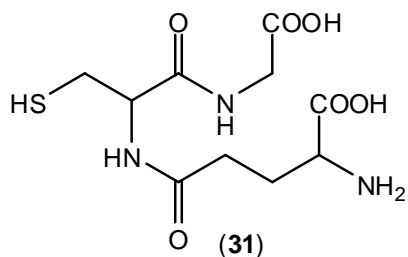
Atherosclerotic fatty streaks occur through cholesterol ester engorgement by cells. This takes place after uptake of LDL_{ox} or modified LDL. This lesion proceeds to a fibrous plaque and then to a complicated thrombus-involved atheroma which may block arterial blood flow and produce clinically manifested CHD^{5zyy}.

LDL_{ox} identified in both human and atherosclerotic animal model arteries and in the bloodstream results from a free-radical reaction sequence with LDL lipids⁵⁹. This produces endothelial cytotoxicity modified platelet functionality, increased monocyte adherence to and chemotaxis through the endothelium. This results in accumulation of arterial foam cells and enhanced smooth muscle proliferation⁶⁰.

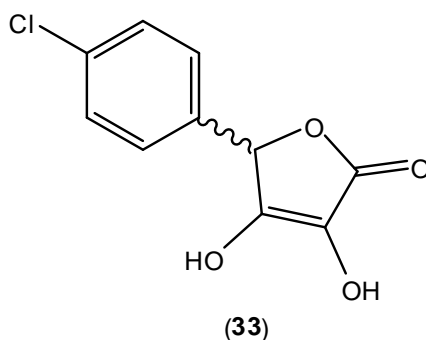
Superoxide anion radicals (O₂⁻) are continuously generated in normal cells by reduction of dioxygen (O₂). Defense mechanisms against O₂⁻ include radical scavengers as such ascorbate and vitamin E and enzymes, superoxide dismutase (SOD), and catalase, among others. O₂⁻ is the anionic form of HOD; having a pK_a = 4.8. Both O₂⁻ and HOO⁻ are reactive species but are not as reactive as their metabolites O₂ upon dismutation catalyzed by SOD yields H₂O and H₂O₂. Preferably, H₂O₂ converts to H₂O and O₂ through the action of catalase.

When normally protective mechanisms cannot handle accumulation of H₂O₂, a Fenton's reaction with ferrous ion (Fe²⁺) may cause further reduction to produce both hydroxyl radical (OH[·]) and hydroxy anion (OH⁻). H₂O₂ and especially HO[·] are highly toxic substances with macromolecules such as DNA, enzymes, proteins and lipoproteins (i.e., LDL). Such free radical reactions may result in pathologic conditions such as CHD, cancer, liver damage, rheumatoid arthritis, immunologic incompetence, among others.

Lipid peroxidation involves highly reactive radicals. Thus, radicals (R[·]) generated under conditions undergo reaction with lipid side-chains (LH) to generate new radicals (L[·]). When L[·] is an unsaturated chain, reaction with O₂ produces allylic peroxides (RCH = CH-CH₂OOH) and epoxides (oxiranes, RCH-CH) which are also reactive species. For example, peroxides may react with LH to generate L[·], which produces more LOO[·] and LOOH. Additionally, O₂ may react with LH by an insertion mechanism, lead to H₂O₂ and HO[·]; LOOH may undergo reduction by Fe²⁺ or oxidation by ferric ion (Fe³⁺) in reinitiation reactions, in which LO[·] and LOO[·], respectively, are product. Glutathione (**31**) and other reducing agents such as ascorbic acid (**32**) may remove LOOH through generation of LOH and H₂O. Unfortunately, ascorbate can also reduce Fe³⁺ generating Fe²⁺, which serves to increase concentrations of HO[·] in Fenton's reaction.



4- (4- Chlorophenyl) - 2- hydroxy-tetronic acid (CHTA) (33) is very effective hypolipidemic drug *in vivo* and an antioxidant *in vitro*³⁴.



In the future, such experimental antiatherosclerotic drugs may more effectively fulfill the antioxidant ascorbate function of protecting endogenous LDL antioxidants.

REFERENCES

- [1]. Wittels, E. H., *Am. J. Cardiol.*, **65**, 432 (1990)
- [2]. Goldman, L., *Am. Heart J.*, **119**, 733 (1990)
- [3]. Frank, C. W., *Bull. N. Y. Acad. Med.*, **44**, 900 (1968)
- [4]. Epstein, F. H., *Bull. N. Y. Acad. Med.*, **44**, 916 (1968)
- [5]. Rinzlör, S. H., *Bull. N. Y. Acad. Med.*, **44**, 936 (1968)
- [6]. Fox, S. M. and Haskell, W. L., *Bull. N. Y. Acad. Med.*, **44**, 950 (1968)
- [7]. Deming, Q. B., *Bull. N. Y. Acad. Med.*, **44**, 968 (1968)
- [8]. Stamler, J., *Bull. N. Y. Acad. Med.*, **44**, 1476 (1968)
- [9]. Garfinkel, L., *Bull. N. Y. Acad. Med.*, **44**, 1495 (1968)
- [10]. Pollak, O. J., *Circulation*, **7**, 702 (1953)
- [11]. Hernandez, H. H., *Proc. Soc. Exp. Biol. Med.*, **83**, 498 (1953)
- [12]. Gerson, T. and Shorland, F. B., *Nature*, **200**, 579 (1963)
- [13]. Blohm, T. R. and Mackenzie, R. D., *Arch. Biochem. Biophys.*, **85**, 245 (1959)
- [14]. Blohm, T. R., *Arch. Biochem. Biophys.*, **85**, 250 (1959)
- [15]. Smith, M. M., *Rep. Prog. Appl. Chem.*, **52**, 146 (1967)
- [16]. Langdon, R., *J. Lipid Res.*, **18**, 24 (1977)
- [17]. Alberts, A. W., *Proc. Natl. Acad. Sci.*, **77**, 3957 (1980)
- [18]. Rodbell, V. W., *Adv. Lipid Res.*, **14**, 1 (1976)
- [19]. Endo, A., *J. Antibiot.*, **29**, 1346 (1976)
- [20]. Brown, A. G., *J. Chem. Soc. Perkin Trans.*, **1**, 1165 (1976)
- [21]. Foye, W. O., Lemke, T. L. and Williams, D. A., *Medicinal Chemistry*, IVth ed. p. 509 (1995)
- [22]. Pento, J. T., *Drugs of Today*, XVIII, 585 (1982)
- [23]. Levy, R. I., in "Proc. 2nd World Conference Chemical Pharmacology, Lemberger, L. and Rvedenberg, M. M., Eds. Bethesda, MD, ASPET, p. 916 (1984)
- [24]. Lupien, P. J., *Can. J. Cardiol.*, **7**, 27 (1991)
- [25]. Smith, T. C., *Proc. R. Soc. Med. (Suppl.2)*, **69**, 24 (1976)
- [26]. Okerholm, R. A., *Fed. Proc.*, **35**, 327 (1976)
- [27]. Merck Index, 10th ed. Rahway, NJ. Merck, 1983
- [28]. Cohen, B. I., *Biochem. Biophys. Acta.*, **369**, 79 (1974)
- [29]. Grundy, S. M., *J. Lipid Res.*, **13**, 531 (1972)
- [30]. Schwartz, C. J., *Clin. Cardiol.*, **14**, 11 (1991)
- [31]. Kinsky, N. I., *Proc. Soc. Exp. Biol. Med.*, **200**, 248 (1992)
- [32]. Esterbauer, H., *Chem. Res. Tox.*, **3**, 77 (1990)
- [33]. Ondrias, K., *Free Radic. Res. Commun.*, **16**, 227 (1992)
- [34]. Nestel, P. J., *J. Clin. Invest.*, **44**, 891 (1963)